



Predictors of sirolimus pharmacokinetic variability identified using a nonlinear mixed effects approach: a systematic review

Janthima Methaneethorn^{1,2*}, Premsuda Art-arsa¹, Ramanya Kosiyaporn¹, Nattawut Leelakanok³

¹Department of Pharmacy Practice, Faculty of Pharmaceutical Sciences, Naresuan University, Phitsanulok, Thailand

²Center of Excellence for Environmental Health and Toxicology, Naresuan University, Phitsanulok, Thailand

³Department of Clinical Pharmacy, Faculty of Pharmaceutical Sciences, Burapha University, Chonburi, Thailand

***Corresponding author:** Janthima Methaneethorn, Faculty of Pharmaceutical Sciences, Naresuan University, Phitsanulok 65000, Thailand. Email: janthima.methaneethorn@gmail.com

Submitted: 3 August 2022; Accepted: 2 September 2022; Published: 24 October 2022

ABSTRACT

Several sirolimus (SRL) population pharmacokinetics (PopPK) were conducted to explain its pharmacokinetic variability, and the results varied across studies. Thus, we conducted a systematic review to summarize significant predictors influencing SRL pharmacokinetic variability. Moreover, discrepancies in model methodologies across studies were also reviewed and discussed. Four databases (PubMed, CINAHL Complete, Science Direct, and Scopus) were systematically searched. The PICO framework was used to identify eligible studies conducted in humans and employ a nonlinear-mixed effects strategy. Based on the inclusion and exclusion criteria, 20 studies were included. SRL pharmacokinetics were explained using 1- or 2-compartment models. Only one study assessed the model using an external approach, while the rest employed basic or advanced internal approaches. Significant covariates influencing SRL pharmacokinetics were bodyweight, age, *CYP3A5* polymorphism, gender, BSA, height, cyclosporine dose or trough concentration, triglyceride, total cholesterol, hematocrit, albumin, aspartate aminotransferase, alanine aminotransferase, and total bilirubin. Of these, bodyweight, age, and *CYP3A5* polymorphism were the three most identified significant predictors for SRL clearance. This review summarizes significant predictors to predict SRL clearance, which can subsequently be used to individualize SRL maintenance dose. However, the PopPK model selected for such prediction should be based on the resemblance of population characteristics

between the target population and those used to conduct the model. Moreover, the predictability of the models in the target population should be assessed before implementation in clinical practice.

Keywords: *anticancer; immunosuppressant; nonlinear mixed-effects; population pharmacokinetics; sirolimus*

INTRODUCTION

Sirolimus (SRL) or rapamycin is an immuno-suppressive agent approved for the prophylaxis of graft rejection in kidney transplant patients aged 13 years or more and for the treatment of patients with lymphangioleiomyomatosis.¹ Moreover, SRL exerts anti-tumor action, and recently its derivative, tem-sirolimus (TEM) or CCI-779, has been developed to treat advanced renal cell carcinoma.² Though SRL has a similar structure to tacrolimus and binds to FK-binding protein, SRL/FK-binding protein complex does not affect calcineurin phosphatase. Instead, it binds to mammalian targets of rapamycin (mTOR), leading to the inhibition of the progression of the cell cycle from the G₁ to the S phase.^{1,3,4}

Following oral administration in stable renal transplant patients, the time to peak concentration (T_{max}) is variable, ranging from 0.5 to 3 hours,⁵ however, the bioavailability is low with a value of approximately 15.0%.^{5,6} In blood, SRL is preferentially distributed to red blood cells (94.5%) in a concentration-independent manner,⁷ with the mean blood to plasma ratio of 34.5:1 in stable kidney transplant patients receiving a single oral dose, nonetheless, this ratio is substantially variable ranging from 10 to 70.⁸ In contrast, in the plasma, approximately 40.0% of the drug is bound to lipoproteins,⁵ therefore, based on the nature of SRL, therapeutic drug monitoring (TDM) should be performed using whole blood as the appropriate biological matrix. Moreover, due to the lipophilic property, SRL is extensively distributed in lipid membranes of various organs, resulting in a high volume of distribution (V_d) of 5.6–16.7 L/kg in stable renal transplant patients.⁸

SRL is extensively metabolized by CYP3A4 and CYP3A5. Also, it is a substrate of P-glycoprotein (P-gp), which contributes to its low oral bioavailability.⁵ Moreover, CYP3A5 also plays a role in SRL metabolism.⁹ The drug exhibits large interindividual variability (IIV) in metabolism, with the apparent clearance ranging from 0.090–0.416 L/h/kg in stable renal transplant patients receiving a 14-day course of SRL with cyclosporine and prednisolone.⁸ The elimination half-life ($t_{1/2}$) was approximately 62 hours,⁸ thus, the steady-state condition is achieved within 1–2 weeks.

Based on preclinical and clinical studies, the efficacy and adverse effects of SRL are related to blood concentrations, with trough concentrations at steady-state ($C_{ss,tr}$) of greater than 5 µg/L associated with an 89.5% negative predictive value for the occurrence of acute rejection episodes, while $C_{ss,tr}$ levels greater than 15 µg/L are related to toxicity such as leucopenia, thrombocytopenia, and hypertriglyceridemia.^{10,14} This suggests that $C_{ss,tr}$ of SRL should be maintained within the range of 5–15 µg/L to achieve an optimal therapeutic outcome,⁵ and thus TDM is an essential process during SRL therapy.

Since it takes approximately one week for SRL to reach steady-state condition, and a dosage adjustment based on the target $C_{ss,tr}$ cannot be performed sooner, a population pharmacokinetic (PopPK) approach can be conducted to aid dosage individualization. This approach and the Bayesian estimation can provide individual pharmacokinetic parameter estimates for the optimization of SRL therapy. To date, several PopPK studies of SRL have been developed to characterize factors influencing SRL pharmacokinetic variability,^{9,15–29} however,

significant predictors obtained from these studies are not consistent; for example, some studies identified a significant effect of gender on SRL clearance (CL_{SRL})²⁷ and V_d ²⁵, but other studies could not find such the effects.^{15,17,19,21,24} Based on the conflicting results, we aimed to systematically summarize factors that significantly influence SRL pharmacokinetic variability and their relationships with pharmacokinetic parameters. In addition, the disparity of model methodologies across studies was also reviewed and discussed.

METHODS

Database Searching and Study Selection

CINAHL Complete, PubMed, Science Direct, and SCOPUS databases were systematically searched to identify apposite studies. The search spanned the period from the database's inception to May 2021. Search terms were developed using the PICO framework as follows: P: human studies, I: sirolimus OR Rapamune or rapamycin, C: none, O: "population pharmacokinetics" OR "pharmacokinetic model" OR "nonlinear mixed effect" OR NONMEM OR "interindividual variability" OR "intersubject variability" OR "residual variability" OR "intrasubject variability." Reference lists were also screened for additional studies.

Title and abstracts were screened to exclude non-relevant articles. Screening of full-text articles was subsequently performed to identify studies to be included in this systematic review based on the following inclusion criteria: 1) PopPK studies conducted in humans, 2) SRL was used as a treatment drug, and 3) studies conducted using a nonlinear mixed-effects approach, while the exclusion criteria included: 1) non-English or non-Thai articles, 2) information on model development methodology was not sufficient, and 3) studies that were not original research articles. The Preferred Reporting Items for Systematic Reviews and Meta-analyses (PRISMA) guideline was adopted and followed during the review process.

