



EVALUATION OF GROWTH PERFORMANCE AND BLOOD BIOCHEMICAL CHANGES IN FISHES (*LABEO ROHITA*) SUPPLEMENTED WITH FRUCTOOLIGOSACCHARIDES UNDER HEAT STRESS

Sadaf Gulzar¹, Saira Parveen¹, Muhammad Usman Saleem², Abdul Asim Farooq³, Maria Gulzar⁴, Maryam Ikram⁵, Ayesha Maqsood¹, Muhammad Nawaz^{1*}

¹Department of Environmental Sciences, Bahauddin Zakariya University, Multan, Pakistan

²Department of Biosciences, Faculty of Veterinary Sciences, Bahauddin Zakariya University, Multan, Pakistan

³Department of Clinical Sciences, Faculty of Veterinary Sciences, Bahauddin Zakariya University, Multan, Pakistan

⁴Department of Botany, Women University, Multan, Pakistan

⁵Department of Botany, Women University, Multan, Pakistan

***Corresponding author:** Dr. Muhammad Nawaz

*Email: mnawaz@bzu.edu.pk

ABSTRACT

This study was conducted to examine the effects of heat stress (HS) on growth performance and selected blood biochemical attributes of fishes supplemented with fructooligosaccharide (FOS). The main objectives revolved around evaluating the negative impacts of HS and amelioration of those harmful effects by using FOS along with exploring the potential benefits of FOS in improving growth and biochemical parameters of (*Labeo rohita*) fishes reared under cyclic heat stress. Moreover, this study offers insightful information related to fish health, management and aquaculture practices. One hundred-eight (n=108) rohu fishes (*Labeo rohita*) were selected and were equally divided into five groups with each group having 3 replicates (n=18) on 21st day. The details of the grouping are T₀, T₁, T₂, T₃, T₄, T₅, in which T₀ was considered as control group which was fed with basal diet. T₁ and T₂ groups were supplemented with 1% and 2% FOS respectively, without HS. The group T₃ was positive control whereas, T₄ and T₅ were exposed to HS at 40 °C supplemented with 1% and 2% FOS respectively. The results revealed that supplementation of fructooligosaccharides significantly improved growth performance and selected biochemical attributes of fishes during 63 days of rearing. The T₀ group exhibited significant decrease in growth and selected biochemical parameters compared to the treatment groups however, after treating with FOS the negative effects of HS were significantly reduced. Moreover, T₁ showed positive improvements in all the selected parameters of this experiment. Growth parameters were found higher in T₂ as in final weight (10.85%), weight gain (51.06%), survival rate (2.09%) at (P ≤ 0.05) supplemented with 2% FOS as compared to T₀ and T₁ but the aforementioned parameters were found lower in T₃ under influence of 40°C of HS as compared to T₀. Although biochemical parameters were found higher in T₂ as in albumin (119.38%), globulin (132.20%), ALP (7.81%) and total protein (34.28%) As compared to T₀ and T₁ but afforested parameters such as globulin and total protein were significantly decreased under HS as compared to rest of the treatment. Furthermore, several serum biochemical markers, including urea (40.86%),

cholesterol (28.58%), and triglycerides (31.24%) were significantly decreased in T₂ as compared to T₀ and T₁. In conclusion, the results revealed that supplementation of FOS alleviate the negative effects of HS in fishes (*Labeo rohita*).

Key words: Cyclic heat stress, Fructooligosaccharides, growth performance, serum biochemical markers, urea, cholesterol, triglycerides.

