



A COMPREHENSIVE REVIEW ON PHYTOCHEMICALS WITH ANTI-DIABETIC ACTIVITY: MECHANISMS AND APPLICATIONS

S. Velmurugan¹, S. Seshai¹, V.J. Ramachandiran¹, A. Reshma¹, S. Anbazhagan²,
S. A. Vadivel^{3*}

¹Research student, Surya School of Pharmacy, Villupuram.

²Professor, Department of Pharmaceutical Chemistry, Surya School of Pharmacy, Villupuram.

^{3*}Associate Professor, Department of Pharmaceutics, Surya School of Pharmacy, Villupuram.

***Corresponding Author:** S. A. Vadivel

*E-mail: vetrivadivel24@gmail.com

ABSTRACT

Diabetes mellitus is a chronic metabolic disorder characterized by impaired glucose homeostasis, insulin resistance, and long-term complications affecting multiple organ systems. The search for effective and safer alternatives to conventional antidiabetic drugs has led to growing interest in phytochemicals derived from medicinal plants. Phytochemicals such as flavonoids, alkaloids, phenolics, terpenoids, and glycosides have demonstrated significant antidiabetic potential through diverse mechanisms, including inhibition of carbohydrate-digesting enzymes, enhancement of insulin secretion, and modulation of glucose uptake and antioxidant activity. This review comprehensively explores the diverse classes of phytochemicals with antidiabetic properties, highlighting their mechanisms of action, pharmacological potential and clinical relevance. Further, the application of these bioactive compounds in functional foods, nutraceuticals and pharmaceutical formulations is discussed, along with challenges such as bioavailability, toxicity, and regulatory considerations. The integration of phytochemicals in diabetes management holds promise for the development of novel, plant-based therapeutics, paving the way for improved treatment strategies.

Keywords: Phytochemicals, Diabetes mellitus, Insulin resistance, Antioxidants, Natural products, Drug discovery

INTRODUCTION

Diabetes mellitus (DM) is a chronic disease caused by inherited and/or acquired deficiency in production of insulin by the pancreas, or by the ineffectiveness of the insulin produced. Such a deficiency results in increased concentrations of glucose in the blood, which in turn damage many of the body's systems, in particular the blood vessels and nerves. As the numbers of people with diabetes multiply worldwide, the disease takes an ever-increasing proportion of national and international health care budgets. It is projected to become one of the world's main disablers and killers within the next 25 years. Regions with greatest potential are Asia and Africa, where DM rates could rise to two- to three-folds than the present rates. Apart from currently available therapeutic options, many herbal medicines have been recommended for the treatment of diabetes. Traditional plant medicines are used throughout the world for a range of diabetic presentations [1]. The prevalence of diabetes in the world was estimated to be 2.8% for all ages in 2000, and that is

expected to increase to approximately 4.4% in 2030. Diabetes-related deaths will increase to 366 million by 2030. The World Health Organization (WHO) estimates that 415 million people will be affected by diabetes in 2015, and this is expected to rise to 642 million by 2040, worldwide. Currently, the numbers of diabetic patients have significantly increased in the population between 45 and 64 years of age in many countries, particularly in China, India, and Southeast Asia. Of various DM subtypes, T2DM, which is characterized by chronic metabolic imbalance, beta-cell failure and insulin resistance, and can be alleviated by changing lifestyle by dietary control and exercise, is the most common type, accounting for more than 90% of all DM patients. The onset of T2DM can be attributed to behavioural, environmental, and genetic factors, leading to insulin resistance and deficiency. Importantly, the involvements of several factors in T2DM that cause resistance of target tissues to insulin, usually resulting from abnormal insulin secretion. T2DM is a common and increasingly prevalent disease and is a major public health problem worldwide [2]. Importantly, prevalence surveys indicate that, in 2017, there are 425 million people with DM in the world. In 2045, estimations suggest that this number will reach 629 million people. Despite the high economic cost for DM treatment (\$ 727 million dollars in 2017), there is still a large part of the population without access to pharmacotherapy, or even with the availability of drug treatment, seeking treatment options in traditional medicine. In fact, a variety of plants with possible antidiabetic properties is used in traditional medicine. Approximately 800 plants have been identified with antidiabetic effect, and more than 200 bioactive compounds have been identified with this potential [3].

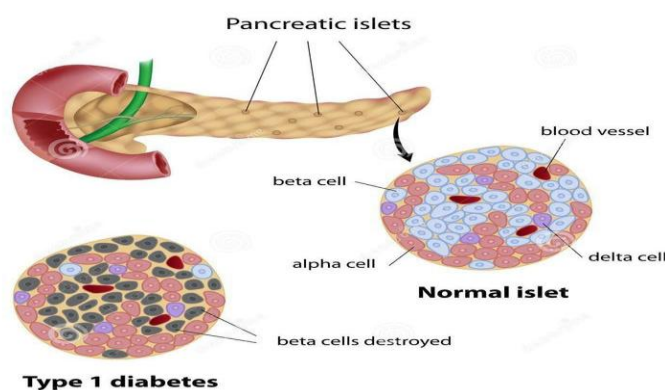


Fig.1 Normal Pancreas Compare to DM affects Pancreas

Types of Diabetes:

- Type-1
- Type-2
- Gestational Diabetes Mellitus

Type-1 DM:

The type-1 diabetes mellitus also known as Insulin dependent diabetes mellitus (IDDM) / juvenile onset diabetes mellitus, in type 1 Diabetes, the pancreatic b-cells undergo autoimmune destruction by CD4+ and CD8+ T cells and macrophages which result in insulin deficiency.

Islet cell antibodies are found in nearly 85% of the patients, and most of them act against the glutamic acid decarboxylase (GAD) found inside the beta-cells of the pancreas. The metabolic disorders related to type 1 diabetes mellitus are a result of deficiency of insulin secretion caused by the immune destruction of islets of Langerhans of the pancreas. Besides, pancreatic alpha-cells start to function abnormally and secrete an excessively large amount of glucagon in the patients with type 1 diabetes mellitus, which further aggravates the metabolic disorders already caused by insulin deficiency. A deficiency of insulin causes lipolysis to occur at an uncontrolled rate which causes the amount of free fatty acids in the blood to rise resulting in a reduction of glucose metabolism in the peripheral tissues. The deficit of insulin also causes a reduction of glucokinase enzyme in the liver

and the GLUT-4 transporter protein in adipose tissue resulting in an inability of the target tissues to respond normally to insulin. Majority of cases are autoimmune (type 1A) antibodies that destroy β cells are detectable in blood, but some are idiopathic (type 1B) - no β cell antibody is found in all type1 cases circulating insulin levels are low or very low, and patients are more prone to ketosis. This type is less common and has a low degree genetic predisposition.[4]

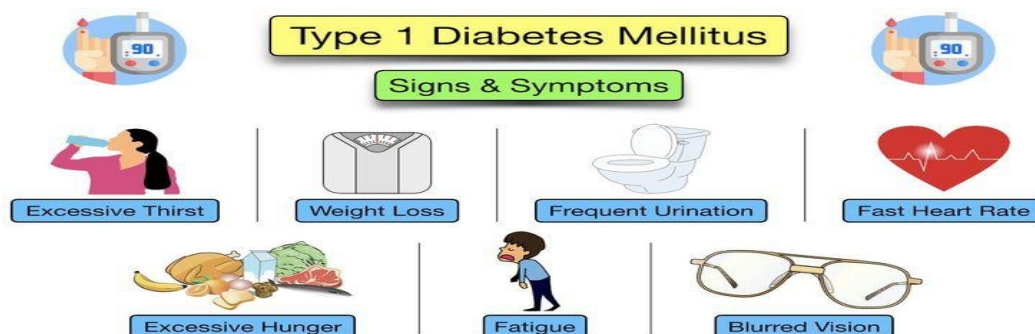


Fig.2 Signs & Symptoms of type -1 DM

Type-2 DM:

Type 2 diabetes mellitus is characterized by insulin resistance, where the body's tissues respond poorly to insulin, often accompanied by reduced insulin secretion. While insulin is produced, it either isn't sufficient or doesn't function properly. Early on, this condition can be managed or even reversed with lifestyle changes and medications that improve insulin sensitivity or reduce liver glucose production. Type 2 diabetes is primarily influenced by genetics and lifestyle factors such as obesity, lack of physical activity, poor diet, stress, and urbanization. It accounts for 90% of diabetes cases and typically occurs in individuals over 40, though it can also develop in younger people with risk factors.[5]

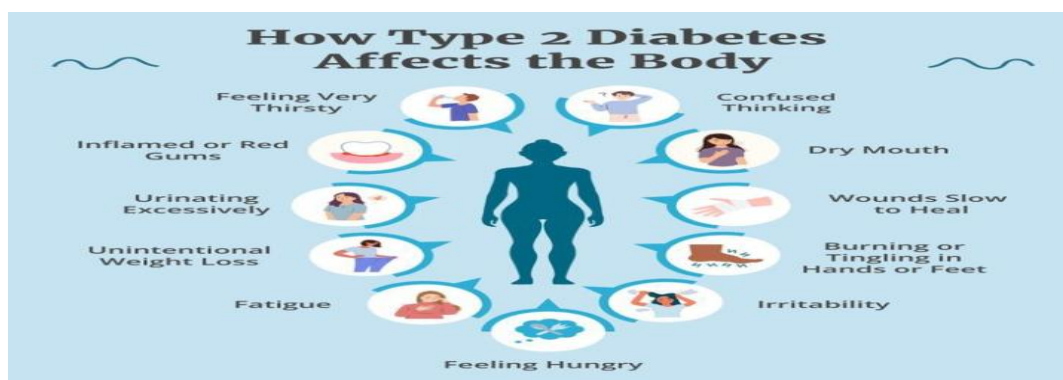


Fig.3 Symptoms of Type -2 DM

Gestational Diabetes Mellitus:

