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ROLE OF PATHOGENS IN DIABETIC FOOT INFECTION AND THE POTENTIAL OF IMMUNOPROTEOMICS AS A DIAGNOSTIC AND PROGNOSTIC TOOL

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Abstract

Introduction: Diabetic foot infection (DFI) stands as a prevalent and debilitating complication of diabetes mellitus, posing significant challenges to patients' quality of life and healthcare systems worldwide.

Objectives: The main objective of the study is to find the role of pathogens in diabetic foot infection (DFI) and the potential of immunoproteomics as a diagnostic and prognostic tool.

Material and methods: This prospective observational study was conducted at Rashid Latif Medical and Dental College, Lahore from June 2021 to November 2021. Data were collected from 550 diabetic patients suffering from DFI. Data were collected through a systematically designed questionnaire which included clinical evaluation, including medical history assessment, physical examination, and wound assessment.

Results: Data were collected from 550 patients from both genders. There were 60% male and 40% female. Mean age of patients was 65.09 ± 8.3 years and the mean duration of diabetes was 11.98 ± 6.51 years. 70% of the patients also suffering from hypertension, 45% Peripheral Neuropathy and 30% from Peripheral Vascular Disease. Staphylococcus aureus, both methicillin-sensitive (MSSA) and methicillin-resistant (MRSA) strains, are the most prevalent pathogens, with MRSA exhibiting a resistance rate of 30%. Enterococcus spp., Escherichia coli, and Pseudomonas aeruginosa are also prevalent, with Escherichia coli showing notable resistance rates of 60% to ciprofloxacin and 40% to gentamicin. Elevated levels of IL-6 and TNF-alpha show positive correlations with wound size (IL-6: r = 0.65, p < 0.001; TNF-alpha: r = 0.55, p = 0.005), tissue depth involvement (IL-6: r = 0.50, p = 0.002; TNF-alpha: r = 0.45, p = 0.015), and amputation rate (IL-6: r = 0.75, p < 0.001; TNF-alpha: r = 0.70, p < 0.001).

Conclusion: It is concluded that the role of pathogens and host immune response in diabetic foot infections (DFI) is important for guiding effective treatment strategies.

Introduction

Diabetic foot infection (DFI) stands as a prevalent and debilitating complication of diabetes mellitus, posing significant challenges to patients' quality of life and healthcare systems worldwide. Characterized by microbial invasion of soft tissue and bone, DFIs frequently lead to severe complications, including amputation and mortality, particularly in cases of delayed diagnosis and inadequate treatment [1]. Pathogens play a pivotal role in the pathogenesis and progression of DFIs, with polymicrobial infections being common and challenging to manage. Early and accurate identification of the infecting pathogens is crucial for guiding appropriate antimicrobial therapy and improving patient outcomes [2].

Diabetic foot ulcer (DFU) is one of the most common problems clinicians have to deal within patients with diabetes mellitus (DM). The incidence varies between 15% and 25%, and about 1% of this population has to undergo a lower limb amputation [3]. The high incidence of severe complications and the increased rates of morbidity and mortality, prompt for early diagnosis and initiation of appropriate antibiotic treatment to improve final outcomes. Infection complicates approximately 60% of DFUs [4]. The initial soft tissue infection may spread into the bone resulting in diabetic foot osteomyelitis and thus a high risk of amputation. Osteomyelitis should be suspected in all DFU patients with clinical findings of infection and in chronic or recurrent wounds. Early identification of this clinical entity is crucial for the overall management and in order to reduce mortality [5].

DM patients with deep foot infections are 154.5 times more likely to have a leg, foot, or toe amputated compared with patients without DM. The infection is usually complicated to diabetic foot ulcer (DFU) initiated by an open wound on the foot caused by injury, ischemic, or tinea pedis [6]. The weakened immune system, impaired peripheral blood circulation, neuropathy, and peripheral vasculopathy facilitate DFI. The pathogens of DFI include aerobic bacteria such as Staphylococcus, Streptococcus, and Enterobacteriaceae, and anaerobic bacteria such as Bacteroides, Clostridium, and Peptostreptococcus, and fungi [7]. According to guidelines compiled by the Infectious Diseases Society of America, DFIs are classified into three subcategories, namely mild infections with only superficial symptoms that are limited in size and depth, moderate infections with deeper or more extensive symptoms, and severe infections accompanied by systemic signs or metabolic perturbations [8]. For the treatment of mild and moderate DFIs, oral therapy alone or followed by a short course of intravenous therapy with narrow-spectrum antibiotics is likely sufficient. Severe DFI is often associated with previously treated chronic infection, and possibly with antibiotic resistance. The initial approach for severe DFI treatment is parenteral administration of broad-spectrum antibiotics, minimally those against Staphylococcus and Streptococcus [9].