INTRODUCTION

Aquaculture production has experienced the profligate growth in the animal food production in the last decade (Stankus, 2021). It is estimated to rise to 53% by the year 2030 (FAO, 2020). Almost 1 billion people make their living through aquaculture industry which is rapidly expanding (Obi *et al.*, 2016). In 2018, 46% of the world's total fish production was attributed to aquaculture (ACTION, 2020). It has been reported that 50% of animal protein consumed by humans comes from aquaculture which is one of the food industry's fastest-growing segments globally. There are some problems such as heat stress (HS) that stop aquaculture from flourishing. (McKenzie *et al.*, 2021). In many parts of the world HS is one of the major environmental factor compromising production performance of animals (El-Tarabany *et al.*, 2017). The HS negatively effects growth of fish. Amur sturgeon (*Acipenser schrenckii*) and Atlantic salmo (*Salmo salar*) (Handeland *et al.*, 2008) are two fishes in which negative effects of HS have been reported which include reduced food intake whereas, in common carp (*Cyprinus carpio communis*) abnormal swimming patterns and respiratory disorders were observed. The HS is a serious issue that demands serious concern due to global warming (Hansen *et al.*, 2010). Ruminants, poultry and pigs are susceptible to HS due to their high metabolic rate, higher basal metabolic heat production, rapid growth and high level of production (Tajima *et al.*, 2007). The HS in fishes negatively impacts their feeding, growth and disease resistance by altering intestinal microbiota, disrupting intestinal barrier function and causing metabolic disorders. (Zhou *et al.*, 2022). Changes in the climate is making HS gradually harmful to fish aquaculture. Cold-water fish like rainbow trout are particularly sensitive to water temperature fluctuations, leading to stress and negative impacts on food consumption, behavior, growth and survival above 18°C. (Zhu *et al.*, 2019). Fructo-oligosaccharides (FOS) are prebiotics derived from sucrose through the action of certain enzyme. They are a low-calorie and non-carcinogen sweetener that promotes ion absorption in the stomach, reduces fat, cholesterol levels and supports the growth of beneficial gut bacteria (Bali *et al.* (2015)

The (FOS) are a beneficial carbohydrate found in a variety of functional food products which include dairy items, bakery items, beverages, jams, candies, chocolates, breakfast cereals, meat, ice cream and confectionery (Kherade *et al.* (2021). The FOS is a type of oligosaccharide that is recognized as safe and have recently drawn particular attention as food ingredients because of their unique aspects and advantages in health (Tódero *et al.*, 2019). Introducing fish to new environments boosts aquaculture yields and water resource management. Fish contribute to 15% of animal protein in human diets (Casal, 2006). Aquaculture is a swiftly growing food industry which involves raising fish, shellfish and aquatic plants. During the last decade aquaculture has supplied one-third of the world's seafood (Reverter *et al.*, 2014). Fish from both inland and marine capture fisheries are highly consumed and traded globally playing a crucial role in supporting livelihoods, food security and improving economic growth (Loring *et al.*, 2019). According to FAO (2020) about 120 million people in developing nations depend on fisheries for their livelihoods with over 90 percent of them engaged in small-scale fisheries (Loring *et al.*, 2019). This study aims to explore the potential of FOS as a dietary supplement to mitigate the negative effects of HS on fish growth. Specifically, this study will examine whether FOS can enhance growth performance and alleviate hazardous effects of HS on fishes (*Labeo Rohita*). By altering dietary strategies the research aims to improve fish performance and resilience under HS. This investigation aims to evaluate the growth performance and selected serum biochemical changes in fish supplemented with FOS during cyclic HS contributing to the development of sustainable aquaculture techniques for countering climate change challenges.

Material and Methods:

2.1. Animal Ethics:

All the experimental procedures were approved by Animal Care and Use Committee Bahauddin Zakariya University, Multan, Pakistan.

2.2. Experimental setup:

The experiment was conducted in fisheries research lab IPAB and the examination of the parameters was executed in the laboratory of Department of Environmental Science, Bahauddin Zakariya University (BZU) Multan. In this experiment one hundred-eight rohu fish (*Labeo Rohita*) were purchased from local fish farm having an average body weight of 15gm which were kept under environmentally controlled conditions.