Gestational diabetes is defined as hyperglycemia with first onset during pregnancy and is one of the common pregnancy complications. An estimated 21 million obstetric patients aged 20 to 49 years around the world are affected by GDM, and the incidence rate is up to 16.7%. The GDM is the initial normal glucose metabolism during pregnancy which affected 5 to 6% to 15 to 20% of pregnant women and 85% of live births worldwide [6]. It occurs in women with a family history of diabetes or certain ethnic backgrounds. Proper management is crucial to prevent complications for both the mother and baby, and the condition typically resolves after childbirth, though it increases the risk of Type 2 diabetes later in life. Managing GDM through diet, exercise, or medication is essential for a healthy pregnancy. [5]



Fig.4 Complication of DM

Treatment of diabetes:

The treatment of diabetes involves both drug and non-drug strategies to maintain proper blood glucose levels. Here's an overview of the major components:

A) Drug treatment for diabetes:

Anti-diabetic drugs are used to lower blood glucose levels, and they can be broadly divided into insulin and oral medications. The choice of medication depends on the type of diabetes and individual patient factors like age and overall health.

Type 1 Diabetes:

This is caused by the body's inability to produce insulin. Therefore, insulin therapy is mandatory for these patients. Insulin can be administered through injections or inhalation.

Type 2 Diabetes:

In type 2 diabetes, the body becomes resistant to insulin. Drug treatment for this type focuses on: Increasing insulin production from the pancreas (e.g., sulfonylureas, meglitinides), Enhancing the sensitivity of cells to insulin (e.g., metformin, thiazolidinediones), Reducing glucose absorption from the gastrointestinal tract (e.g., alpha-glycosidase inhibitors). Other injectable drugs like Exenatide (a GLP-1 receptor agonist) and Pramlintide (an amylin analogue) are also used, particularly in combination with other medications.

B) Non-Drug Treatment for Diabetes:

Non-drug management is equally important for all types of diabetes. It includes: Dietary management: A balanced diet, low in refined sugars and rich in fiber, helps control blood sugar. Exercise: Regular physical activity improves insulin sensitivity and helps maintain healthy body weight. Monitoring blood glucose levels: This is crucial for adjusting treatment as needed and preventing complications. Together, these strategies help maintain optimal blood glucose levels and reduce the risk of diabetes-related complications. [5]

INSULIN:

Insulin was discovered in 1921 by William Banting and Best who demonstrated the hypoglycemic action of an extract of pancreas prepared after degeneration of the exocrine part due to ligation of pancreatic duct. It was first obtained in pure crystalline form in 1926 and the chemical structure was fully worked out in 1956 by Sanger. Insulin is a peptide hormone produced by beta cells of the pancreatic islets encoded in humans by the insulin gene. It is made up of the 51 amino acids. It contains 2 chains: A and B. A chain with 21 amino acids, B chain with 30 amino acids.

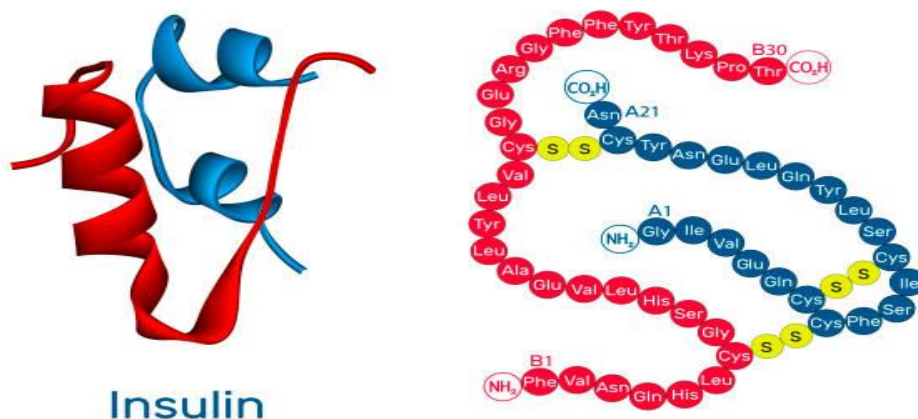


Fig: 5 Human Insulin Structure

Insulin Mediated Glucose Uptake:

The insulin plays the most prominent role. Insulin is powerful anabolic hormone regulates the transport of glucose into the cell through translocation of glucose transporter from an intracellular pool to the plasma membrane mainly in metabolically active tissues like skeletal muscles, adipose tissue, or liver (GLUT4). This translocation occurs through multiple steps of PI3K/AKT signalling pathway. In this chapter, we will focus on molecular events leading to GLUT4 translocation, starting with activation of insulin receptors through signalling cascade involving phosphatidylinositol 3-kinase (PI3K) and protein kinase B (PKB) and finally, the action of their effectors. We will present regulatory mechanisms and modulators of insulin-mediated glucose uptake.

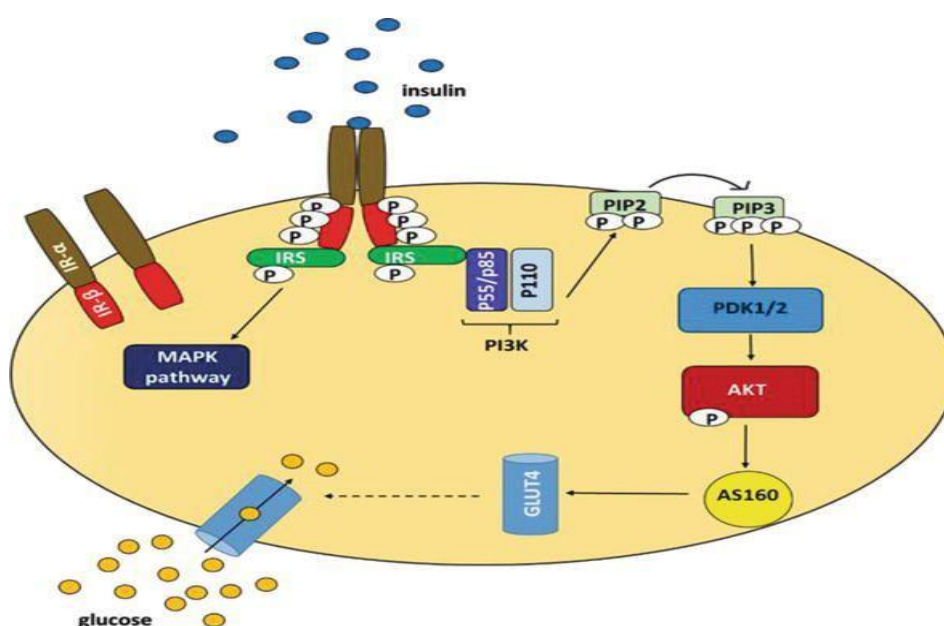


Fig: 6 PI3K/AKT Pathway

Insulin is usually given subcutaneously, either by injections or by an insulin pump. Research is underway of other routes of administration. In acute care settings, insulin may also be given intravenously. There are several types of insulin, characterized by the rate which they are metabolized by the body. Insulin is essential for the treatment of type1 diabetes. For many years it was assumed, as an act of faith, that normalizing plasma glucose would prevent diabetic complications. The diabetes control and complications trial (American diabetes association, 1993) showed that this faith was well placed: type diabetic patients were randomly allocated to intensive or conventional management.[5]

Methodology:**Why We Chose Medicinal Plants:**

Challenges in the Treatment of DM. Existing approaches to the management of DM relied on keeping BGLs with normal limits via administration of appropriate medications together with lifestyle modifications. So far, the accessible medicines for DM are various preparations of insulin and oral antihyperglycemic agents. Older oral hypoglycemics are sulphonylureas, alpha glucosidase inhibitors, thiazolidinediones, and biguanides, while the newer medicines include incretin-based therapies, sodium-glucose cotransporter 2 (SGLT-2) inhibitors, glucokinase activators, and injectable glucagon-like peptide (GLP-1) agonists. Medicines are used either as monotherapy or in combination to achieve better treatment outcomes, conventional and newer agents are still with their shortcomings, and successful treatment of diabetes is being a global challenge requiring further investigations. In fact, these medications are associated with unnecessary drug reactions or side effects including hepatocellular injury, exacerbate renal diseases, blood dyscrasias, gastrointestinal irregularities, hypoglycemia, hypersensitivity reactions, weight gains, and lactic acidosis, which decrease their effectiveness and compliance rates.

For instance, 3.9 and 32.7% severe and no severe hypoglycemic events were reported in 826 (2.8%) patients during their most recent year of sulfonylurea treatment, respectively. Clearly, the weight gain associated with the use of thiazolidinediones (pioglitazone and rosiglitazone), sulphonylureas (glibenclamide), and insulin is also the major drawback for treating diabetes. Several drawbacks have been reported associated with the use of antihyperglycemic medications, such as decreasing effectiveness and increasing adverse effects and toxicities. For instance, sulphonylureas lose their efficacy after six years of therapy in nearly 44% of patients, while BG lowering medicines are stated to be unable to control hyperlipoproteinemia.