Data Extraction

JM, PA, and RK independently extracted the data using the data abstraction form developed by JM. The extracted data were discussed, and the consensus was made by all authors. Three data categories were extracted, which included: 1) population characteristics such as study design, sample size, underlying diseases, age, body size, sex, functions of elimination organs, measurements of relevant laboratory values; 2) pharmacokinetic related information, that is, SRL dosage regimens, concurrent medications, analytical methods of SRL concentrations, and sampling strategy which is categorized into sparse sampling if the number of samples per patients was less than 6, otherwise it was considered an extensive sampling approach, and 3) model development methodologies and model evaluations. For model development, structural and statistical models for IIV and residual variability (RV) were summarized. In addition, tested and significant covariates on SRL pharmacokinetic parameters were compared across studies. Model evaluations were classified into three categories including basic internal, advanced internal, and external evaluation, as previously described by Brendel et al.³⁰

Transparent Report and Clarity of the Included Studies

All reviewers independently assessed the study report quality using the Clinical PK checklist developed by Kanji et al.³¹ and the PopPK model-building strategies introduced by Dartois et al.³²

RESULTS

Study Identification

Based on the systematic search, 992 articles were identified from all databases. Following the removal of duplicates, titles and abstracts of 904 non-redundant studies were screened, and 873 articles were excluded as irrelevant, leaving 31 studies for full-text assessment. Of these, 20 studies published between 1997 and 2021 met the inclusion

criteria and were included in this review. Details on study exclusion are summarized and presented in a PRISMA diagram (Figure 1).

Population Characteristics, Study Design, and Pharmacokinetic Data

Most SRL PopPK studies were performed on various types of transplant patients, including kidney transplants,^{9,15,17,20,26} heart transplants,¹⁹ pancreatic islet transplants,¹⁸ and bone marrow transplants,²² while cancer was the second most disease in which SRL PopPK studies were conducted,^{21,27,28,33–35} which included renal cell carcinoma, Kaposiform heman-gioendothelioma, and several types of advanced

or recurrent solid tumor, and of these two studies developed SRL PopPK models as an active metabolite of TEM.^{33,34} Other underlying diseases included immune cytopenia,²⁹ tuberous sclerosis complex,³⁶ and vascular anomalies.^{23,24} Moreover, one study was conducted solely on healthy Chinese subjects,²⁵ and the other one was performed in both healthy Chinese and kidney transplant patients.³⁷ In terms of age category, respective nine and 11 studies were conducted on children and adults. Most SRL PopPK studies were conducted to characterize SRL pharmacokinetics and its variability, but a few studies specifically aimed to determine the initial SRL dose using the developed PopPK models^{28,36} or create a

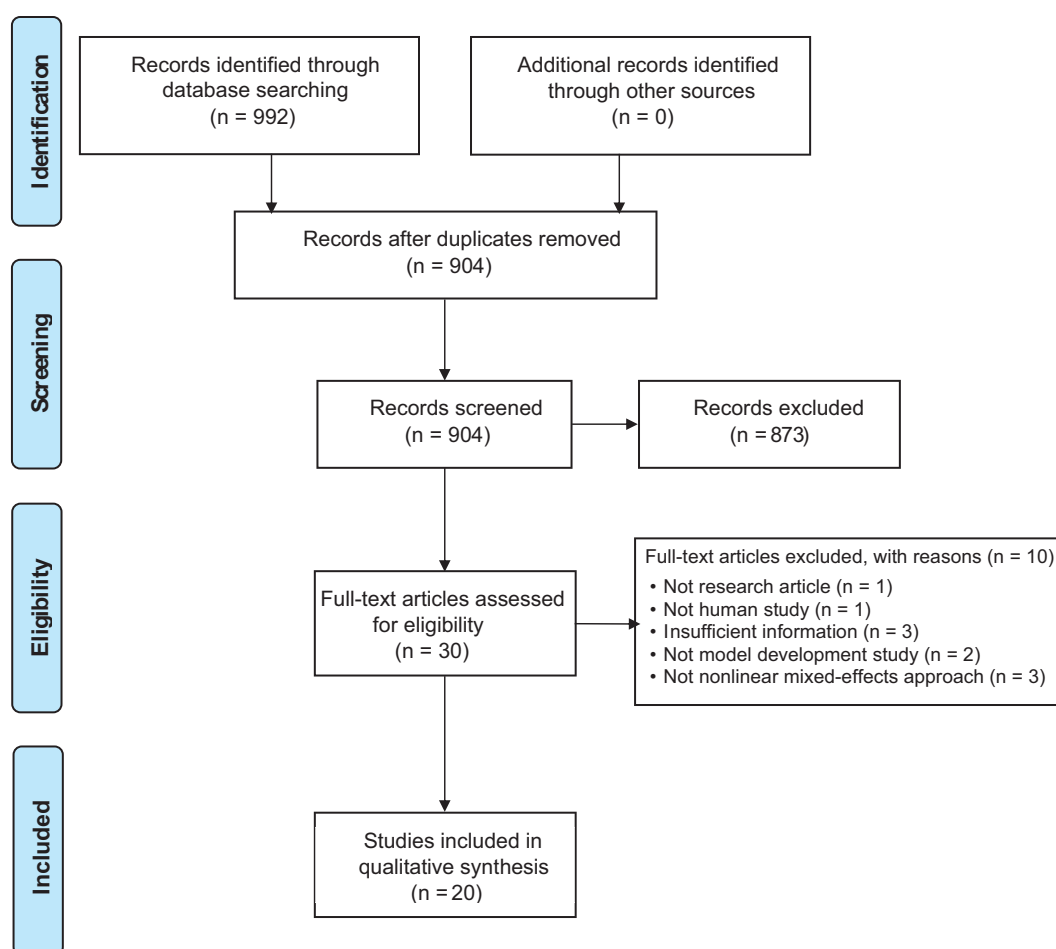


FIGURE 1. A PRISMA diagram of the study identification.

Bayesian estimator for estimating an individual's pharmacokinetic parameters.¹⁷ The sample sizes of the included studies ranged from 6 to 127 subjects, with half of the included studies retrospectively conducted using data from a clinical trial^{19–21,23,34,37} and TDM data,^{26–28,36} while the rest of the studies were prospective.^{9,15,17,18,22,24,25,29,33} Most studies used sparse sampling strategy,^{9,18,20,23,24,26–29,36} whereas six and four studies employed an intensive sampling approach^{15,17,22,25,34,35} and a mixture of both intensive and sparse sampling.^{19,21,33,37} Concerning bioassay, most studies used liquid chromatography with tandem mass spectrometry (LCMS/MS),^{17,19–21,23–25,33,34,37} mass spectrometry (MS),^{15,22,33,35} or high performance liquid chromatography with ultraviolet detector (HPLC/UV),^{18,20} whereas the rest utilized various types of immunoassays.^{9,26–29,36} Table 1 summarizes population characteristics, study design, and pharmacokinetic data.

Population Pharmacokinetic Model Development and Model Evaluation

Most studies conducted a PopPK model using NONMEM® software, except for four studies in which Phoenix NLME,^{25,29} Monolix,³⁵ and P-Pharm¹⁵ were used. Ten studies developed a model with a 1-compartment structure,^{9, 18–20,24,27–29,35,36} while nine studies used a 2-CMT disposition,^{15,17,21,22,25,26,33,34,37} and the other one that explored the developmental trajectory of CL_{SRL} employed a sigmoidal E_{max} model.²³ The first-order absorption process was used in all models, except studies by Djebli et al.¹⁷ and Wu et al.²¹ in which the Erlang distribution and the Michaelis-Menten kinetics were used to explain the absorption process. In addition, one study reported an absorption lag-time of 0.24 h.¹⁶ Eight studies^{9,19, 20,24,27–29,36} had to fix the absorption rate constant (k_a) at 0.485, 0.752, 2.2, or 2.77 h⁻¹ since the information during the absorption phase was insufficient to estimate k_a , whereas the estimated k_a ranged from 0.0535 to 2.65 h⁻¹, with the IIV ranging from 17.5% to 80.0%. The estimated CL_{SRL} without covariate effects ranged from 3.2 to 14.4 L/h, with

a wider IIV range than the k_a (11.4% to 103.0%). As for the V_d , the estimated values ranged from 88.9 to 3670 L for those with the 1-compartment structure, with the magnitude of IIV of 5.5% to 115.5%. While for the 2-compartment models, the central (V_c) and peripheral (V_p) volumes of distribution were in the range of 26.9 to 676.0 L and 72.8 to 1380 L, with the respective IIV of 7.8% to 164.0% and 10.3% to 38.7%. Concerning the statistical model, the IIV was modeled using an exponential relationship in all studies, except for one study in which an additive model was employed,¹⁵ while the proportional model was the most commonly used relationship for the RV, followed by a combined additive and proportional relationship.