2.3. Experimental Design:

One hundred-eight rohu fishes (*Labeo Rohita*) with an average weight of 15 ± 2 gm were selected and divided into six groups T₀, T₁, T₂, T₃, T₄, T₅. All groups were fed with basal diet on daily basis (twice daily) and grown at normal temperature (26°C to 28°C) for 3 weeks. Three replicates were selected for each treatment. The experimental continued for 6 weeks and selected fish groups were subjected to HS during the last three weeks of experiment. After 21 days, the weight of the fish increased from 15 ± 2 gm to 23.5 gm. Twenty- one days old fish with average weight of 23.5gm was exposed to FOS and HS. The treatment design for each group was set as:

- Group I: Basal diet +No heat stress + No FOS named as T₀
- Group II: Basal diet +No heat stress + 1% FOS named as T₁
- Group III: Basal diet+ No heat stress + 2% FOS kept as T₂
- Group IV: Basal diet+ only heat stress (40°C) + NO FOS kept as T₃
- Group V: Basal diet +1% FOS + heat stress Kept as T₄
- Group VI: Basal diet +2% FOS + heat stress kept as T₅

The appropriate sizes of the basal diet were utilized as the basal feed in the experiment and its composition is shown in Table 1. The percentage composition of the ingredients for basal diet was formulated in line as in previously used by (Guerreiro *et al.*, 2016; Genç *et al.*, 2020).

Table 1. Percentage composition of ingredients for basil diet

Ingredients	%
Crude protein	48
Crude lipid	18
Crude Ash	12
Crude cellulose	2
Phosphorous	1.5
Moisture	11
Fat	13
Fiber	2.5
Starch	5

2.4. Assessment of selected parameters:

The growth parameters such as weight gain, final weight, feed intake and survival rate were estimated using digital electronic balance. The biochemical parameters such as aspartate aminotransferase (AST) (IU/L), alanine aminotransferase, glucose(mg/dL), triglycerides, total cholesterol and total protein (TP) was measured in the automatic biochemical analyzer (Mindray BS-400 bio- chemistry analyzer) using kits assay specially designed for fish detection. The alanine aminotransferase (ALT) and Aspartate aminotransferase (AST) enzymes were measured calorimetrically from plasma with a kit (Abcam Cambridge, UK) in 96-well microplates. Amylase activity was determined by the starch

hydrolysis method, according to the Somogy–Nelson colorimetric method. According to Hidalgo et al. (1999) the casein hydrolysis method was used to measure total protease activity.

2.5 Statistical Analysis

The collected data was analyzed statistically by using excel sheets and statistix 8.1 version. For the calculation of mean and standard error, MS office-2010 was used while one way Analysis of Variance (ANOVA) was done through statistical software statistix 8.1. Significant difference between the treatments was also observed through the Duncan Multiple Range Test (DMRT). The differences were considered significant at $P < 0.05$.

3. RESULTS AND DISCUSSIONS

RESULTS

The FOS are type of prebiotic carbohydrate that selectively stimulates the formation of beneficial bacteria in the gut and mitigate the impact of heat stress on fish (*Labeo Rohita*).

3.1. Effect of heat stress and fructooligosaccharides supplementation on growth Parameters:

There was a significant effect of FOS on growth parameter except feed intake. As illustrated in (fig.1a) final weight was significantly increased 4.91% in T₁ supplemented with 1% FOS while T₂ showed significant increase in final weight which was found to be 10.85% compared to T₀ and T₁. The final weight of T₃ fish (*Labeo Rohita*) under the effect of 40 °C heat showed prominent difference as compared to control as well as rest of the treatments which was found to be less 9.84% than T₀ at $P \leq 0.05$. The final weight of T₄ was significantly increased that was found to be 2.81% compared to control while T₅ exhibited an increase in final weight that was 7.14% compared to T₄. As shown in (fig.1b&d) weight gain and survival rate were significantly increased in T₁ that was found to be 37.95%, 1.27% respectively supplemented with 1% FOS As compared to T₀. While T₂ showed significant increase in weight gain and survival rate that was found to be 51.06%, 2.09% respectively as compared to T₀ and T₁. The T₃ showed significant decrease in weight gain 12.75% and survival rate 11.76% As compared to T₀. As demonstrated in (fig.1b) that the weight gain of T₄ was significantly increased that was found to be 5.63% As compared to T₀ and T₃ while T₅ showed some extent of increase in weight gain that was 42.98% as compared to T₄ and T₃.