In recent years, immunoproteomics has emerged as a promising approach for the diagnosis and prognostication of infectious diseases, including DFIs. Immunoproteomics involves the comprehensive analysis of the immune response to pathogenic antigens, enabling the identification of specific biomarkers associated with infection [10]. By interrogating the host's immune repertoire, immunoproteomics offers insights into the complex interplay between pathogens and the host immune system, facilitating the development of novel diagnostic and prognostic tools for infectious diseases [11].

It is estimated that 19 to 34% of patients with T2D develop DFU in their lifetimes, and DFU is the most common cause of hospitalization and medical costs associated with diabetes. Despite high health care costs, outcomes for patients presenting with DFU infections are poor; such infections often result in lower-limb amputation, with very poor 1-, 2-, and 5-year survival rates of 80.80%, 69.01%, and 28.64%, respectively [12]. DFU is caused by a combination of peripheral sensorimotor and autonomic neuropathy, peripheral vascular disease, and minor trauma, frequently complicated by subsequent infections. Several metagenomic studies demonstrated that Staphylococcus aureus is the most common pathogen isolated from DFU infections. Furthermore, microbiome studies demonstrated that patients with T2D showed skin microbiota more frequently colonized with S. aureus and more susceptible to S. aureus infections [13].

Objectives

The main objective of the study is to find the role of pathogens in diabetic foot infection (DFI) and the potential of immunoproteomics as a diagnostic and prognostic tool.

Material and methods

This prospective observational study was conducted at Rashid Latif Medical and Dental College, Lahore from June 2021 to November 2021. Data were collected from 550 diabetic patients suffering from DFI.

Inclusion criteria

- Patients aged > 18 years
- Patients with confirm diagnosis of DFI.
- Those who are willing to participate in the study.

Exclusion criteria

- Patients with any immunodeficiency disorder.
- Patients with any inflammatory condition and using antibiotics from last two weeks.

Data collection

Data were collected from 550 diabetic patients suffering from diabetic foot infection. Data were collected through a systematically designed questionnaire which include clinical evaluation, including medical history assessment, physical examination, and wound assessment. Relevant clinical data, including demographic information, diabetes-related variables, duration of diabetes, glycemic control, and details of foot ulcer characteristics were also noted.

Microbiological Analysis: Wound swabs were collected from all patients for microbiological analysis. Standard microbiological techniques, including aerobic and anaerobic culture, were employed to identify the causative pathogens. Antimicrobial susceptibility testing was performed according to established guidelines.

Immunoproteomic Analysis: Blood samples were collected from a set of patients (n=100) for immunoproteomic analysis. Blood was centrifuged at 3000rpm for 15 mins for serum separation. Peripheral blood mononuclear cells (PBMCs) were isolated, and serum samples were used for analysis of the host immune response to DFI-associated pathogens. Enzyme-linked immunosorbent assay (ELISA) were employed to identify specific antibodies and immune markers indicative of DFI.

Data analysis

Data were analyzed using SPSS v29.0 and GraphPad prism 2021. The prevalence of different pathogens, antimicrobial resistance patterns, and immune response profiles were analyzed. P-value <0.005 were considered as significant.

Ethical consideration

All the data collected according to ethical committee of hospital and data of patients remains confidential.

Results

Data were collected from 550 patients from both genders. There were 60% male and 40% female. Mean age of patients was 65.09 ± 8.3 years and mean duration of diabetes was 11.98 ± 6.51 years. 70% of the patients also suffering from hypertension, 45% Peripheral Neuropathy and 30% from Peripheral Vascular Disease. Mean level of Hb was 11.2 ± 1.5 g/dL. Demographic data is represented in table 01.

Table 01: Demographic and baseline values of patients				
Value				
60%				
40%				
65.09 ± 8.3				
11.98 ± 6.51				
8.5 ± 1.2				
70%				
45%				
30%				
11.2 ± 1.5				
11.8 ± 3.2				
70.5 ± 8.7				
22.0 ± 6.3				
275 ± 60				
35.2 ± 15.6				

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Grade 1 infections, characterized as superficial, corresponded to the lowest numbers, while Grade 4, indicative of limb-threatening conditions, exhibited the highest. This underscores the correlation between infection severity and microbial burden, emphasizing the importance of early intervention and appropriate management strategies to mitigate the risk of complications in diabetic foot infections.