Effects of numerous covariates on SRL pharmacokinetics were tested (Table 2), including age, body size (i.e., weight, height, body mass index; BMI, and body surface area; BSA), sex, SRL dose, TEM dose, duration of SRL therapy (DTT), enzyme polymorphisms, concurrent medication, postoperative days, and various laboratory measures such as hemoglobin (Hb), hematocrit (Hct), red blood cell (RBC), white blood cell (WBC), platelet, mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC), aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), albumin (ALB), total protein (TP), bilirubin (BIL), serum creatinine (SCr), creatinine clearance (CLCR), total cholesterol (TC), triglyceride (TG), high-density lipoprotein (HDL), low-density lipoprotein (LDL), and glucose. Other investigated covariates included study site, ethnicity, graft origin, tumor age, dialysis before transplant, and the presence of ischemic heart disease (IHD). Of the tested covariates, weight was the significant covariate most commonly identified, whereas *CYP3A5* polymorphisms and age were the second most commonly identified significant covariates. Other factors significantly affecting SRL pharmacokinetics were gender, BSA, height, cyclosporine concentration at time 0 h ($CsA C_0$), TEM dose, TG, TC, Hct, ALB, AST, ALT, and BIL.

TABLE 1. Population characteristics, study design, and pharmacokinetic data.

No	Author	Study design	Study site	Patient characteristics	N (male)	Mean age (range) years	Mean weight (range) kg	Mean BSA (range) m ²	Route	Sampling strategy	Bioassay
1	Ferron et al., 1997 ¹⁵	Randomized, double-blind, placebo-controlled ascending single-dose study	Multicenter	KT: high risk of chronic rejection KT: compromise kidney function KT: stable transplant	12 (11) 12 (11) 12 (9)	52.3 ± 24 51 ± 25.5 43.3 ± 29.0	73.9 ± 17.5 78.2 ± 28.7 75.6 ± 18.9	1.87 ± 10.1 1.90 ± 15.7 1.88 ± 10.0	Oral	Intensive	HPLC/MS
2	Boni et al., 2005 ³³	Randomized, double-blind, multicenter trial, once-weekly IV infusion	Multicenter	Advanced renal cell carcinoma	50 (33)	57.9 ± 9.6 (40-81)	82.9 ± 17.1 (53.7-124.7)	1.97 ± 0.2 (1.62-2.45)	IV inf. (30 min)	Intensive and Sparse	LCMS/MS
3	Djebli et al., 2006 ¹⁷	Prospective		Adult KT	22 (13)	48.6 (20-69)	Wk 1: 60 (48-90) Wk 2: 63 (48-90) Mth 1: 58 (41-90) Mth 2: 58 (41-90)		Oral	Intensive	HPLC-MS/MS
4	Sato et al., 2006 ¹⁸	Prospective	Single-center	Pancreatic islet transplant	6 (2)	Med: 39 (35-58)	Med: 57 (37-30)		Oral	Sparse	HPLC/UV
5	Zahir et al., 2006 ¹⁹	Retrospective of heart transplant trial	Multicenter	Adult heart transplant with or without ischemic heart disease	31 (24)	49.0 ± 12.0 (18-66)	78.3 ± 12.7 (51-12.5)		Oral	Intensive and Sparse	LCMS/MS
6	Jiao et al., 2009 ²⁰	Retrospective of clinical trial	Multicenter	Chinese adult KT	112 (78)	42 ± 9.9	60.4 ± 9.43	1.7 ± 0.144	Oral	Intensive	LCMS/MS and HPLC/UV
7	Wu et al., 2012 ²¹	Retrospective of clinical trial	Single-center	Patients with advanced solid tumors	76 (39)	57.7 (22-83)	79.76 (32.8-154.6)		Oral	Sparse	LCMS/MS

(continues)

TABLE 1. Continued

No	Author	Study design	Study site	Patient characteristics	N (male)	Mean age (range) years	Mean weight (range) kg	Mean BSA (range) m ²	Route	Sampling strategy	Bioassay
8	Goyal et al., 2013 ²²	Prospective	Multicenter	Pediatric blood and bone marrow transplant	37 (27)	Med: 10.1 ± 5 (4-22)	Med: 34.8 ± 19 (13.2-84.3)		Oral	Sparse	HPLC/MS
9	Shi et al., 2016 ⁹	Prospective	Single-center	Chinese adult KT	108 (79)	47±11 (21-72)	58.8 ± 5 (45-72)		Oral	Intensive	MEIA
10	Emoto et al., 2016 ²³	Retrospective of clinical trial		Neonates and infants with complicated vascular anomalies	52 (20)	Med: 4.8 (0.058-19)	Med: 18 (4-101)	Med: 0.77 (0.23-2.2)		Sparse	LCMS/MS
11	Mizuno et al., 2016 ³⁴	Retrospective of clinical trial		Children with recurrent solid tumor	19 (11)	11.9 ± 5.7 (21 days-19 years)	45.7 ± 28 (7.3-114.7)		IV inf. (15-60 min)	Intensive and Sparse	LCMS/MS
12	Mizuno et al., 2017 ²⁴	Prospective	Multicenter	Pediatric patients with vascular anomalies	52 (20)	Med:4.9 (0.1-18.6)	Med: 18.4 (4-100.6)		Oral	Intensive	LCMS/MS
13	Wang et al., 2016 ³⁷	Retrospective of bioavailability and post-market study	Multicenter	Healthy Chinese and KT patients	127 (89)	37.44±1.02 (19-64)	62±0.91 (37-89)		Oral	Sparse	LCMS/MS
14	Peng et al., 2018 ²⁵	Prospective	Single-center	Healthy Chinese subjects	27 (12)	Med: 25.48 (20-36)	Med: 61.93 (45-75)	Med: 1.71 (1.45-1.93)	Oral	Sparse	LCMS/MS/MS
15	Golubovic et al., 2019 ²⁶	Retrospective of TDM data	Single-center	Adult KT	25 (18)	43.22 ± 12.62 (16-64)	77.07 ± 18.76 (44-128)			Sparse	CMIA
16	Wang et al., 2019 ²⁷	Retrospective of TDM data	Single-center	Pediatric Chinese patients with KHE	17 (11)	1.21 ± 1.2 (0.2-6)	7.99 ± 3.04 (3.6-18)		Oral	Sparse	EMIT
17	Chen et al., 2020 ²⁸	Retrospective	Single-center	Pediatric Chinese patients with KHE	14 (9)	1.53 ± 1.40	8.87 ± 4.12		Oral	Sparse	EMIT

(continues)

TABLE 1. Continued

No	Author	Study design	Study site	Patient characteristics	N (male)	Mean age (range) years	Mean weight (range) kg	Mean BSA (range) m ²	Route	Sampling strategy	Bioassay
18	Chen et al., 2020 ²⁹	Prospective	Single-center	Children with immune cytopenia	27 (18)	8.16 ± 3.60	27.03 ± 10.87		Oral	Sparse	FPIA
19	Wang et al., 2020 ³⁶	Retrospective	Single-center	Children with tuberous sclerosis complex	15 (7)	6.16 ± 2.80	23.83 ± 8.88		Oral	Sparse	EMIT
20	Sabo et al., 2021 ³⁵	Prospective	Multicenter	Pediatric oncology	Day 1: 27 (16) Day 8: 34 (21)	11.7 ± 5.9 12.6 ± 5.6	38.5 ± 18.6 40.7 ± 17.4		Oral	Intensive	Reverse phase LCMS

BSA: body surface area, CMIA: Chemiluminescent microparticle immunoassay, EMIT: enzyme multiplied immunoassay technique, HPLC/MS: high performance liquid chromatography/mass spectrometry, HPLC-MS/MS: high performance liquid chromatography with tandem mass spectrometry, HPLC/UV: high performance liquid chromatography/ultraviolet, Inf: infusion, IV: intravenous, KHE: kaposiform hemangioendothelioma, KT: kidney transplant, LCMS/MS: liquid chromatography with tandem mass spectrometry, Med: median, MEIA: microparticle enzyme immunoassay, Mth: month, TDM: therapeutic drug monitoring, Wk: week.