In spite of the extensive improvement made in the management of DM using various antihyperglycemic medicines in the past, the results of treatment are still far from successful. Because of the limitations of these agents, there remains a clear need for the identification of new antidiabetic drugs. Therefore, seeking extra safe and effective antidiabetic medicines from plant sources is becoming an active area of research in the scientific community.[7]

Mechanism of Medicinal Plants in Diabetes Mellitus:

Possible Mechanism of Action of Medicinal Plants in Treating DM. Bioactive compounds that are obtained from various MPs have been reported to have potent BG-lowering potentials. Mechanism of lowering BGL might be a result of activation of releasing insulin from β -cells, reduction of glucose absorption, stimulation of glycogenesis, and/or enhancement of glucose. In addition to lowering BGL, secondary metabolites obtained from MPs have the capacity to restore the impaired β -cells and terminate oxidative stress on β -cells. Furthermore, inhibiting cellular apoptosis, reducing renal glucose reabsorption, enhancing the metabolic rate of oxygen consumption, and promoting glucose transporter (GLUT-2) expression and translocation of GLUT-4 are also important mechanisms in diabetic mellitus treated with certain secondary metabolites that are responsible for antidiabetic effects. Blocking pancreatic β -cell K^+ channel, stimulating cyclic adenosine monophosphate (cAMP), and providing some essential elements (calcium, zinc, magnesium, manganese, and copper) for the β -cell are also some mechanisms that are possibly participated in β -cell dysfunction found in DM. Blocking the actions of α -amylase and α -glucosidase enzymes, which are essential for carbohydrate digestion, is used as an optional treatment approach for type 2 diabetes.

Folkloric MPs with antihyperglycemic effects via inhibition of these enzymes and their free radical scavenging potentials are becoming promising modalities in treating type 2 DM and associated complications. Medicinal plants have central roles to discover newer medicines and have begun to get greater attention as sources of bioactive constituents as well as antioxidants. Antioxidant activity has a protective effect in restoring β -cell function in diabetes. Because free radicals are known to

damage and mutation of cells, and hence, oxidative stress has a vital role in the pathogenesis of DM and its complications, MPs with antioxidant effect will have paramount importance in treating diabetes and its complications via scavenging free radicals.[7]

RESULT AND DISCUSSION:

In this article, we discuss medicinal plants used for diabetic mellitus. Medicinal plants have been used for centuries in folk medicine. The medicinal plants in the treatment and management of several ailments and diseases, including diabetes. Traditionally the medicinal plants are used in daily food preparation especially in Tamilnadu, more over south India like. Now the development of science and medicinal technology: the traditionally used spices are identified as having medicinal value phytoconstituents and proved scientifically. There are thousands of plants identified for use in the treatment of diabetes mellitus. Now we discuss some plants that have antidiabetic activity, their Phyto-constituents and effects.

S. No	Biological Name	Family	Parts Used	Phytomolecules	Activity	Reference
1	<i>Mangifera indica</i>	<i>Anacardiaceae</i>	Leaves/ Stem bark	Phenolic Acid, Benzophenones, Flavonoids.	Inhibits glucose absorption in the gut.	[8]
2	<i>Azadirachta indica</i>	<i>Meliaceae</i>	Bark	Isoprenoids, Alkaloids, Flavonoids.	Inhibits key enzymes linked to diabetes.	[9]
3	<i>Caesalpinia crista</i>	<i>Caesalpinaceae</i>	Seed	Flavonoids, Tannins, Alkaloids, Saponins.	Shows antidiabetic and wound healing properties.	[10]
4	<i>Caesalpinia volkensii</i>	<i>Fabaceae</i>	Leaf	Sterols, Flavones, Chalcones, Flavonoids.	Has antidiabetic, anti-malarial, and pain-relieving effects.	[11]
5	<i>Sarcopoterium spinosum</i>	<i>Rosaceae</i>	Root	Corilagin, Pedunculagin, Castalagin.	Used traditionally for diabetes, anti-inflammation, and wound repair.	[12]
6	<i>Cocos nucifera</i>	<i>Arecaceae</i>	Flower	Alkaloids, Flavonoids, Triterpenoids.	Has an antidiabetic and antioxidant property.	[13]
7	<i>Sida acuta</i>	<i>Malvaceae</i>	Root	Diterpenes, Alkaloids, Steroids, Lactones.	Lowers blood glucose via extra-pancreatic pathways.	[14]
8	<i>Withania coagulans</i>	<i>Solanaceae</i>	Fruit	Amino Acids, Fatty Oils, Essential Oils.	Improves insulin sensitivity.	[15]
9	<i>Salacia chinensis</i>	<i>Hippocrateaceae</i>	Roots	Salacinol, Kotalanol, Mangiferin.	Inhibits enzyme activity to reduce diabetes.	[16]
10	<i>Rheum emodi</i>	<i>Polygonaceae</i>	Root	Stilbene, Rhaponticin.	Enhances glucose utilization and mimics insulin.	[17]
11	<i>Leucas aspera</i>	<i>Lamiaceae</i>	Stem, flower, fruit, and leaves	Terpenoids, Glycosides, Flavonoids, Alkaloids, Saponins.	Inhibits glycosylation and reduces haemoglobin levels.	[18]
12	<i>Justicia adhatoda</i>	<i>Acanthaceae</i>	Leaves, root	Alkaloids (Vasicine, Vasicinone).	Reduces blood sugar and tissue lipids.	[19]
13	<i>Ficus amplissima</i>	<i>Moraceae</i>	Bark	Flavonoids, Saponins, Glycosides, Alkaloids.	Shows anti-hyperglycaemic activity.	[20]
14	<i>Helicteres isora</i>	<i>Sterculiaceae</i>	Root	Carbohydrates, Fat, Protein.	Reduces blood glucose, cholesterol, and triglycerides.	[21]
15	<i>Ficus religiosa</i>	<i>Moraceae</i>	Leaves, stem	Flavonoids, Beta Sitosteryl-D-Glucoside.	Used mainly for Type 2 diabetes mellitus.	[22]

16	<i>Costus igneus</i>	<i>Costaceae</i>	Leaves	Steroids, Alkaloids.	Improves glycemic control and insulin secretion.	[23]
17	<i>Ferula asafoetida</i>	<i>Apiaceae</i>	Root	Resins.	Stimulates pancreatic beta cells to secrete insulin.	[24]
18	<i>Lawsonia inermis</i>	<i>Lythraceae</i>	Leaves	Tannins, Saponins, Flavonoids, Terpenoids.	Shows hypoglycemic and other medicinal properties.	[25]
19	<i>Combretum micranthum</i>	<i>Combretaceae</i>	Leaves	Alkaloids, Tannins, Glycosides, Flavonoids.	Has potent antidiabetic and lipid-lowering effects.	[26]
20	<i>Ricinus communis</i>	<i>Euphorbiaceae</i>	Root	Alkaloids, Terpenoids, Flavonoids.	Promising for diabetes treatment with high safety.	[27]
21	<i>Syzygium cumini</i>	<i>Myrtaceae</i>	Seed kernels	Flavonoids, Saponins, Triterpenoids.	Kernel extract shows better hypoglycemic activity.	[28]
22	<i>Murraya koenigii</i>	<i>Rutaceae</i>	Leaf	Alkaloids, Flavonoids, Glycosides.	Shows hypoglycemic and antioxidant activity.	[29]
23	<i>Aerva lanata</i>	<i>Amaranthaceae</i>	Leaf	Alkaloids, Steroids, Kaempferol.	Inhibits α -amylase and α -glucosidase, acting as an antidiabetic agent.	[30]
24	<i>Aeglemarmelos</i>	<i>Rutaceae</i>	Leaf and Seed	Aegeline, Coumarin, Flavonoid, Alkaloid.	Decreases glucose levels, decreases glycosylated haemoglobin.	[31-38]
25	<i>Allium cepa</i>	<i>Alliaceae</i>	Bulb	Allylpropyldisulphide, S-Methylcysteine Sulphoxide.	Inhibit alpha amylase thereby decrease the blood glucose level and anti-oxidant properties.	[39,40]
26	<i>Allium sativum</i>	<i>Alliaceae</i>	Root	Diallyldisulphideoxide ,Ajoene, Allylpropyldisulfide,S-Allylcysteine, S-Allyl Mercaptocysteine.	Decreases glucose levels, decreases lipid levels, increases insulin levels, decreases oxidative stress.	[41,42,43]
27	<i>Aloe barbadensis</i>	<i>Asphodelaceae</i>	Leaf	Lophenol,24-Methyl-Lophenol,24-Ethyllophenol.	Improve Insulin sensitivity, decrease the triglycerides and sugar level.	[49]
28	<i>Brassica juncea</i>	<i>Brassicaceae</i>	Leaf	Isorhamnetin, Diglucoside.	Reducing serum glucose level and preventing insulin resistance.	[46]
29	<i>Lepidium sativum</i>	<i>Brassicaceae</i>	Seed	Flavonoids, Alkaloids, Coumarins, Glycosinolates.	Decreases glucose levels and triglycerides.	[47]