Table 02. DTT grade and bacterial isolation in 550 patients					
Diabetic Foot Infection Grade	Number of Patients	Number of Bacteria Isolated			
Grade 1 (Superficial)	100	120			
Grade 2 (Deep)	200	260			
Grade 3 (Extensive)	150	300			
Grade 4 (Limb-threatening)	100	200			

Table 02. DFI grade and bacterial isolation in 550 natients

Staphylococcus aureus, both methicillin-sensitive (MSSA) and methicillin-resistant (MRSA) strains, are the most prevalent pathogens, with MRSA exhibiting a resistance rate of 30%. Enterococcus spp., Escherichia coli, and Pseudomonas aeruginosa are also prevalent, with Escherichia coli showing notable resistance rates of 60% to ciprofloxacin and 40% to gentamicin.

Table 05: Microbiological analysis and prevalence					
Pathogen	Prevalence (%)	Antimicrobial Resistance (%)			
Staphylococcus aureus	40%				
(MSSA)		30% (Methicillin-resistant)			
Enterococcus spp.	25%				
Escherichia coli	20%				
Pseudomonas	15%	60% (Ciprofloxacin), 40% (Gentamicin)			
aeruginosa					

Table 03. Microbiological analysis and provalance

Elevated levels of IL-6 and TNF-alpha show positive correlations with wound size (IL-6: r = 0.65, p < 0.001; TNF-alpha: r = 0.55, p = 0.005), tissue depth involvement (IL-6: r = 0.50, p = 0.002; TNFalpha: r = 0.45, p = 0.015), and amputation rate (IL-6: r = 0.75, p < 0.001; TNF-alpha: r = 0.70, p < 0.001; TNF-alpha: r = 0.70; p < 0.001; TNF-alpha: r = 0.001; r = 0.001; TNF-alpha: r =

0.001).	Furthermore,	the p	presence	of	antibodies	against	DFI-associated	pathogens	correlates
positivel	ly with treatme	ent fail	ure(r = 0)).60,	p = 0.003	and recu	urrent infections	(r = 0.55, p = 0.55)	= 0.008).

Table 04. Correlation of miniunomarkers in DF1						
Immune Marker	Clinical Outcome	Correlation Coefficient	P-value			
		(r)				
IL-6 (pg/mL)	Wound size	0.65	< 0.001			
	Tissue depth	0.50	0.002			
	Amputation rate	0.75	< 0.001			
TNF-alpha (pg/mL)	Wound size	0.55	0.005			
	Tissue depth	0.45	0.015			
	Amputation rate	0.70	< 0.001			
Antibodies against	Treatment failure	0.60	0.003			
DFI-associated Recurrent infections		0.55	0.008			
pathogens						

Staphylococcus aureus, both methicillin-sensitive (MSSA) and methicillin-resistant (MRSA) strains, exhibit notable resistance rates, with MRSA showing particularly high levels of resistance to methicillin (80%). Enterococcus spp. demonstrate moderate resistance to vancomycin (20%), while Escherichia coli and Pseudomonas aeruginosa display substantial resistance to ciprofloxacin (60%) and piperacillin/tazobactam (50%), respectively.

Pathogen	athogen Antibiotic		Intermediate	Susceptible			
		(%)	(%)	(%)			
Staphylococcus aureus (MSSA)	Methicillin	30	10	60			
Staphylococcus aureus (MRSA)	Methicillin	80	15	5			
Enterococcus spp.	Vancomycin	20	5	75			
Escherichia coli	Ciprofloxacin	60	20	20			
	Gentamicin	40	30	30			
Pseudomonas aeruginosa	Piperacillin/Tazobactam	50	10	40			

Table 05: Antimicrobial resistance pattern in DFI

Discussion

The study identified Staphylococcus aureus, Enterococcus spp., Escherichia coli, and Pseudomonas aeruginosa as the most prevalent pathogens in diabetic foot infections. The high prevalence of these pathogens underscores their significant role in DFI pathogenesis. Moreover, antimicrobial resistance was observed, particularly among Gram-negative bacteria, highlighting the challenge of treating DFI effectively [14]. Pathogenic bacteria and drug sensitivity spectrums vary regionally and are affected by the widespread use of antibiotics. Appropriate antibiotic selection for DFI is controversial because to date no empirical antimicrobial regimen has been shown to be superior. Thus, definitive therapy should be based on the identification of pathogens and their drug sensitivity [15]. The current study generated drug susceptibility results for a variety of bacterial pathogens isolated from patients with DFI [16].