As for the model evaluation, all studies used basic and advanced internal approaches; however, one study assessed the model based solely on the basic internal method,²² and the other one employed all types of model evaluation techniques.²⁶ Two studies did not have information on the model evaluation.^{15,18} Software, structural models, and model evaluation are presented in Table 3, while Table 4 summarizes the final models of the included studies and their magnitudes of IIV and RV.

Transparent Report of the Included Studies

Most studies complied with the guideline of the transparent report for clinical pharmacokinetic studies,^{31,32} with a compliance rate greater than 80%. Only one study had a compliance rate of approximately 60%,¹⁸ while four studies reported their results with compliance rates between 70% to 80%.^{16,29,36,38} The most common non-reported items of each section, identified in more than 10 studies were “route of administration” in the introduction section, “formulation details” and “sample storage” in the method section, and “study withdrawals or lost to follow-up” in the result section.

DISCUSSION

Several PopPK models of SRL have been conducted to determine factors influencing its pharmacokinetic variability. The impacts of these factors on SRL pharmacokinetics were summarized and discussed below.

Absorption

The rate of SRL absorption was variable, with the estimated k_a ranging from 0.0535 to 2.65 h⁻¹, which was consistent with a traditional pharmacokinetic study in stable transplant patients that reported a wide range of T_{max} of 0.5–3 h.³⁹ The difference in k_a among studies could not be clearly explained, however, it has been reported that administration of SRL with food results in a 3.5-fold increase in T_{max} , while the maximum concentration (C_{max}) is

TABLE 2. Tested covariates for sirolimus pharmacokinetics.

No	Author	Tested Covariates															Concurrent med	Polymorphism	Other
		Age	Body size	Sex	Blood profile	LFT	Lipid profile	Protein	Urea	BIL	RFT	SRL dose	CsA dose/ conc.						
1	Ferron et al., 1997 ¹⁵	✓	WT, HT, BSA									✓	✓					Study site	
2	Boni et al., 2005 ³³	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR			NR	NR	
3	Djebli et al., 2006 ¹⁷	✓	WT, HT, BMI, BSA		HCT, HB, RBC, WBC, platelet	AST, ALT	TC, TG	TP, ALB	✓		SCr						CYP3A5, CYP3A4, MDR1		
4	Sato et al., 2006 ¹⁸	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND			ND	ND	
5	Zahir et al., 2006 ¹⁹	✓	WT	✓	HCT, RBC		TC, TG, HDL, LDL	TP, ALB					✓					Ethnicity, IHD, POD	
6	Jiao et al., 2009 ²⁰	✓	WT, HT, BMI, BSA	✓	HCT, HB, RBC, WBC	AST, ALT	TC, TG, HDL, LDL		✓		SCr, CLCR	✓	✓				✓	Organ source, POD	
7	Wu et al., 2012 ²¹	✓	WT, HT	✓	HCT, HB, RBC, WBC, platelet	AST, ALT, ALP	TC, TG	TP, ALB	✓	✓	SCr, CLCR							Glucose	
8	Goyal et al., 2013 ²²	✓		✓	HB	AST, ALT		ALB	✓	✓	SCr								
9	Shi et al., 2016 ⁹	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR			NR	NR	
10	Emoto et al., 2016 ²³	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND	ND			ND	ND	
11	Mizuno et al., 2016 ³⁴	✓ (for TEM)		✓ (for TEM)														TEM dose	
12	Mizuno et al., 2017 ²⁴	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR	NR			NR	NR	

(Continues)

TABLE 2. Continued

No	Author	Tested Covariates												Concurrent med	Other
		Age	Body size	Sex	Blood profile	LFT	Lipid profile	Protein	Urea	BIL	RFT	SRL dose	CsA dose/conc.		
13	Wang et al., 2016 ³⁷	✓	WT, HT, BMI	✓		AST			✓	✓	SCr		✓		population
14	Peng et al., 2018 ²⁵	✓	WT, HT, BSA	✓	HCT, HB, RBC, platelet	AST, ALT	TC, TG	TP, ALB	✓		SCr		CYP3A5, MDR1		
15	Golubovic et al., 2019 ²⁶	✓	WT	✓	HCT	AST, ALT, ALP	TC, TG	TP			SCr			✓	Graft origin, dialysis before transplant
16	Wang et al., 2019 ²⁷	✓	WT	✓	HCT, HB	AST, ALT			✓		SCr	✓		✓	Duration of treatment with SRL
17	Chen et al., 2020 ²⁸	✓		✓	HCT, HB, MCH, MCHC	AST, ALT		TP, ALB		✓	SCr		CYP3A5	✓	
18	Chen et al., 2020 ²⁹	✓	WT	✓	HCT, HB, RBC, platelet	AST, ALT, ALP	TC, TG, LDL	ALB	✓	✓	SCr				
19	Wang et al., 2020 ³⁶	✓	WT	✓	HCT, HB, MCH, MCHC	AST, ALT		TP	✓	✓	SCr		ABCB1, ABCC4, ABCC8, ABCG2, CYP2C9, CYP2C19, CYP3A4, CYP3A5, CYP4F2, UGT1A1, UGT1A8, UGT2B15		
20	Sabo et al., 2021 ³⁵	✓	WT, HT, BMI, BSA											✓	Tumor age, CNS tumor diagnostic, irinotecan dose on day 1, glucose

ABC: ATP binding cassette, ALB: albumin, ALP: alkaline phosphatase, ALT: alanine aminotransferase, AST: aspartate aminotransferase, BIL: bilirubin, BMI: body mass index, BSA: body surface area, CLCR: creatinine clearance, CsA: cyclosporine, CYP: cytochrome P450, HB: hemoglobin, HCT: hematocrit, HDL: high density lipoprotein, HT: height, IHD: ischemic heart disease, LDL: low density lipoprotein, LFT: liver function test, MCH: mean corpuscular hemoglobin, MCHC: mean cell hemoglobin concentration, MDR1: multidrug resistant protein 1, ND: not determined, NR: not reported, POD: postoperative day, RBC: red blood cell, RFT: renal function test, SCr: serum creatinine, SRL: sirolimus, TC: total cholesterol, TEM: temsirolimus, TG: triglyceride, TP: total protein, UGT: Uridine 5'-Diphospho-Glucuronosyl Transferase, WBC: white blood cell, WT: weight.

TABLE 3. Software, structural models, and model evaluation.

No	Author	Software	Structural model	Model evaluation
1	Ferron et al., 1997 ¹⁵	P-Pharm	2-CMT with first-order absorption and Lag-time	NR
2	Boni et al., 2005 ³³	NONMEM	2-CMT with first-order formation into the central CMT (SRL was model as a metabolite of TEM)	Basic and advanced internal evaluation
3	Djebli et al., 2006 ¹⁷	NONMEM	2-CMT with Erlang distribution and first-order elimination	Basic and advanced internal evaluation
4	Sato et al., 2006 ¹⁸	NONMEM	1-CMT with first-order elimination	NR
5	Zahir et al., 2006 ¹⁹	NONMEM	1-CMT with first-order absorption and elimination	Basic and advanced internal evaluation
6	Jiao et al., 2009 ²⁰	NONMEM	1-CMT with first-order absorption and elimination	Basic and advanced internal evaluation
7	Wu et al., 2012 ²¹	NONMEM	2-CMT with Michaelis-Menten absorption and first-order elimination	Basic and advanced internal evaluation
8	Goyal et al., 2013 ²²	NONMEM	2-CMT with first-order absorption and elimination	Basic internal evaluation
9	Shi et al., 2016 ⁹	NONMEM	1-CMT with first-order absorption and elimination	Basic and advanced internal evaluation
10	Emoto et al., 2016 ²³	NONMEM	Sigmoidal $E_{\max}$	Basic and advanced internal evaluation
11	Mizuno et al., 2016 ³⁴	NONMEM	Temsirolimus: 3-CMT with zero-order infusion Sirolimus (as a metabolite): 2-CMT with first-order elimination	Basic and advanced internal evaluation
12	Mizuno et al., 2017 ²⁴	NONMEM	1-CMT with first-order input	Basic and advanced internal evaluation
13	Wang et al., 2016 ³⁷	NONMEM	2-CMT with first-order absorption and elimination	Basic and advanced internal evaluation
14	Peng et al., 2018 ²⁵	Phoenix NLME	2-CMT with first-order absorption and elimination	Basic and advanced internal evaluation
15	Golubovic et al., 2019 ²⁶	NONMEM	2-CMT with first-order absorption and elimination	Basic and advanced internal and external evaluation
16	Wang et al., 2019 ²⁷	NONMEM	1-CMT with first-order absorption and elimination	Basic and advanced internal evaluation
17	Chen et al., 2020 ²⁸	NONMEM	1-CMT with first-order absorption and elimination	Basic and advanced internal evaluation
18	Chen et al., 2020 ²⁹	Phoenix NLME	1-CMT with first-order absorption and elimination	Basic and advanced internal evaluation
19	Wang et al., 2020 ³⁶	NONMEM	1-CMT with first-order absorption and elimination	Basic and advanced internal evaluation
20	Sabo et al., 2021 ³⁵	Monolix	1-CMT with first-order absorption and elimination	Basic and advanced internal evaluation

CMT: compartment, NR: not reported, SRL: sirolimus, TEM: temsirolimus.