30	<i>Raphanus sativus</i>	<i>Brassicaceae</i>	Root	Flavonoids, Alkaloids, Saponins, Glycosinolate.	Decreases glucose levels, decreases lipid levels, decreases insulin levels.	[48]
31	<i>Beta vulgaris</i>	<i>Chenopodiaceae</i>	All parts of plant	Sugarbeetpectin, Polydextrose.	Enhanced glucose uptake, increase insulin sensitivity.	[44,45]
32	<i>Momordica charantia</i>	<i>Cucurbitaceae</i>	Full plant	Charantin, Momordicin, Galactose-Binding Lectin Nonbitter, Diosgenin, Cholesterol, Lanosterol, Beta Sitosterol, Cucurbitacinyglycoside.	Decreases glucose levels, decreases glycosylated haemoglobin, decreases oxidative stress, increases glycogen levels, decreases lipid peroxidation.	[53-59]
33	<i>Artemisia sphaerocephala</i>	<i>Asteraceae</i>	Fruit	Polysaccharide.	Decreases glucose levels, decreases lipid levels, increases glucose tolerance.	[62,63]
34	<i>Taraxacum officinale</i>	<i>Asteraceae</i>	Fruit	Terpen.	Decreases glucose levels by increase glucose uptake	[64]
35	<i>Terminalia chebula</i>	<i>Combretaceae</i>	Seed Fruit	Palitic acid, Beta-Sitosterol, Daucosterol, Shikimic, Gallic, Triacontanoic.	Decreases glucose levels by increase insulin secretion.	[68,69]
36	<i>Ziziphus spinarum</i>	<i>Rhamnaceae</i>	Leaf	Christinin-A, Fatty acid.	Decreases glucose levels and triglycerides.	[70]
37	<i>Morinda citrifolia</i>	<i>Rubiaceae</i>	Fruit	Saponin, Triterpene, Steroid, Flavonoid.	Decreases glucose levels and increase insulin sensitivity	[66,67]
38	<i>Psidium guajava</i>	<i>Myrtaceae</i>	Leaf Fruit	Terpen, Flavonoid, Strictinin, Isostrictinin, Pedunculagin, Polysaccharide.	Decreases glucose levels in blood and anti-oxidant properties.	[49,50]
39	<i>Syzygium jambolanum</i>	<i>Myrtaceae</i>	Fruit	Anthocyanin, Citric, Malic, Gallic acid.	Decreases glucose levels, increases hepatic glycogen levels.	[51,52]
40	<i>Terminalia brownii</i> Fresen.	<i>Combretaceae</i>	Stem bark	Tannins, Saponins, Polyphenols, Flavonoids, Terpenoids, and Steroids.	Exhibits α -glucosidase inhibitory activity	[71]
41	<i>Artemisia afra</i> Jacq. ex Willd	<i>Asteraceae</i>	Aerial parts	Tannins, Saponins, Chromophores, Phosphosteroid, Withanoids, Flavonoids, and Anthraquinone.	Demonstrates α -glucosidase and α -amylase inhibitory activities.	[72]

42	<i>Stevia Rebaudiana Bertoni</i>	<i>Asteraceae</i>	Leaf	Alkaloids, Steroids, Phenols, and Flavonoids.	Contains steroids, flavonoids which has been shown to have antidiabetic and anti-inflammatory effects.	[73]
43	<i>Pentas schimperiana (A. Rich).</i>	<i>Rubiaceae</i>	Leaf	Flavonoids, Saponins, Steroids, and Tannins.	Exhibits antioxidant and anti-inflammatory activities, which can help manage diabetes.	[74]
44	<i>Aloe Megalacantha Baker</i>	<i>Asphodelaceae</i>	Leaf	Alkaloids, Flavonoids, Phenols, Tannins, Saponins, Glycosides, Anthraquinone & Terpenoids.	Contains aloin, which has been shown to have antidiabetic and anti-inflammatory effects.	[75]
45	<i>Melia azedarach Linn.</i>	<i>Meliaceae</i>	Leaf	Alkaloids, Flavonoids, Triterpenoids, Limonoids.	Exhibits α -glucosidase inhibitory activity and improves insulin sensitivity.	[76]
46	<i>Hagenia Abyssinica</i>	<i>Rosaceae</i>	Flower	Saponins, Tannins, Terpenoids, Phenols, Flavonoids, Glycosides, Steroids, and Anthraquinones.	Shows antioxidant and anti-inflammatory effects, which can help manage diabetes.	[77]
47	<i>Bersama abyssinica Fresen</i>	<i>Melanthaceae</i>	Seeds	Acarbose Alkaloids, Glycosides, Flavonoids, Steroids, Phenols, Tannins, Triterpene, Anthraquinone, Polysterols, and Coumarins.	Exhibits α -glucosidase inhibitory activity.	[78]
48	<i>Aloe Megalacantha Baker</i>	<i>Asphodelaceae</i>	Leaf	Aloe-Emodin, Chrysophanic acid, Flavonoids, Saponins.	Contains aloin, which has been shown to have antidiabetic and anti-inflammatory effect,	[79]
49	<i>Aloe Monticola Reynolds</i>	<i>Asphodelaceae</i>	Leaf	Aloesin, Aloenin, Aloeresin D, Aloin A, Aloin B.	Exhibits antioxidant and anti-inflammatory activities, which can help manage diabetes.	[80]
50	<i>Aloe pulcherrima Gilbert and Sebsebe</i>	<i>Asphodelaceae</i>	Leaf	Flavonoids, Anthraquinone, Saponins, Glycosides, Tannins, Phenols, and Alkaloids.	Shows antioxidant and anti-inflammatory effects, which can help manage diabetes.	[80]
51	<i>Datura stramonium Linn.</i>	<i>Solanaceae</i>	Seed	Saponins, Alkaloids, Flavonoids, Phenols, Tannins, Terpenoids, Glycosides, and Steroids.	Exhibits α -glucosidase inhibitory activity.	[81]

52	<i>Cathaedulis Forsk</i>	<i>Celastraceae</i>	Leaf	Alkaloids, Flavonoids, Quercetin.	Shows antioxidant and anti-inflammatory effects, which can help manage diabetes.	[82]
53	<i>Otostegia integrifolia Benth.</i>	<i>Lamiaceae</i>	Leaf	Phenols, Saponins, Reducing Sugars, and Flavonoids.	Exhibits α -glucosidase inhibitory activity.	[83]
54	<i>Calpurnia aurea (Ait.) Benth.</i>	<i>Fabaceae</i>	Seed	Alkaloids, Phenols, Flavonoids, and Terpenoids.	Shows antioxidant and anti-inflammatory effects, which can help manage diabetes.	[84]
55	<i>Caylusea abyssinica (Fresen.)</i>	<i>Resedaceae</i>	Leaf	Saponins, Flavonoids, and Alkaloids.	Exhibits α -glucosidase inhibitory activity.	[85]
56	<i>Becium grandiflorum Lam.</i>	<i>Lamiaceae</i>	Leaf	Flavonoids Terpenoids, Tannins, Saponins, Phenols, and Steroids.	Shows antioxidant and anti-inflammatory effects, which can help manage diabetes.	[86]
57	<i>Calpurnia aurea (Ait.) Benth.</i>	<i>Fabaceae</i>	Leaf	Phenols, Alkaloids, Terpenoids, and Flavonoids.	Exhibits α -glucosidase inhibitory activity.	[87]
58	<i>Acanthus polystachyus Delile</i>	<i>Acanthaceae</i>	Root	Tannins, Flavonoids, Saponins, Polyphenols, Terpenoids, Glycosides, Anthraquinone.	Shows antioxidant and anti-inflammatory effects, which can help manage diabetes.	[88]
59	<i>Justicia Schimperiana T. Anderson</i>	<i>Acanthaceae</i>	Leaf	Alkaloids, Phenols, and Terpenoids.	Exhibits α -glucosidase inhibitory activity.	[89]
60	<i>Aloe debrana</i>	<i>Asphodelaceae</i>	Leaf	Flavonoids, Phenols, Triterpenoids, Photosterols.	Shows antioxidant and anti-inflammatory effects, which can help manage diabetes.	[90]
61	<i>Croton macrostachys Hocsht. ex Del</i>	<i>Euphorbiaceae</i>	Root	Alkaloids, Phenols, Tannins, Terpenoids, Saponins, Phlobatannins, and Flavonoids.	Exhibits α -glucosidase inhibitory activity.	[91]
62	<i>Lens culinaris</i>	<i>Fabaceae</i>	Seed	Polyphenols, Flavonoids, Saponins, Triterpenoids, Phytates, Lectins Phytosterols, and Defensins.	Shows antioxidant and anti-inflammatory effects, which can help manage diabetes.	[92]