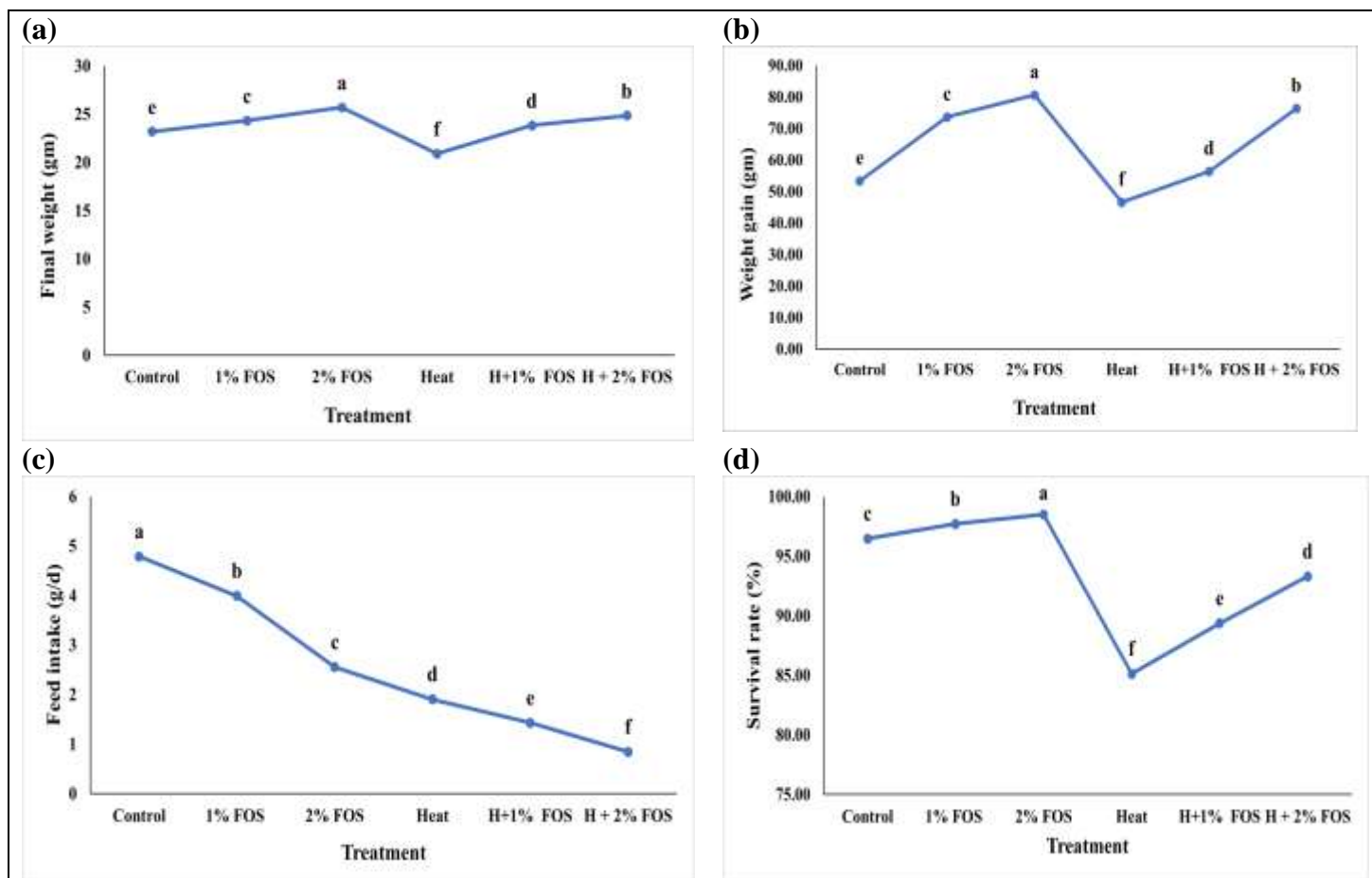


Figure 1: *Labeo rohita* fish fed with different level of fructooligosaccharides and heat on (a)final weight(gm) (b)weight gain(gm) (c)Feed intake (g/d) (d) Survival rate (%). Superscript denotes significant differences between control and treatment groups. ($p < 0.05$)

As shown in (fig.1d) survival rate of T₄ that was found to be 7.35% which was significantly lower as compared to T₁ and T₀. Whereas, T₅ compared to T₄ exhibited significant decrease in survival rate which was found to be 3.28%. As demonstrated in (fig.1c) feed intake of T₁ was significantly reduced to 16.63% compared to T₀. Whereas, T₂ exhibited a further significant decrease in feed intake 46.62% as compared to T₀ and T₁. The results revealed that supplementation of FOS produced significant effects and countered the negative effects of HS at $P \geq 0.01$. Similarly, T₃ showed significant decrease in feed intake that was found to be 60.19% under HS compared to rest of the treatment groups as shown in (fig.1c) whereas, feed intake was found increase in T₄ 70.14% as compared to T₃ and T₁. Feed intake of T₅ compared to T₄ was significantly decreased which was found to be 82.39% compared to T₃ and T₂.

3.2. Effect of heat stress and fructooligosaccharides supplementation on biochemical parameters:

As shown in Table 2 HS and FOS supplementation showed significant impact on albumin of fishes. The albumin of T₁ (4.61 ± 0.16) was significantly increased 57.02% compared to T₀. The albumin content was also significantly increased in T₂ (6.45 ± 