TABLE 4. Final models, interindividual variability, and residual variability.

No	Authors	Final model						RV (%CV)	
		Absorption	IIV (%CV)	Distribution	IIV (%CV)	Elimination	IIV (%CV)		
1	Ferron et al., 1997 ¹⁵	k_a (h^{-1}) = 2.18 t_{lag} (h) = 0.24	41.3% 40.1%	V_c (L) = 112.9 V_p (L) = 452	31.8% 26.4%	CL/F (L/h) = 8.91	38.2%	Prop: NR	
2	Boni et al., 2005 ³³	k_a (h^{-1}) = 0.087	34.6%	V_c (L) = 10.4 V_p (L) = 12.9*Dose ^{0.302} *HCT ^{0.719}	16.4% 22.8%	CL (L/h) = 2.05*Dose ^{0.422} Q (L/h) = 44.1	63.7% NR	Prop: 23.24%	
3	Djebl et al., 2006 ¹⁷	k_{tr} (h^{-1}) = 5.25	42.7%	V_c (L) = 218	52.7%	CL/F (L/h) = 14.1 + 14.2*CYP3A5	49.3%	Prop: 5.86% Add: 3.08 ng/mL	
4	Sato et al., 2006 ^{18,ε}	NA	NA	V_p /F (L) = 292 V/F (L) = 790 ± 659	20.2% NA	Q/F (L/h) = 38.7 CL/F (L/h) = (0.0776 + 0.167*POD/12)*WT for POD ≤ 12 CL/F (L/h) = (0.0776 + 0.167)*WT for POD > 12	78.1% NA	NA	
5	Zahir et al., 2006 ¹⁹	k_a (h^{-1}) = 0.752 (fixed)	NA (fixed)	V/F (L) = 1350	NR	CL/F (L/h) = 7.09 - 0.0147*C _{sA} dose-1.37*(1-TG) + 2.2*(1-IHD)	27.5%	Add: 24.1%	
6	Jiao et al., 2009 ²⁰	k_a (h^{-1}) = 0.752 (fixed)	NA (fixed)	V/F (L) = 3670-7.27*(C _{sA} C ₀ -104)	56.7%	CL/F (L/h) = [10.1-0.662*(TC-5.66)-0.00417*(C _{sA} C ₀ -104)] *0.65 ^{SLM} *0.661 ^{GLZ} *(DDS/2) ^{0.479}	23.8%	Expo: 29.9%	
7	Wu et al., 2012 ²¹	V_{max} (μg/L*h) = 4.56	NE	V_c /F (L) = 53.4	52.4%	CL/F (L/h) = 12.9*(35.1/HCT) ^{0.14}	52.4%	Prop: 2.17% Add: 0.5 ng/mL	
		K_m (mg) = 13.8	NE	V_p /F (L) = 611	19.3%	Q/F (L/h) = 29.0	52.4%		
8	Goyal et al., 2013 ²²	k_a (h^{-1}) = 0.0535±0.0104	NR	V_c /F (L) = 26.9±7.7	91%	CL/F (L/h) = 6.66±1.10	78%	Prop: 21% Add: 0.84 ng/mL	
				V_p /F (L) = 630±171	NR	Q/F (L/h) = 4.62±2	NR		

(continues)

TABLE 4. Continued

No	Authors	Final model					RV (%CV)		
		Absorption	IIV (%CV)	Distribution	IIV (%CV)	Elimination	IIV (%CV)		
9	Shi et al., 2016 ⁹	k_a (h^{-1}) = 2.20 (fixed)	NA (fixed)	V_d/F (L) = 322	22.6	CL/F (L/h) = $14.4 \cdot (1+W/T/58.6 \cdot 0.19) \cdot \exp((-ALB/38.9) \cdot 0.26) \cdot \exp(CYP3A5 \cdot -0.30)$	19.6	Expo: 25.6%	
10	Emoto et al., 2016 ²³					$CL = CL_{matured} \cdot PMA^{Hill}/+$	11.3%	Prop: 25.7%	
						$CL_{matured}$ (L/h/70 kg) = 18.7			
						TM_{50} (weeks) = 62.9	17.4%		
11	Mizuno et al., 2016 ³⁴	CL_{TEM} (L/h/70 kg) = $3.4 \cdot Dose^{0.855}$	70.9%	V_c (L/h/70kg) = 48	121%	CL_{SRL} (L/h/70kg) = 6.08	103%	Prop: 25.5%	Add: 1.69 ng/mL
				V_p (L/h/70kg) = 72.8	NE	Q (L/h/70kg) = 11.6	NE		
12	Mizuno et al., 2017 ²⁴	k_a (h^{-1}) = 2.77 (fixed)	NA (fixed)	$V_c^{pediatric} = V_{adult} \cdot (BW/70)$ $V_{adult} = 1030$	62.3%	$CL_{pediatric} = CL_{adult} \cdot (BW/70)^{0.73} \cdot MF$ $MF = PMA^{Hill}/(1 + PMA^{Hill})$ $CL_{adult} = 18.5$	31.2%	Prop: 38.1%	
				V_p/F (L) = $676 \cdot (SCR/592.3)^{1.4}$	10.3%	CL/F (L/h) = $8.81 \cdot [1 - 0.219 \cdot (CsA/300)] \cdot [1 - 0.0171 \cdot (age - 40)]$	50.9%	Prop: 62.2%	
13	Wang et al., 2016 ³⁷	k_a (h^{-1}) = 0.24	0	V_p/F (L) = 1380	103.4%	Q/F (L/h) = 32.9	12.7%		
				$V_c = 184.461$ for female $V_c = 184.461 \cdot \exp(0.266)$ for male	7.8	$CL = 10.813 \cdot (CYP3A5 \cdot 1)^{1.1}$ $CL = 10.813 \cdot \exp(0.034) \cdot (CYP3A5 \cdot 1)^{1.3}$ $CL = 10.813 \cdot \exp(0.250) \cdot (CYP3A5 \cdot 1)^{1.3}$	11.4%	Prop: 17.5%	
14	Peng et al., 2018 ²⁵	k_a (h^{-1}) = 2.651		$V_p = 170.029 \cdot BSA^{2.165}$	38.7	Q (L/h) = 23.596	7.6%		
				V_c/F (L) = 118	55.3	CL/F (L/h) = $12.2 \cdot 0.63^{AST} \cdot (1 - (age/44) \cdot 0.388)$ $AST = 0$ if ≤ 37 IU/L $AST = 1$ if > 37 IU/L	23.4%	Prop: 24.9%	Add: 1.93 ng/mL
15	Golubovic et al., 2019 ²⁶	k_a (h^{-1}) = 2.19	38.1%	V_p/F (L) = 609	25.6	Q/F (L/h) = 5.07	32.1%		

(continues)