63	<i>Indigofera spicata Forsk</i>	<i>Fabaceae</i>	Leaf	Alkaloids, Glycoside, Tannins, Saponins, Phytosterols, Flavonoids, and Diterpenes.	Exhibits α -glucosidase inhibitory activity.	[93]
64	<i>Vigna radiate</i>	<i>Fabaceae</i>	Grain	Nthocyanin and Bound Phenolic Acids.	Shows antioxidant, anti-inflammatory effects.	[94]
65	<i>Ajuga remota Benth</i>	<i>Lamiaceae</i>	Leaf	Phenols, Flavonoids, Saponins, Tannins, and Steroids.	Exhibits α -glucosidase inhibitory activity.	[95]
66	<i>Tymus vulgaris L.</i>	<i>Lamiaceae</i>	Leaf	Phenols and Flavonoids.	Shows antioxidant and anti-inflammatory effects, which can help manage diabetes.	[96]
67	<i>Salvia tiliifolia Vahl.</i>	<i>Lamiaceae</i>	Aerial parts	Alkaloid, Saponins Flavonoids, and Phytosterols.	Exhibits α -glucosidase inhibitory activity.	[97]
68	<i>Hibiscus deflersii Schweinf ex</i>	<i>Malvaceae</i>	Leaf	Phytosterols, Flavonoids, and Glycosides.	Shows antioxidant and anti-inflammatory effects, which can help manage diabetes.	[98]
69	<i>Moringa stenopetala Baker</i>	<i>Moringaceae</i>	Leaf	Alkaloids, Flavonoids, Tannins, Saponins, Terpenoids.	Exhibits α -glucosidase inhibitory activity and improves insulin sensitivity.	[99]
70	<i>Myrtus communis L.</i>	<i>Myrtaceae</i>	Leaf	Polyphenols, Tannins, and Glucosides.	Shows antioxidant and anti-inflammatory effects, which can help manage diabetes.	[100]
71	<i>Psidium guajava L.</i>	<i>Myrtaceae</i>	Leaf	Alkaloids, Phenols, Flavonoids, Tannins, and Saponins.	Exhibits α -glucosidase inhibitory activity.	[101]
72	<i>Caylusea abyssinica (Fresen.)</i>	<i>Resedaceae</i>	Leaf	Saponins, Flavonoids, and Alkaloids.	Shows antioxidant and anti-inflammatory effects, which can help manage diabetes.	[102]
73	<i>Capsicum frutescens L.</i>	<i>Solanaceae</i>	Fruit	Phenols and Flavonoids.	Exhibits α -glucosidase inhibitory activity and improves insulin sensitivity.	[103]

Table: 1. Plants used for anti-Diabetic

Above the medicinal plants have good antidiabetic activity, which is confirmed by the extraction and fractionation of the plant parts are used for in vivo studies such as aqueous, methanol, ethanol, butanol, hydroalcoholic extraction, chloroform, and petroleum ether fraction. The models for used invitro studies of chromogenic DNSA & in vivo studies of tSTZ-induced diabetic mice. Alloxan-induced diabetic mice. The phytochemicals antidiabetic activity compares to standard drugs such as Acarbose and glibenclamide. From the evaluation, the identified herbs are proved to have potent antidiabetic activity.

CONCLUSION

Finally, we concluded above Phytochemicals have emerged as promising natural agents in the management of diabetes mellitus due to their diverse mechanisms of action, including enzyme inhibition, insulin sensitization, glucose uptake modulation, and antioxidant activity. This review highlights the significant pharmacological potential of flavonoids, alkaloids, phenolics, terpenoids, and glycosides in regulating glucose metabolism and reducing diabetes-associated complications. Their integration into functional foods, nutraceuticals, and pharmaceutical formulations offers a novel and potentially safer alternative to conventional antidiabetic therapies. Despite their therapeutic promise, challenges such as low bioavailability, potential toxicity, and regulatory constraints remain critical barriers to their widespread clinical application. Future research should focus on advanced formulation strategies, rigorous preclinical and clinical evaluations, and exploring synergistic phytochemical combinations to enhance efficacy and therapeutic outcomes. The development of standardized, evidence-based phytochemical therapeutics could pave the way for more effective, sustainable, and patient-friendly approaches in diabetes management.

ACKNOWLEDGMENT


We are sincerely acknowledged to Surya School of Pharmacy, Villupuram, Tamil Nadu, for the great support for our research work.

CONFLICT OF INTERESTS

We have No conflict interest.

AUTHOR CONTRIBUTIONS

Equal contribution for all authors.

S.A. Vadivel  <https://orcid.org/0000-0001-5814-7775>

REFERENCE

1. A.N. Nagappa a, P.A. Thakurdesai a, N. Venkat Rao b Jiwan Singh b, Antidiabetic activity of Terminalia catappa Linn fruits, Journal of Ethnopharmacology 88 (2003) 45–50 DOI: 10.1016/S0378-8741(03)00085-1 .
2. Jirawat Riyaphan, Dinh-Chuong Pham, Max K. Leong, and Ching-Feng Weng 4,Review In Silico Approaches to Identify Polyphenol Compounds as α -Glucosidase and α -Amylase Inhibitors against Type-II Diabetes, Biomolecules 2021, 11(12), 1877; <https://doi.org/10.3390/biom11121877>.
3. Andréia C.F. Salgueiro, Vanderlei Folmer, Felipa E.M. Bassante, Márcia H.S. Cardoso, Hemerson S. da Rosa, Gustavo O. Puntel, Predictive antidiabetic activities of plants used by persons with Diabetes mellitus Complementary Therapies in Medicine Volume 41, December 2018, Pages 1-9. <https://doi.org/10.1016/j.ctim.2018.08.009>.
4. Safaet Alam, Md. Moklesur Rahman Sarker, Taposhi Nahid Sultana, Md. Nafees Rahman Chowdhury, Mohammad A. Rashid, Nusrat Islam Chaity, Chao Zhao, Jianbo Xiao, Elsayed E. Hafez, Shah Alam Khan and Isa Naina MohamedAntidiabetic Phytochemicals from Medicinal Plants: Prospective Candidates for New Drug Discovery and Development, DOI: 10.3389/fendo.2022.800714.

5. S. A. Vadivel, K. Reeta Vijayarani, M. Thennarasi, S. Sandhiya, A. Karthigayani, T. Hari Priya and G. Aathisvadharshan, Bridging Tradition and Modern Science: A Review of Herbal Remedies for The Treatment of Diabetes, European Journal of Biomedical and Pharmaceutical sciences, ISSN 2349-8870 Volume: 11 Issue: 11 Year: 2024.
6. M.Abinaya, R. Kalki, E. Sibichanduru, A. Sivabalan, Sankar Saravanan and S. A. Vadivel A Comprehensive Review Of Phytomolecules Used In Anti-Diabetic Activities, European Journal of Biomedical and Pharmaceutical sciences, ISSN 2349-8870 Volume: 11 Issue: 6 77-90 Year: 2024. <https://www.researchgate.net/publication/382918220>.
7. Berhan Begashaw Yikna, Awgichew Shewasinad Yehualashet, Medicinal Plant Extracts Evaluated In Vitro and In Vivo for Antidiabetic Activities in Ethiopia: Bases for Future Clinical Trials and Related Investigations, Hindawi Evidence-Based Complementary and Alternative Medicine Volume 2021, Article ID 9108499, 24 pages, <https://doi.org/10.1155/2021/9108499>.
8. Amrita Bhowmik, Liakot Ali Khan, Masida Akhter and Begum Rockeya, Studies on the Antidiabetic Effect of Magnifera Indica Stem-Barks and Leaves on Nondiabetic, Type 1 and Type 2 Diabetic Model Rats, A Journal of the Bangladesh Pharmacological Society, 2004; 4: 110-114. DOI: 10.3329/bjp.v4i2.2488.
9. Olakunle Sanni, Ochuko L, Erukaiure, Chika I Chukwuma, Neil A Koorbanally, Collinsu. Ibeji, Md.Shalidul Islam, Azadirchta Indica Inhibit key Enzyme Linked to Type 2 diabetes In Vitro Abates Oxidative Hepatic Injury and Enhance Muscle Glucose uptake Ex -vivo, Biomedicine and Pharmacotherapy, 2019; (109): 734-743.DOI: 10.1016/j.biopha.2018.10.099.
10. Nakul Gupta, Isha Sharma, Meetu Agarwal, Safhi M. Mohammed, Prerna Chauhan, Tarique Anwer, GyasKhan, Anti-Diabetic activity of seed extract of Caesalpinia Crista Linn in Experimental Animals, African Journal of Pharmacy and Pharmacology, 2013; 7(26): 1808-1813. DOI: 10.5897/AJPP2013.3594.
11. Njaji Joan Murugi, Ngugi Mathew Piero, kibiti Cromwell Mwithi, Ngeranwa N.J.Joseph, Njagi Eliud N Mwanniki, Njue M Wilson, Maina David and Gathumbi Peter Karuri, Hypoglycemic Effect of Caesalpinia Wolkensi on Alloxan Induced Diabetes, Asian Journal of Pharmaceutical and Clinical Research, 2012; 5(2): 69-74.DOI: 10.22159/ajpcr.2012.v5i2.1460.
12. Uriel Elyasiyan, Adi Nudel, Nir Skalka, Konstantin Rozenberg, Elyashiv Drori, Rachela Oppenheimer, Zoha Kerem and Tovit Rosenzweig, Anti-Diabetic Activity of Aerial Parts of Sarcopoterium Spinosum, BMC Complementary and Alternative Medicine, 2017; 17: 1-12. DOI: 10.1186/s12906-017-1780-1.
13. Emojevwe Victor and Jerroh E, Antidiabetic Effect of the Cocos Nucifera coconut husk extract, Journal of Medical and Applied Bioscience, 2012; 4: 16-18.
14. Kumari, K., & Augusti, K. T. "Antidiabetic and antioxidant effects of S-methyl cysteine sulfoxide isolated from onions (*Allium cepa* Linn) as compared to standard drugs in alloxan diabetic rats." Indian Journal of Experimental Biology, 2002; 40(9): 1005-1009. DOI: 10.56042/ijeb.v40i9.21760.
15. Samesh S. Elhady, Fadia S.Youssef, Abdulrahman M. Alahdal, Diena M.Almasri and Mohamed L.Ashour, Anti-hyperglycaemic Evaluation of Buddleia indica leaves Using in vitro, in vivo and in silico Studies and its Correlation with the Major Phytoconstituents, Multidisciplinary Digital Publishing Institute, 2021; 10(2351): 1-16. DOI: 10.3390/plants10112351.
16. Kakara Gift Kumar Deepak, Surekha Challa, Ganta Suhasin, Neelapu Nageseawara Rao Reddy, Hosam OElansary and Diaa O. El-Ansary, Antidiabetic and Antilipidemic Activity of Root Extracts of Salacia oblonga against Streptozotocin-Induced Diabetes in Wistar Rats, Multidisciplinary Digital Publishing Institute, 2020; 8: 1-10,doi: 10.3390/pr8030301.
17. Ankur Ptwardhan, Makarand Pimputkar, Radhika Josh, Evaluation of Anti-diabetic Property of Extracts of Different Plant Parts of Salacia chinensis linn., Journal of Biodiversity, Bioprospecting and Development, 2014; 1(1): 1-4. Doi:10.4172/2376-0214.1000107.