Immunoproteomics is a powerful tool to identify immunoreactive molecules and develop candidate vaccines against pathogens. Immunoproteomics combines proteomics for the detection of immunoreactive antigens expressed during infections [17]. High throughput immunoproteomics

arrays offer a rapid, sensitive, and specific diagnosis of pathogens and their drug resistance profiles, which are otherwise difficult to culture or are multi-species infections. A viral proteome array comprising 646 viral antigens was developed by to examine the relationship between viral infections and the early onset of DM1 [18]. Immunoproteomic analysis revealed elevated levels of proinflammatory cytokines such as interleukin-6 (IL-6) and tumor necrosis factor-alpha (TNF-alpha) in patients with diabetic foot infections [19]. Additionally, specific antibody responses against DFI-associated pathogens were detected, suggesting a robust immune response to infection. These findings support the potential utility of immunoproteomics as a diagnostic and prognostic tool for DFI [20].

The study demonstrated significant correlations between immune markers and clinical outcomes in diabetic foot infections. Elevated levels of IL-6 and TNF-alpha were associated with increased wound size, greater tissue depth involvement, and higher rates of amputation, indicating their potential as prognostic indicators for disease severity and progression. Furthermore, the presence of antibodies against DFI-associated pathogens was predictive of treatment failure and recurrent infections, highlighting the importance of host immune response in determining clinical outcomes [21].

The study identified concerning rates of antimicrobial resistance among pathogens isolated from diabetic foot infections. Resistance to commonly used antibiotics, particularly among Gram-negative bacteria, poses challenges in selecting appropriate antimicrobial therapy and emphasizes the need for judicious antibiotic use and antimicrobial stewardship strategies [22]. *Staphylococcus aureus* grows and secretes virulent factors in glucose-rich diabetic conditions, where insulin deficiency prevents or delays immune response. *Staphylococcus aureus* has expanded its glycolytic capacity by acquiring several additional glucose transporters. Carbohydrate transporters in *S. aureus* allow efficient uptake of carbohydrates and support anaerobic growth in inflamed tissues [23]. Eleven carbohydrate transporters have been identified in *S. aureus*, while four of them (glcA, glcB, glcC, and glcU) are strictly responsible for glucose transporter might reduce glucose uptake and attenuate *S. aureus* growth [24].

The findings of this study have important clinical implications for the management of diabetic foot infections. Understanding the role of pathogens and host immune response in DFI pathogenesis can inform targeted therapeutic interventions and personalized treatment approaches. Additionally, addressing antimicrobial resistance through antibiotic stewardship programs and exploring alternative treatment modalities, such as immunomodulatory therapies, may improve outcomes in patients with diabetic foot infections.

Conclusion

It is concluded that the role of pathogens and host immune response in diabetic foot infections (DFI) is important for guiding effective treatment strategies. According to our study potential of immunoproteomics as a diagnostic and prognostic tool also emphasizing the concerning rates of antimicrobial resistance among DFI-associated pathogens.

References

- 1. Chai, Wei, et al. "The Profile of Microbiological Pathogens in Diabetic Foot Ulcers." *Frontiers in Medicine*, vol. 8, 2021, https://doi.org/10.3389/fmed.2021.656467.
- Barwell ND, Devers MC, Kennon B, Hopkinson HE, McDougall C, Young MJ, et al.. Diabetic foot infection: antibiotic therapy and good practice recommendations. *Int J Clin Pract.* (2017) 71. 10.1111/ijcp.13006
- 3. Fabiani S., Fallahi P., Ferrari S. M., Miccoli M., Antonelli A. (2018). Hepatitis C virus infection and development of type 2 diabetes mellitus: systematic review and meta-analysis of the literature. *Rev. Endocr. Metab. Disord.* 19, 405–420. doi: 10.1007/s11154-017-9440-1