TABLE 4. Continued

No	Authors	Final model					RV (%CV)	
		Absorption	IIV (%CV)	Distribution	IIV (%CV)	Elimination	IIV (%CV)	
16	Wang et al., 2019 ²⁷	k_a (h^{-1}) = 0.485 (fixed)	NA (fixed)	V/F (L) = $165 * \exp(0.0783 * DTT/10)$	115.5	CL/F (L/h) = $3.19 * \exp(0.215 * \text{age}) * \exp(0.0108 * ALT)^* \exp(-0.818 * \text{sex})$ Sex = 1 for female, 0 otherwise	46.6%	Expo: 60.4%
17	Chen et al., 2020 ²⁸	k_a (h^{-1}) = 0.485 (fixed)	NA (fixed)	V/F (L) = $1840 * (WT/70)$	NR	CL/F (L/h) = $7.55 * (WT/70)^{0.75} * (1 - (0.999)^{CYP3A5})$	59.0%	Prop: 62.4%
18	Chen et al., 2020 ²⁹	k_a (h^{-1}) = 0.752 (fixed)	NA (fixed)	V/F (L) = 144.16	42.9%	CL/F (L/h) = $5.63 * (TBIL/11.29)^{-0.32} * (WT/28.5)^{0.5}$	21.9%	NR: 11.8%
19	Wang et al., 2020 ³⁶	k_a (h^{-1}) = 0.485 (fixed)	NA (fixed)	V/F (L) = $124 * (WT/70)$	5.5%	CL/F (L/h) = $6.48 * (WT/70)^{0.75}$	25.7%	Prop: 55.9% Add: 1.25 ng/mL
20	Sabo et al., 2021 ³⁵	For day 1: k_a (h^{-1}) = 0.46	80%	For day 1: V/F (L/h) = $88.9 * (BSAi/Med BSA)^{1.35}$	62.5%	For day 1: CL/F (L/h) = $23.9 * (BSAi/Med BSA)$	92.9%	Prop: Day 1: 0.54%
		For day 8: k_a (h^{-1}) = 0.97	168.5%	For day 8: V/F (L/h) = $238 * (BSAi/Med BSA)^{1.41}$	29.4%	For day 8: CL/F (L/h) = $11.9 * (BSAi/Med BSA)^{1.09}$	50.1%	Prop: Day 8: 0.31%

^ε This study compared two models and did not determine the magnitude and sources of variability

ALB: albumin, AST: aspartate aminotransferase, BSA: body surface area, BSAi: individual body surface area, CL : clearance, $CL_{matured}$: clearance at fully matured level, $C_{sA} C_o$: cyclosporine trough concentration, C_{sA}: cyclosporine, CYP: cyclosporine, P450, DDS: daily dose sirolimus, F: bioavailability, GLZ: glycyrrhizin, HCT: hematocrit, IHD: ischemic heart disease, k_a : absorption rate constant, k_m : sirolimus amount at 50% of V_{max} , k_{tr} : transfer rate constant, Med: median, MF: maturation function, NA: not applicable, NE: Not estimated, PMA: postmenstrual age, POD: postoperative day, Q: intercompartmental clearance, SLM: silymarin, SRL: sirolimus, TBIL: total bilirubin, TC: total cholesterol, TEM: tensirolimus, TG: triglyceride, t_{lag} : absorption lag time, TM: postmenstrual age at which clearance is half of $CL_{matured}$, V_c : central volume of distribution, V_{max} : maximum absorption rate, V_p : peripheral volume of distribution, WT: weight.

decreased by 34%.¹ The administration process of some of the included studies, e.g., fasted or fed, was not described and might contribute to such difference. Though the magnitude of IIV on k_a ranged from 17.5% to 80.0%, no studies identified significant predictors for k_a , and this was consistent with a study by Kahan et al. that reported no association between sex, age, weight, or ethnicity and C_{max} , the minimum concentration at steady-state ($C_{min,ss}$), or AUC.¹⁴ Notably, the slowest k_a of 0.0535 h⁻¹ was reported by a study conducted in pediatric bone marrow transplant patients whose ages ranged from 4 years to 22 years, and some patients were co-administered fluconazole.²² It has been shown that neonates and infants have longer gastric emptying, which can delay the rate of drug absorption, however, the age at which gastric emptying time approaches that in adults was not specifically determined.⁴⁰

Distribution

The estimated V_d for the 1-compartment studies ranged from 88.9 L to 1840 L, excluding the one with a substantially high value of 3670 L, while those of the 2-compartment studies had the V_d of 120.8 L to 2056 L. The large V_d of SRL can be explained by its lipophilic property, which contributes to the distribution of the lipid membrane of various tissues.⁵ The ranges of V_d from SRL PopPK studies were more comprehensive than those of the traditional pharmacokinetics conducted in stable renal transplant patients (392 L to 1169 L for a 70 kg patient),⁸ which could be due to different patients' characteristics. PopPK studies that reported the low V_d values (88.9 L to 165 L) were conducted in children,^{27,29,35,36} and evidence has indicated that children contain lower fat mass than adults.⁴¹

Significant predictors for SRL V_d included CsA C_0 ,²⁰ weight,^{24,28,36} BSA,³⁵ Scr,³⁷ sex,²⁵ and DTT.²⁷ Jiao et al.²⁰ indicated that 1 ng/mL increase in CsA C_0 from the median value of 104 ng/mL resulted in a decrease in the apparent volume of distribution (V_d/F) of 7.27 L. CsA is a substrate and an inhibitor

of intestinal CYP3A4 and P-gp,⁴² thus co-administration of CsA and SRL increased SRL bioavailability, and in turn, a decrease in V_d/F .

The effect of weight on V_d was explained using an allometric scaling relationship with the exponent of one, suggesting that the V_d is linearly related to weight. This relationship is well accepted since it is based on physiologic principles describing size in relation to blood volume and vital capacity.⁴³ Another study; however, used BSA instead of weight as an index of body size with the exponent of 1.35 and 1.09 for V_d on day 1 and V_d on day 8.³⁵ This was deemed feasible since this study was conducted in pediatric patients with solid tumors in which SRL dose was titrated based on BSA.

Wang et al. reported a nonlinear increase in V_c/F with an increase in SCr.³⁷ This could be rationalized by a decrease in plasma protein binding in patients with impaired renal function,⁴⁴ increasing free SRL concentrations that can distribute to red blood cells. Moreover, Peng et al. reported higher V_c of SRL in males than in females, which was expected given that males generally have larger body configurations than females.²⁵ As for the DTT, V_d of SRL increased as the DTT increased, which could be due to an increase in the number of erythrocytes with the improvement of clinical outcomes as the duration of treatment was lengthened.²⁷

Elimination

Significant predictors for CL_{SRL} were different among studies. Jiao et al.²⁰ reported a nonlinear increase in CL_{SRL}/F with an increase in SRL dose. However, this could be due to the TDM effects since subjects with higher CL_{SRL}/F tend to receive higher SRL doses. This effect was previously described by Ahn et al.⁴⁵ Boni et al.³³ also reported a similar effect using a model conducted in patients with advanced renal cancer receiving TEM, in which CL_{SRL} increased with an increase in TEM dose.

Several studies found a significant effect of CsA on CL_{SRL} . Zahir et al.¹⁹ identified that a 100 mg increase in CsA dose led to approximately 20.7%

decrease in CL_{SRL}/F , while Wang et al.³⁷ reported a smaller effect of a 7.3% decrease in CL_{SRL}/F . Moreover, Jiao et al.²⁰ found a 4.5% decrease in CL_{SRL}/F for a 100 ng/mL increase in CsA C_0 . CsA is an inhibitor of CYP3A4 and P-gp. Thus concomitant administration of CsA and SRL leads to an increase in SRL bioavailability and in turn, a decrease in CL_{SRL}/F .⁴² Therefore, in patients undergoing CsA dose minimization, the SRL dose should be increased by 7% to 20% for a 100 mg decrease in the CsA dose.