18. Muhammad Torequl Islam, Anti-diabetic potential of *Ocimum sanctum* linn., International Journal of Advanced Biochemistry Research, 2022; 6(2): 38-40 doi: 10.33545/26174693.2022.v6.i2a.132.
19. Battu Gango Rao, Anupoju Asha Naga Sai, Devarkonda Ramadevi, Pharmacognostic and in vitro anti diabetic activity on aerial parts of *Leucas Aspera* (willd). Link, Journal of Global Trends in Pharmaceutical Sciences, 2018; 9(2): 5634-5642.
20. Muhammad Gulfraz, Asma Ahmad, Muhammad Javaid Asad, Aliya sadiq, Umara Afzal, Muhammad Imran, Pervez Anwar, Asiya Zeenat, Kashif Sarfraz Abbasi, Sadaf Maqsood and Rahmat Ullah Qureshi, Antidiabetic activities of leaves and root extracts of *Justicia adhatoda* Linn against alloxan induced diabetes in rats, African Journal of Biotechnology, 2011; 10(32): 6101-6105.
21. Sama Venkatesh, G. Dayanada Reddy, B. Madhava Reddy, M.Lakshman, Anti-diabetic activity of *Helicteres isora* root, Aruna Sindhe M, Yadav D Bodke, Kenchappa R, Vinoda BM, Venkatesh Talwara and Nagaraja O, In vivo Antihyperglycemic activity of *Ficus amplissima* smith Bark Extracts, Journal of Chemical and Pharmaceutical Research, 2016; 8(10): 164-167.
22. Palak Tiwari, Nathiya R, Gayathiri Mahalingam, Antidiabetic activity of Endophytic fungi Isolated from *Ficus religiosa*, Asian Journal of Pharmaceutical and Clinical Research, 2017; 10(4): 59-61.
23. Bhat Vishnu, Asuti Naveen, Kamat Akshay, Sikarwar Mukesh. S., Patil M. B., Antidiabetic activity of insulin plant (*costus igneus*) leaf extract in diabetes rats, Journal of Pharmacy Research, 3(3): 608-610.
24. V. M. Nivedha, R. Priyadharshini, S. Rajeshkumar and Palati Sinduja, Anti-diabetic and Antioxidant Activity of *Pterocarpus santalinus* and *Stevia* Herbal Formulation, Journal of Pharmaceutical Research International, 2021; 33(62B): 125-131. doi:10.9734/jpri/2021/v33i62B35178.
25. Syamsudin, Inawati and Hendig Winarno, The Effect of Inai *Lawsonia Inermis* Leaves extract on Blood Sugar Level, Research Journal of Pharmacology, 2008; 2(2): 20-23.
26. Odoh, Uchenna Estella, Osadeb, Patience Ogomaka and Osadebe, Onyinye Hilda, Evaluation of Antidiabetic and Associated Effect of Methanol Extract of *Combretum Micranthum* G Don Combretaceaeor Normoglycemic and Alloxan Induced Diabetes rats, Journal of Global Bioscience, 2020; 9(5): 7260-7279.
27. Poonam shokeen, Prachi Anand,Y. krishna Murali, Vibha Tandon, Antidiabetic activity of ethanolic extract of *Ricinus Communis* and its purified fractions, Food and chemical toxicology, 2008; 46: 3458-3466.
28. Ravi, K. Sivagnanam and S. Subramanian, Antidiabetic Activity of *Eugenia jambolana* seed kernels on streptozotocin – induced diabtes rats, Journal of Medicinal Food, 2004; 7(2): 187-191.
29. Imad M Ani, Rahajoe I Santosa, Muhammad H Yankuzo, Anil K Saxena, Khalid S Alazzawi, The Antidiabetic Activity of Curry leaves *Murraya Koengii* on the glucose Level, Kidney, and Islet of Langerhans of rats with Streptozotocin Induced Diabetes, Makara journal of healthresearch, 2017; (2): 54-60.
30. Musbau Adewunmi Akanji, Samson Olasunkanmi Olukolu and Mutiu Idowu Kazeem, Leaf Extracts of *Aerva lanata* Inhibit the Activities of Type 2 Diabetes – Related Enzyme and possess Antioxidant Properties, Hindawi Oxidative Medicine and Cellular Longevity, 2018; 1: 1-7.
31. Vikrant A, Sharma R. A Review on Fruits Having AntiDiabetic Potential. Journal of Chemical and Pharmaceutical Research. J Chem Pharm Res 2011; 3(2):204-212.
32. Makheswari MU, Sudarsanam D. Database on Antidiabetic indigenous plants of Tamil Nadhu, India. Int J Pharma Sci Res 2012; 3(2):287-293.
33. Kamalakkannan N, Prince PS. The effect of *Aegle marmelos* fruit extract in streptozotocin diabetes: a histopathological study. J Herbal Pharmacother 2005; 5:87-96.

34. Adebajo AC, Ayoola OF, Iwalewa EO, Akindahunsi AA, Omisore NO, Adewunmi CO et al. Antitrichomonal, biochemical and toxicological activities of methanolic extract and some carbazole alkaloids isolated from the leaves of *Murraya koenigii* growing in Nigeria. *Phytomed* 2006; 13(4):246-524.
35. Kesari AN, Gupta RK, Singh SK, Diwakar S, Watal G. Hypo-glycemic and antihyperglycemic activity of *Aegle marmelos* seed extract in normal and diabetic rats. *J Ethnopharmacol* 2006; 107:374-379.
36. Narendhirakannan RT, Subramanian S, Kandaswamy M. Biochemical evaluation of antidiabetogenic properties of some commonly used Indian plants on streptozotocin-induced diabetes in experimental rats. *Clin Experiment Pharmacol Physiol* 2006; 33:1150-1157.
37. Narender T, Shweta S, Tiwari P, Papi Reddy K, Khaliq T, Prathipati P et al. Antihyperglycemic and antidyslipidemic agent from *Aegle marmelos*. *Bioorg Med Chem Lett* 2007; 17(6):1808-1811.
38. Frade TS, Medeiros YS. Animal models to test drugs with potential antidiabetic activity. *J Ethnopharmacol* 2008; 115:173-183.
39. Fetrow CW, Avila JR. *Professional's Handbook of Complementary and Alternative Medicines*. Springhouse, PA: Springhouse Corporation, 1999.
40. Hattori A, Yamada N, Nishikawa T, Fukuda H, Fujino T. Antidiabetic effects of ajoene in genetically diabetic KK-A(y) mice. *J Nutr Sci Vitaminol* 2005; 51(5):382-384.
41. Liu CT, Wong PL, Lii CK, Hse H, Sheen LY. Antidiabetic effect of garlic oil but not diallyl disulfide in rats with streptozotocin-induced diabetes. *Food Chem Toxicol* 2006; 44:1377-1384.
42. El-Demerdash FM, Yousef MI, El-Naga NI. Biochemical study on the hypoglycemic effects of onion and garlic in alloxan-induced diabetic rats. *Food Chem Toxicol* 2005; 43:57-63.
43. Eidi A, Eidi M, Esmaeili E. Antidiabetic effect of garlic (*Allium sativum* L.) in normal and streptozotocin-induced diabetic rats. *Phytomedicine* 2006; 13:624-629.
44. Tanaka M, Misawa E, Ito Y, Habara N, Nomaguchi K, Yamada M et al. Identification of five phytosterols from *Aloe vera* gel as antidiabetic compounds. *Biol Pharm Bull* 2006; 29:1418-1422.
45. Tan BK, Tan CH, Pushparaj PN. Anti-diabetic activity of the semi-purified fractions of *Averrhoa bilimbi* in high fat diet fed-streptozotocin-induced diabetic rats. *J Life Sci* 2005; 76:2827-2839. DOI: 10.1016/j.lfs.2004.10.051.
46. Eddouks M, Maghrani M, Zeggwagh NA, Michel JB. Study of the hypoglycaemic activity of *Lepidium sativum* L. aqueous extract in normal and diabetic rats. *J Ethnopharmacol* 2005a; 97:391-395. DOI: 10.1016/j.jep.2004.11.030.
47. Taniguchi H, Kobayashi-Hattori K, Tenmyo C, Kamei T, Uda Y, Sugita-Konishi Y et al. Effect of Japanese radish (*Raphanus sativus*) sprout (Kaiware-daikon) on carbohydrate and lipid metabolisms in normal and streptozotocin-induced diabetic rats. *Phytother Research* 2006; 20:274-278. DOI: 10.1002/ptr.1840.
48. Kubo H, Kobayashi J, Higashiyama K, Kamei J, Fujii Y, Ohmiya S. The hypoglycemic effect of (7R*, 9aS*)-7-phenyl-octahydroquinolizin-2-one in mice. *Biol Pharm Bull* 2000; 23(9):1114-1117. DOI: 10.1248/bpb.23.1114.
49. Ojewole JA. Hypoglycemic and hypotensive effects of *Psidium guajava* Linn. (Myrtaceae) leaf aqueous extract. *Methods Findings Experiment Clin Pharmacol* 2005d; 27:689-695. DOI: 10.1358/mf.2005.27.10.932303.
50. Musabayane CT, Mahlalela N, Shode FO, Ojewole JA. Effects of *Syzygium cordatum* (Hochst.) [Myrtaceae] leaf extract on plasma glucose and hepatic glycogen in streptozotocin-induced diabetic rats. *J Ethnopharmacol* 2005; 97:485-490. DOI: 10.1016/j.jep.2004.11.031.
51. Chauhan A, Sharma PK, Srivastava P, Kumar N, Duehe R. Plants having potential antidiabetic activity: a review. *Der Pharm Lett* 2010; 2(3):369-387.
52. Cherian S, Augusti KT. Antidiabetic effect of a glycoside of pelargonidin isolated from the bark of *Ficus bengalensis* Linn. *Indian J Exp Biol* 1993; 31(1):26-29.