- Hine J. L., De Lusignan S., Burleigh D., Pathirannehelage S., Mcgovern A., Gatenby P., et al.. (2017). Association between glycaemic control and common infections in people with type 2 diabetes: a cohort study. *Diabet. Med.* 34, 551–557. doi: 10.1111/dme.13205
- Klekotka R. B., Mizgała-Izworska E., Drzastwa W., Mazur B. (2018). The role of in the clinical diagnosis of diabetic patients. *Postępy Mikrobiol. Advancem. Microbiol.* 57, 166–178. doi: 10.21307/PM-2018.57.2.166
- 6. Kundu J., Bakshi S., Joshi H., Bhadada S. K., Verma I., Sharma S. (2020). Proteomic profiling of peripheral blood mononuclear cells isolated from patients with tuberculosis and diabetes copathogenesis A pilot study. *PLoS One* 15:e0233326. doi: 10.1371/journal.pone.0233326
- Pouget, C., Dunyach-Remy, C., Pantel, A., Schuldiner, S., Sotto, A., & Lavigne, J. P. (2020). Biofilms in diabetic foot ulcers: Significance and clinical relevance. *Microorganisms*, 8(10), 1580.
- 8. Shettigar, K., & Murali, T. S. (2020). Virulence factors and clonal diversity of Staphylococcus aureus in colonization and wound infection with emphasis on diabetic foot infection. *European Journal of Clinical Microbiology & Infectious Diseases*, *39*(12), 2235-2246.
- 9. Boulton, A. J., Armstrong, D. G., Hardman, M. J., Malone, M., Embil, J. M., Attinger, C. E., ... & Kirsner, R. S. (2020). Diagnosis and management of diabetic foot infections.
- 10. Macdonald, K. E., Boeckh, S., Stacey, H. J., & Jones, J. D. (2021). The microbiology of diabetic foot infections: a meta-analysis. *BMC infectious diseases*, *21*, 1-10.
- 11. Guzman, N. A., & Guzman, D. E. (2021). Immunoaffinity capillary electrophoresis in the era of proteoforms, liquid biopsy and preventive medicine: a potential impact in the diagnosis and monitoring of disease progression. *Biomolecules*, *11*(10), 1443.
- 12. Chen, Z. Z., & Gerszten, R. E. (2020). Metabolomics and proteomics in type 2 diabetes. *Circulation research*, 126(11), 1613-1627.
- 13. Ball, B., Langille, M., & Geddes-McAlister, J. (2020). Fun (gi) omics: Advanced and diverse technologies to explore emerging fungal pathogens and define mechanisms of antifungal resistance. *MBio*, *11*(5), 10-1128.
- 14. Thurlow, L. R., Stephens, A. C., Hurley, K. E., & Richardson, A. R. (2020). Lack of nutritional immunity in diabetic skin infections promotes Staphylococcus aureus virulence. *Science advances*, *6*(46), eabc5569.
- 15. Moritz, C. P., Paul, S., Stoevesandt, O., Tholance, Y., Camdessanché, J. P., & Antoine, J. C. (2020). Autoantigenomics: holistic characterization of autoantigen repertoires for a better understanding of autoimmune diseases. *Autoimmunity reviews*, 19(2), 102450.
- 16. Taunk, K., Kalita, B., Kale, V., Chanukuppa, V., Naiya, T., Zingde, S. M., & Rapole, S. (2020). The development and clinical applications of proteomics: An Indian perspective. *Expert Review of Proteomics*, *17*(6), 433-451.
- 17. Fulton, K. M., Baltat, I., & Twine, S. M. (2019). Immunoproteomics methods and techniques. *Immunoproteomics: methods and protocols*, 25-58.
- 18. Ceciliani, F., Eckersall, D., Burchmore, R., & Lecchi, C. (2014). Proteomics in veterinary medicine: applications and trends in disease pathogenesis and diagnostics. *Veterinary pathology*, *51*(2), 351-362.
- 19. Chen, Z. Z., & Gerszten, R. E. (2020). Metabolomics and proteomics in type 2 diabetes. *Circulation research*, 126(11), 1613-1627.
- 20. Guzman, N. A., & Guzman, D. E. (2021). Immunoaffinity capillary electrophoresis in the era of proteoforms, liquid biopsy and preventive medicine: a potential impact in the diagnosis and monitoring of disease progression. *Biomolecules*, *11*(10), 1443.
- 21. Thurlow, L. R., Stephens, A. C., Hurley, K. E., & Richardson, A. R. (2020). Lack of nutritional immunity in diabetic skin infections promotes Staphylococcus aureus virulence. *Science advances*, *6*(46), eabc5569.

- 22. Moritz, C. P., Paul, S., Stoevesandt, O., Tholance, Y., Camdessanché, J. P., & Antoine, J. C. (2020). Autoantigenomics: holistic characterization of autoantigen repertoires for a better understanding of autoimmune diseases. *Autoimmunity reviews*, 19(2), 102450.
- 23. de Almeida, J. R. F., Jannuzzi, G. P., Kaihami, G. H., Breda, L. C. D., Ferreira, K. S., & de Almeida, S. R. (2018). An immunoproteomic approach revealing peptides from Sporothrix brasiliensis that induce a cellular immune response in subcutaneous sporotrichosis. *Scientific Reports*, 8(1), 4192.
- 24. Taunk, K., Kalita, B., Kale, V., Chanukuppa, V., Naiya, T., Zingde, S. M., & Rapole, S. (2020). The development and clinical applications of proteomics: An Indian perspective. *Expert Review of Proteomics*, *17*(6), 433-451.