Significant effects of several laboratory values were identified. First, an increase in Hct contributed to a modest decrease in CL_{SRL}/F .²¹ This effect was previously reported for tacrolimus.⁴⁶ Since SRL is concentrated in erythrocytes,⁷ as Hct increases, free SRL levels available for elimination decrease. Second, CL_{SRL}/F is decreased as TC²⁰ or TG increases.¹⁹ This could be due to increased SRL bioavailability following high-fat meals since patients with hyperlipidemia tend to consume high-fat meals.^{19,20} Zahir et al.¹⁹ also proposed that SRL may be a low intrinsic clearance drug in which hepatic clearance depends on fraction unbound. Patients with hyperlipidemia may have a lower unbound SRL fraction since the drug extensively distributes across the plasma membrane and binds to erythrocytes, decreasing hepatic clearance. The authors also reported that heart transplant patients with non-IHD had lower CL_{SRL}/F than IHD patients, which might also be associated with dyslipidemia since IHD patients tend to consume a high-fat diet. Nonetheless, this covariate (IHD) can be confounded by the effect of TG and should be interpreted with a caveat. Third, a one-fold of ALB lower than average level contributed to a modest increase in CL_{SRL}/F of 17%.⁹ This would be expected since it increased the free SRL fraction available for elimination. Fourth, and impaired liver function, expressed as AST above upper limit normal contributed to lower CL_{SRL}/F ,²⁶ which is not surprising given that SRL is extensively metabolized by the liver. Fifth, Cheng et al. reported that the higher BIL was associated

with, the lower CL_{SRL}/F ; however, underlying mechanism of this effect could not be clarified.²⁹

Several studies identified a significant association between weight and CL_{SRL}/F , with an increase in CL_{SRL}/F with bodyweight.^{9,24,28,29,36} This association is commonly described using a power relationship⁴³ and is widely applied since higher body weight may relate to larger elimination organs. Whereas Sabo et al.³⁵ reported a significant association between BSA and CL_{SRL}/F in pediatric patients with solid tumors using a power relationship, which is deemed appropriate given that SRL dose is given based on BSA in patients with cancer. Moreover, one study reported that females had approximately 40% lower CL_{SRL}/F than males,²⁷ which is incongruent with a physiological basis that females have lower body weight, corresponding to smaller elimination organs.

Golubovic et al. identified that CL_{SRL}/F of adults decreased with advancing age,²⁶ which is in agreement with physiological basis, while Wang et al.²⁷ reported an increase in CL_{SRL}/F with age in a pediatric population aged 0.2-6 years. This was expected based on the development of elimination organs that approaches adults with increasing age. Moreover, Emoto et al.²³ described the developmental trajectory of CL_{SRL} in neonates and infants using postmenstrual age (PMA) and a sigmoidal E_{max} model, which could aid dosing recommendations in this population. Based on their model, CL_{SRL} approached the mature level at the PMA of approximately 144-196 weeks.

CYP3A5 polymorphisms significantly influence CL_{SRL}/F , as Djebli et al.¹⁷ and Chen et al.²⁸ found that non-expressers (*CYP3A5**3/*3) had approximately 50% lower CL_{SRL}/F than expressers (*CYP3A5**1/*1 and *CYP3A5**1/*3). With a similar trend, Shi et al.⁹ and Peng et al.²⁵ reported that patients carrying *CYP3A5**1/*1, *CYP3A5**1/*3, *CYP3A5**3/*3 had a ratio of CL_{SRL}/F of 1: 0.74: 0.55 and 1: 0.96: 0.78, respectively. This effect could assist SRL dosing when *CYP3A5* genotyping is available.

In conclusion, PopPK models of SRL conducted using a nonlinear mixed-effects approach

were summarized, and significant predictors for CL_{SRL} were identified. These models, with Bayesian forecasting, can be used to guide SRL dosage individualization. However, the choice of model selection should be based on the characteristics of the target population in which the model is to be used. Moreover, most models were not externally evaluated. Therefore, the predictive performance of such models should be assessed before applying them in clinical practice.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

AUTHORSHIP CONTRIBUTIONS

Conceptualizations: Janthima Methaneethorn. Data screening and extraction: Janthima Methaneethorn, Premasuda Art-arsa, and Ramanya Kosiyaporn. Quality assessment: Janthima Methaneethorn, Premasuda Art-arsa, Ramanya Kosiyaporn, and Nattawut Leelakanok. Drafting the manuscript: Janthima Methaneethorn. Reviewing and approving the manuscript: Janthima Methaneethorn, Premasuda Art-arsa, Ramanya Kosiyaporn, and Nattawut Leelakanok.

REFERENCES

1. Rapamune [package insert]. Philadelphia, PA: Wyeth-Ayerst Pharmaceuticals; 2017.
2. Rini BI. Temsirolimus, an inhibitor of mammalian target of rapamycin. *Clin Cancer Res*. 2008;14(5):1286–90. <https://doi.org/10.1158/1078-0432.CCR-07-4719>
3. Stenton SB, Partovi N, Ensom MH. Sirolimus. *Clin Pharmacokinet*. 2005;44(8):769–86. <https://doi.org/10.2165/00003088-200544080-00001>
4. MacDonald A, Scarola J, Burke JT, Zimmerman JJ. Clinical pharmacokinetics and therapeutic drug monitoring of sirolimus. *Clin Ther*. 2000;22:B101–B21. [https://doi.org/10.1016/s0149-2918\(00\)89027-x](https://doi.org/10.1016/s0149-2918(00)89027-x)
5. Mahalati K, Kahan BD. Clinical pharmacokinetics of sirolimus. *Clin Pharmacokinet*. 2001;40(8):573–85. <https://doi.org/10.2165/00003088-200140080-00002>
6. Yatscoff R. Pharmacokinetics of rapamycin. *Transplant Proc*. 1996;28(2):970–3.
7. Yatscoff R, LeGatt D, Keenan R, Chackowsky P. Blood distribution of rapamycin. *Transplantation*. 1993;56(5):1202–6. <https://doi.org/10.1097/00007890-199311000-00029>
8. Zimmerman JJ, Kahan BD. Pharmacokinetics of sirolimus in stable renal transplant patients after multiple oral dose administration. *J Clin Pharmacol*. 1997;37(5):405–15. <https://doi.org/10.1002/j.1552-4604.1997.tb04318.x>
9. Shi HQ, Yang J, Zhang LQ, et al. The population pharmacokinetics of sirolimus and CYP3A5*3 polymorphism in Chinese renal transplant patients. *Int J Clin Exp Med*. 2016;9(3):5854–66. <https://doi.org/10.1111/j.1365-2125.2009.03392.x>
10. Fryer J, Yatscoff RW, Pascoe EA, et al. The relationship of blood concentrations of rapamycin and cyclosporine to suppression of allograft rejection in a rabbit heterotopic heart transplant model. *Transplantation*. 1993;55(2):340–5. <https://doi.org/10.1097/00007890-199302000-00021>
11. Granger DK, Cromwell JW, Chen SC, et al. Prolongation of renal allograft survival in a large animal model by oral rapamycin monotherapy. *Transplantation*. 1995;59(2):183–6.
12. Kahan BD, Julian BA, Pescovitz MD, et al. Sirolimus reduces the incidence of acute rejection episodes despite lower cyclosporine doses in Caucasian recipients of mismatched primary renal allografts: a phase II trial. *Transplantation*. 1999;68(10):1526–32. <https://doi.org/10.1097/00007890-199911270-00016>
13. Kahan BD, Podbielski J, Napoli KL, et al. Immunosuppressive effects and safety of a sirolimus/cyclosporine combination regimen for renal transplantation. *Transplantation*. 1998;66(8):1040–6. <https://doi.org/10.1097/00007890-199810270-00013>
14. Kahan B, Napoli K, Kelly P, et al. Therapeutic drug monitoring of sirolimus: correlations with efficacy and toxicity. *Clin Transplant*. 2000;14(2):97–109. <https://doi.org/10.1034/j.1399-0012.2000.140201.x>