53. Saxena A, Vikram NK. Role of selected Indian plants in management of type 2 diabetes: a review. *J Altern Complement Med* 2004; 10(2):369-378. DOI: 10.1089/10755304323062365.
54. Shetty AK, Kumar GS, Sambaiah K, Salimath PV. Effect of bitter gourd (*Momordica charantia*) on glycaemic status in streptozotocin induced diabetic rats. *Plant Foods Human Nut* 2005; 60:109-112. DOI: 10.1007/s11130-005-6838-2.
55. Sathishsekar D, Subramanian S. Beneficial effects of *Momordica charantia* seeds in the treatment of STZ-induced diabetes in experimental rats. *Biol Pharm Bull* 2005; 28:978-983. DOI: 10.1248/bpb.28.978.
56. Sekar DS, Sivagnanam K, Subramanian S. Antidiabetic activity of *Momordica charantia* seeds on streptozotocin induced diabetic rats. *Pharmazie* 2005; 60:383-387 DOI: 10.1691/ph.2005.5.4550.
57. Yadav UC, Moorthy K, Baquer NZ. Combined treatment of sodium orthovanadate and *Momordica charantia* fruit extract prevents alterations in lipid profile and lipogenic enzymes in alloxan diabetic rats. *Molecular Cellular Bioch* 2005; 268:111-120. DOI: 10.1007/s11010-005-3720-0.
58. Harinantenaina L, Tanaka M, Takaoka S, Oda M, Mogami O, Uchida M et al. *Momordica charantia* constituents and antidiabetic screening of the isolated major compounds. *Chem Pharm Bull* 2006; 54:1017- 1021. DOI: 10.1248/cpb.54.1017.
59. Reyes BA, Bautista ND, Tanquilut NC, Anunciado RV, Leung AB, Sanchez GC et al. Anti-diabetic potentials of *Momordica charantia* and *Andrographis paniculata* and their effects on estrous cyclicity of alloxan-induced diabetic rats. *J Ethnopharmacol* 2006; 105:196-200. DOI: 10.1016/j.jep.2005.10.022.
60. Hernández-Galicia E, Calzada F, Roman-Ramos R, Alarcón-Aguilar FJ. Monoglycerides and fatty acids from *Ibervillea sonorae* root: isolation and hypoglycemic activity. *Planta Med* 2007; 73(3):236-240. DOI: 10.1055/s-2007-967121.
61. Suryanarayana P, Kumar PA, Saraswat M, Petrash JM, Reddy GB. Inhibition of aldose reductase by tannoid principles of *Embllica officinalis*: implications for the prevention of sugar cataract. *Mol Vis* 2004; 10:148-154. DOI: 10.1167/molvis.10.20.
62. Sharma SB, Nasir A, Prabhu KM, Murthy PS. Antihyperglycemic effect of the fruit-pulp of *Eugenia jambolana* in experimental diabetes mellitus. *J Ethnopharmacol* 2006; 104:367-373. DOI: 10.1016/j.jep.2005.10.033.
63. K, Rajasekaran S, Subramanian S. Antihyperlipidemic effect of *Eugenia jambolana* seed kernel on streptozotocin-induced diabetes in rats. *Food Chem Toxicol* 2005; 43:1433-1439. DOI: 10.1016/j.jep.2004.11.034.
64. Singh LW. Traditional medicinal plants of Manipur as antidiabetics. *J Med Plant Res* 2011; 5(5):677-687. DOI: 10.1055/s-2005-873137.
65. Shirwaikar A, Rajendran K, Punitha IS. Antidiabetic activity of alcoholic stem extract of *Coscinium fenestratum* in streptozotocin-nicotinamide-induced type 2 diabetic rats. *J Ethnopharmacol* 2005; 97:369-374. DOI: 10.1016/j.jep.2005.07.016.
66. Navarrete A, Mata R. Antihyperglycemic effect of constituents from *Hintonia standleyana* in streptozotocin-induced diabetic rats. *Planta Medica* 2005; 71:1099-1105. DOI: 10.1016/j.pls.2005.08.027.
67. Ojewole JA. Antinociceptive, anti-inflammatory and antidiabetic properties of *Hypoxis hemerocallidea* Fisch. and C.A. Mey. (Hypoxidaceae) corm ['African Potato'] aqueous extract in mice and rats. *J Ethnopharmacol* 2006; 103:126-134. DOI: 10.1016/j.jep.2005.04.009.
68. Lo HC, Tsai FA, Wasser SP, Yang JG, Huang BM. Effects of ingested fruiting bodies, submerged culture biomass, and acidic polysaccharide lucuronoxylomannan of *Tremella mesenterica* Retz: Fr. on glycemic responses in normal and diabetic rats. *J Life Sci* 2006; 78:1957- 1966. DOI: 10.1159/000497725.

69. Abdel-Zaher AO, Salim SY, Assaf MH, Abdel-Hady RH. Antidiabetic activity and toxicity of *Zizyphus spinachristi* leaves. *J Ethnopharmacol* 2005; 101:129-138. DOI: 10.1016/j.jep.2005.04.007.
70. Mohamed B, Ziyat A, Mekhfi H, Tahri A, Legssyer A. Medicinal plants with potential antidiabetic activity - A review of ten years of herbal medicine research (1990- 2000). *Int J Diabetes & Metabol* 2006; 14:1-25. DOI: 10.1159/000497588.
71. N. M. Alema, G. Periasamy, G. G. Sibhat, G. H. Tekulu, and M. G. Hiben, "Antidiabetic activity of extracts of *Terminalia brownii* Fresen. Stem bark in mice," *Journal of Experimental Pharmacology*, vol. 12, pp. 61–71, 2020. DOI: 10.2147/JEP.S240266.
72. A. Issa and M. Hussien Bule, "Hypoglycemic effect of aqueous and methanolic extract of *Artemisia afra* on alloxan induced diabetic Swiss albino mice," *Evidence-Based Complementary and Alternative Medicine*, vol. 2015, Article ID 752486, 5 pages, 2015. DOI: 10.1155/2015/752486.
73. T. Bekele, "Antidiabetic activity and phytochemical screening of crude extracts of *Stevia rebaudiana* Bertoni and *Ajuga remota* Benth grown in Ethiopia on alloxan-induced diabetic mice," pp. 1–60, 2008, <https://www.researchgate.net/publication/271193466>.
74. T. Dinku, S. Tadesse, and K. Asres, "Antidiabetic activity of the leaf extracts of *Pentas schimperiana* subsp. *schimperiana* (A. Rich) vatke on alloxan-induced diabetic mice," *Ethiopian Pharmaceutical Journal*, vol. 28, pp. 12–20, 2010.
75. W. W. Hammes, Y. K. Emiru, K. Ayalew Getahun, and W. Kahaliw, "Antidiabetic and antihyperlipidemic activities of the leaf latex extract of *Aloe megalacantha* baker (Aloaceae) in streptozotocin-induced diabetic model," *EvidenceBased Complementary and Alternative Medicine*, vol. 2019, Article ID 8263786, 9 pages, 2019.
76. D. Seifu, L. Gustafsson, and R. Chawla, "Antidiabetic and gastric emptying inhibitory effect of herbal *Melia azedarach* leaf extract in rodent models of diabetes type 2 mellitus," *Journal of Experimental Pharmacology*, vol. 9, pp. 23–29, 2017. DOI: 10.2147/JEP.S129826.
77. Z. D. Kifle, J. S. Yesuf, and S. A. Atnafie, "Evaluation of in vitro and in vivo anti-diabetic, anti-hyperlipidemic and antioxidant activity of flower crude extract and solvent fractions of *Hagenia abyssinica* (Rosaceae)," *Journal of Experimental Pharmacology*, vol. 12, pp. 151–167, 2020. DOI: 10.2147/JEP.S249964.
78. Kifle and E. F. Enyew, "Evaluation of in vivo antidiabetic, in vitro alpha-amylase inhibitory, and in vitro antioxidant activity of leaves crude extract and solvent fractions of *Bersama abyssinica* Fresen. (melianthaceae)," *Journal of Evidence-Based Integrative Medicine*, vol. 25, pp. 1–11, 2020. DOI: 10.1177/2515690X20935827.
79. G. H. Tekulu, E. M. Araya, and H. G. Mengesha, "In vitro α -amylase inhibitory effect of TLC isolates of *Aloe megalacantha* baker and *Aloe monticola* Reynolds," *BMC Complementary and Alternative Medicine*, vol. 19, no. 1, pp. 1–7, 2019. DOI: 10.1186/s12906-019-2470-0.
80. G. G. Amare, B. G. Meharie, and Y. M. Belayneh, "Evaluation of antidiabetic activity of the leaf latex of *Aloe pulcherrima* gilbert and *sebsebe* (Aloaceae)," *Evidence-based Complementary and Alternative Medicine*, vol. 2020, Article ID 8899743, 9 pages, 2020. DOI: 10.1155/2020/8899743.
81. B. C. Melaku and G. Getnet, "Evaluation of antidiabetic and antioxidant potential of hydromethanolic seed extract of *Datura stramonium* in mice," *Journal of Clinical and Experimental Pharmacology*, vol. 10, no. 2, pp. 1–7, 2020. DOI: 10.35248/2161-1459.20.10.264.
82. D. A. Debecho, Y. A. Kinfu, T. T. Dugul, and D. S. Melka, "Effect of fresh juice of *Khat* (*Catha edulis*) on blood glucose levels of normoglycemic and streptozotocin-induced diabetic rats," *International Journal of Pharmaceutical Sciences and Research*, vol. 9, no. 2, pp. 784–789, 2028. DOI: 10.13040/IJPSR.0975-8232.9(2).784-89.
83. Z. Shewamene, M. Abdelwuhab, and Z. Birhanu, "Methanolic leaf extract of *Otostegia integrifolia* Benth reduces blood glucose levels in diabetic, glucose loaded and normal rodents,"