15. Ferron GM, Mishina EV, Zimmerman JJ, et al. Population pharmacokinetics of sirolimus in kidney transplant patients. *Clin Pharmacol Ther.* 1997;61(4):416–28. [https://doi.org/10.1016/S0009-9236\(97\)90192-2](https://doi.org/10.1016/S0009-9236(97)90192-2)
16. Ferron GM, Mishina EV, Jusko WJ, et al. Population pharmacokinetics of sirolimus. *Clin Pharmacol Ther.* 1998;63(4):494. <https://doi.org/10.1111/j.1365-2125.2009.03392.x>
17. Djebli N, Rousseau A, Hoizey G, et al. Sirolimus population pharmacokinetic/pharmacogenetic analysis and bayesian modelling in kidney transplant recipients. *Clin Pharmacokinet.* 2006;45(11):1135–48. <https://doi.org/10.2165/00003088-200645110-00007>
18. Sato E, Shimomura M, Masuda S, et al. Temporal decline in sirolimus elimination immediately after pancreatic islet transplantation. *Drug Metab Pharmacokinet.* 2006;21(6):492–500.
19. Zahir H, Keogh AM, Akhlaghi F. Apparent clearance of sirolimus in heart transplant recipients: impact of primary diagnosis and serum lipids. *Ther Drug Monit.* 2006;28(5):614–22. <https://doi.org/10.1097/01.ftd.0000246765.05248.fa>
20. Jiao Z, Shi XJ, Li ZD, et al. Population pharmacokinetics of sirolimus in de novo Chinese adult renal transplant patients. *Br J Clin Pharmacol.* 2009;68(1):47–60. <https://doi.org/10.1111/j.1365-2125.2009.03392.x>
21. Wu K, Cohen EE, House LK, et al. Nonlinear population pharmacokinetics of sirolimus in patients with advanced cancer. *CPT: Pharmacomet Syst Pharmacol.* 2012;1(12):e17. <https://doi.org/10.1038/psp.2012.18>
22. Goyal RK, Han K, Wall DA, et al. Sirolimus pharmacokinetics in early postmyeloablative pediatric blood and marrow transplantation. *Biol Blood Marrow Transplant.* 2013;19(4):569–75. <https://doi.org/10.1016/j.bbmt.2012.12.015>
23. Emoto C, Fukuda T, Mizuno T, et al. Characterizing the developmental trajectory of sirolimus clearance in neonates and infants. *CPT: Pharmacomet Syst Pharmacol.* 2016;5(8):411–7. <https://doi.org/10.1002/psp4.12096>
24. Mizuno T, Emoto C, Fukuda T, et al. Model-based precision dosing of sirolimus in pediatric patients with vascular anomalies. *Eur J Pharm Sci.* 2017;109s:s124–s31. <https://doi.org/10.1016/j.ejps.2017.05.037>
25. Peng M, Zhao G, Li X, et al. Population pharmacokinetics of sirolimus in healthy Chinese subjects. *Indian J Pharm Sci.* 2018;80(2):291–7. <https://doi.org/10.4172/pharmaceutical-sciences.1000357>
26. Golubović B, Vučićević K, Radivojević D, et al. Exploring sirolimus pharmacokinetic variability using data available from the routine clinical care of renal transplant patients - population pharmacokinetic approach. *J Med Biochem.* 2019;38(3):323–31. <https://doi.org/10.2478/jomb-2018-0030>
27. Wang D, Chen X, Li Z. Population pharmacokinetics of sirolimus in pediatric patients with kaposiform hemangioendothelioma: a retrospective study. *Oncol Lett.* 2019;18(3):2412–9. <https://doi.org/10.3892/ol.2019.10562>
28. Chen X, Wang DD, Xu H, et al. Initial dose recommendation for sirolimus in paediatric kaposiform haemangioendothelioma patients based on population pharmacokinetics and pharmacogenomics. *J Int Med Res.* 2020;48(8):300060520947627. <https://doi.org/10.1177/0300060520947627>
29. Cheng X, Zhao Y, Gu H, et al. The first study in pediatric: population pharmacokinetics of sirolimus and its application in Chinese children with immune cytopenia. *Int J Immunopathol Pharmacol.* 2020;34:2058738420934936. <https://doi.org/10.1177/2058738420934936>
30. Brendel K, Dartois C, Comets E, et al. Are population pharmacokinetic and/or pharmacodynamic models adequately evaluated? *Clin Pharmacokinet.* 2007;46(3):221–34. <https://doi.org/10.2165/00003088-200746030-00003>
31. Kanji S, Hayes M, Ling A, et al. Reporting guidelines for clinical pharmacokinetic studies: the ClinPK statement. *Clin Pharmacokinet.* 2015;54(7):783–95. <https://doi.org/10.1007/s40262-015-0236-8>
32. Dartois C, Brendel K, Comets E, et al. Overview of model-building strategies in population PK/PD analyses: 2002–2004 literature survey. *Br J Clin Pharmacol.* 2007;64(5):603–12. <https://doi.org/10.1111/j.1365-2125.2007.02975.x>
33. Boni JP, Leister C, Bender G, et al. Population pharmacokinetics of CCI-779: correlations to

- safety and pharmacogenomic responses in patients with advanced renal cancer. *Clin Pharmacol Ther.* 2005;77(1):76–89.
34. Mizuno T, Fukuda T, Christians U, et al. Population pharmacokinetics of temsirolimus and sirolimus in children with recurrent solid tumours: a report from the Children's oncology group. *Br J Clin Pharmacol.* 2017;83(5):1097–107. <https://doi.org/10.1111/bcp.13181>
35. Sabo AN, Jannier S, Becker G, et al. Sirolimus pharmacokinetics variability points to the relevance of therapeutic drug monitoring in pediatric oncology. *Pharmaceutics.* 2021;13(4):470. <https://doi.org/10.3390/pharmaceutics13040470>
36. Wang DD, Chen X, Xu H, et al. Initial dosage recommendation for sirolimus in children with tuberous sclerosis complex. *Front Pharmacol.* 2020;11:890. <https://doi.org/10.3389/fphar.2020.00890>
37. Wang M, Bao ling D, Yin-jin Y, et al. Population pharmacokinetic characteristics of sirolimus in healthy Chinese subjects and renal transplant patients. *Int J Clin Pharmacol Ther.* 2016;54(6):433. <https://doi.org/10.5414/cp202499>
38. Boni JP, Zhou S, Burns J, et al. 552 POSTER integrated population pharmacokinetic analysis of temsirolimus in cancer patients following weekly IV treatments. *Eur J Cancer Suppl.* 2006;4(12):167–8. <https://doi.org/10.1053/j.seminoncol.2009.10.009>
39. Johnson EM, Zimmerman J, Duderstadt K, et al. A randomized, double-blind, placebo-controlled study of the safety, tolerance, and preliminary pharmacokinetics of ascending single doses of orally administered sirolimus (rapamycin) in stable renal transplant recipients. *Transplant Proc.* 1996;28(2):987.
40. Benedetti MS, Whomsley R, Baltes EL. Differences in absorption, distribution, metabolism and excretion of xenobiotics between the paediatric and adult populations. *Expert Opin Drug Metab Toxicol.* 2005;1(3):447–71. <https://doi.org/10.1517/17425255.1.3.447>
41. Weber DR, Leonard MB, Zemel BS. Body composition analysis in the pediatric population. *Pediatr Endocrinol Rev.* 2012;10(1):130–9.
42. Zimmerman JJ, Harper D, Getsy J, et al. Pharmacokinetic interactions between sirolimus and microemulsion cyclosporine when orally administered jointly and 4 hours apart in healthy volunteers. *J Clin Pharmacol.* 2003;43(10):1168–76. <https://doi.org/10.1177/0091270003257227>
43. Anderson BJ, Holford NH. Mechanism-based concepts of size and maturity in pharmacokinetics. *Annu Rev Pharmacol Toxicol.* 2008;48:303–32. <https://doi.org/10.1146/annurev.pharmtox.48.113006.094708>
44. Gibaldi M. Drug distribution in renal failure. *Am J Med.* 1977;62(4):471–4. [https://doi.org/10.1016/0002-9343\(77\)90399-0](https://doi.org/10.1016/0002-9343(77)90399-0)
45. Ahn JE, Birnbaum AK, Brundage RC. Inherent correlation between dose and clearance in therapeutic drug monitoring settings: possible misinterpretation in population pharmacokinetic analyses. *J Pharmacokinet Pharmacodyn.* 2005;32(5-6):703–18. <https://doi.org/10.1007/s10928-005-0083-6>
46. Staatz CE, Willis C, Taylor PJ, et al. Population pharmacokinetics of tacrolimus in adult kidney transplant recipients. *Clin Pharmacol Ther.* 2002;72(6):660–9. <https://doi.org/10.1067/mcp.2002.129304>