- BMC Complementary and Alternative Medicine, vol. 15, no. 19, pp. 1–7, 2015. DOI: 10.1186/s12906-015-0547-0.
84. Y. M. Belayneh, M. E. Birru, and D. Ambikar, “Evaluation of hypoglycemic, antihyperglycemic and antihyperlipidemic activities of 80% methanolic seed extract of *Calpurnia aurea* (Ait.) Benth. (Fabaceae) in mice,” *Journal of Experimental Pharmacology*, vol. 11, pp. 73–83, 2019. DOI: 10.2147/JEP.S200923.
85. T. Gebreyohannis, W. Shibeshi, and K. Asres, “Effects of solvent fractions of *Caylusea abyssinica* (Fresen.) fisch. & meyer. On blood glucose levels of normoglycemic, glucose loaded and streptozotocin-induced diabetic rodents,” *Journal of Natural Remedies*, vol. 14, no. 1, pp. 67–75, 2014. DOI: 10.18311/jnr/2014/206.
86. L. Gebremeskel, K. B. Tuem, and T. Teklu, “Evaluation of antidiabetic effect of ethanolic leaves extract of *Becium grandiflorum* lam. (Lamiaceae) in streptozotocin-induced diabetic mice,” *Diabetes, Metabolic Syndrome and Obesity: Targets and %erapy*, vol. 13, pp. 1481–1489, 2020.
87. Y. M. Belayneh and E. M. Birru, “Antidiabetic activities of hydromethanolic leaf extract of *Calpurnia aurea* (ait.) Benth. Subspecies *aurea* (Fabaceae) in mice,” *Evidence-Based Complementary and Alternative Medicine*, vol. 2018, Article ID 3509073, 9 pages, 2018. DOI: 10.1155/2018/3509073.
88. D. Derebe, M. Wubetu, and A. Alamirew, “Hypoglycemic and antihyperglycemic activities of 80% methanol root extract of *Acanthus polystachyus delile* (Acanthaceae) in type 2 diabetic rats,” *Clinical Pharmacology: Advances and Applications*, vol. 12, pp. 149–157, 2020. doi: 10.2147/CPAA.S273501.
89. A. Tesfaye, E. Makonnen, and S. Gedamu, “Hypoglycemic and antihyperglycemic activity of aqueous extract of *Justicia Schimperiana* leaves in normal and streptozotocin-induced diabetic mice,” *International Journal of Pharma Sciences and Research*, vol. 7, no. 2, pp. 107–113, 2016. <https://www.researchgate.net/publication/312120806>.
90. A. Suleyman, N. Gnanasekaran, and S. Daniel, “Amelioration of streptozotocin-induced hyperglycemia and dyslipidemia through *Aloe debrana*,” *International Journal of Pharmacy and Pharmaceutical Sciences*, vol. 7, no. 2, pp. 290–293, 2015. <https://doi.org/10.1155/2021/9108499>.
91. L. Bantie and E. Gebeyehu, “Antidiabetic activity of hydroalcoholic extract of the root of *Croton macrostachys* in Streptozotocin induced diabetic mice,” *World Journal of Pharmaceutical Sciences*, vol. 3, no. 2, pp. 185–191, 2015. <https://www.researchgate.net/publication/271589637>.
92. M. M. Tefera, B. M. Altaye, E. M. Yimer, D. F. Berhe, and S. T. Bekele, “Antidiabetic effect of germinated *Lens culinaris medik* seed extract in streptozotocin-induced diabetic mice,” *Journal of Experimental Pharmacology*, vol. 12, pp. 39–45, 2020. <https://doi.org/10.2147/JEP.S228834>.
93. E. M. Birru, M. Abdelwuhab, and Z. Shewamene, “Effect of hydroalcoholic leaves extract of *Indigofera spicata* Forssk. on blood glucose level of normal, glucose loaded and diabetic rodents,” *BMC Complementary and Alternative Medicine*, vol. 15, p. 321, 2015. DOI:10.1186/s12906-015-0852-8.
94. K. D. Ayenewu, “Antidiabetic Activity of aqueous extract from *Vigna radiata* in streptozotocin induced diabetic mice,” *Indo American Journal of Pharmaceutical Research*, vol. 7, no. 2, pp. 7718–7721, 2017. <https://doi.org/10.1155/2021/9108499>.
95. T. B. Tafese, A. Hymete, Y. Mekonnen, and M. Tadesse, “Antidiabetic activity and phytochemical screening of extracts of the leaves of *Ajuga remota* Benth on alloxan-induced diabetic mice,” *BMC Complementary and Alternative Medicine*, vol. 17, no. 243, pp. 1–9, 2017. DOI 10.1186/s12906-017-1757-5.
96. E. Dessalegn, G. Bultosa, G. D. Haki, and H. P. V. Rupasinghe, “Evaluation of In vitro antidiabetic potential of *Thymus schimperi* R. And *%ymus vulgaris* L,” *Journal of Health, Medicine and Nursing*, vol. 69, pp. 1–19, 2029.

97. K. Gebru, G. G. Sibhat, and M. G. Hiben, "Preliminary phytochemical screening and blood glucose lowering activity of methanol extract of *Salvia tilifolia* vahl aerial part, International Journal of Pharmaceutical & Biological Archives, vol. 5, no. 5, pp. 122–125, 2014. <https://www.researchgate.net/publication/282603756> .
98. S. Lakshman, B. A. Jyothi, V. Mounica, A. R. Kumar, K. M. S. Rathinam, and K. Rajesh, "Antidiabetic activity of methanolic extract of streptozotocin induced diabetic rats *Hibiscus deflersii*," International Research Journal of Pharmaceutical and Applied Sciences, vol. 4, no. 1, pp. 1–3, 2014 <https://doi.org/10.1155/2021/9108499> .
99. Nardos, E. Makonnen, and A. Debella, "Effects of crude extracts and fractions of *Moringa stenopetala* (Baker f) Cufodontis leaves in normoglycemic and alloxan-induced diabetic mice," African Journal of Pharmacy and Pharmacology, vol. 5, no. 20, pp. 2220–2225, 2011. DOI: 10.5897/AJPP11.318.
100. Toma, E. Makonnen, Y. Mekonnen, A. Debella, and S. Addisakwattana, "Extract of leaves of *Moringa stenopetala* alleviates glycation in in vitro model," EC Nutrition, vol. 11, no. 6, pp. 218–222, 2017. DOI: 10.2147/JEP.S371354.
101. Godebo, E. Makonnen, and N. Mekonnen, "Hypoglycemic and antihyperglycemic effect of leaves extracts of *Psidium guajava* in normoglycemic and streptozotocin-induced diabetic mice," International Journal of Pharmacognosy, vol. 4, no. 8, pp. 262–268, 2017. doi: 10.1186/1472-6882-12-151.
102. W. Tamiru, E. Engidawork, and K. Asres, "Evaluation of the effects of 80% methanolic leaf extract of *Caylusea abyssinica* (Fresen.) fisch. & Mey. on glucose handling in normal, glucose loaded and diabetic rodents," BMC Complementary and Alternative Medicine, vol. 12, pp. 1–7, 2012.
103. E. Dessalegn, "Effect of extraction solvents on antioxidant and α -amylase inhibition activities of spiced green chili paste: an in vitro study," Advances in Life Science and Technology, vol. 77, pp. 1–9, 2020. DOI: 10.7176/ALST/77-02.