0.16) that was 119.38% at 2% dose of FOS compared to control at $P \leq 0.05$. Similarly, it increased significantly in T₃ (1.38 ± 0.12) which was significantly lower 52.94% compared to T₁ and T₂. However, compared to T₃ and the rest of the treatment groups T₄ (3.52 ± 0.30) and T₅ (5.59 ± 0.22) exhibited increase in albumin which was 19.95% and 90.36% respectively. Globulin and total protein of T₁ (5.68 ± 0.12 and 6.57 ± 0.29) significantly increased 80.61%, 20.25% respectively compared to T₀. Moreover, in T₂ a comparable rise in globulin (7.30 ± 0.12) and total protein (7.33 ± 0.15) was found to be 132.20%, 34.28% in

comparison to the T₀ and T₁. The T₃ demonstrated a considerable decrease in globulin (2.80±0.17) and total protein (2.68± 0.22) which was found to be 10.91% and 50.82% respectively, compared to the other groups whereas, globulin content of T₅ (6.79±0.04) was significantly higher and was found to be 115.78% compared to T₄. In this study (Table 2) revealed significant impact of FOS on protease which increased significantly 51.27% in T₁ (8.49±0.18).

Table:2: Biochemical parameters of *Labeo rohita* fed different levels of fructooligosaccharide

Parameters	Treatment groups					
	Control	1% FOS	2% FOS	Heat ONLY	Heat + 1% FOS	Heat + 2% FOS
Albumin (g d/l)	2.94±0.06	4.61±0.16	6.45± 0.16	1.38± 0.12	3.52± 0.30	5.59± 0.22
Cholesterol (mg d/l)	106.05±0.05	83.35±0.12	75.74±0.04	109.48±0.30	89.51± 0.38	79.11± 0.06
Globulin (g d/l)	3.14± 0.08	5.68± 0.12	7.30 ±0.12	2.80 ±0.17	4.81± 0.18	6.79± 0.04
Protease (µm/g)	5.61± 0.17	8.49± 0.18	10.78±0.16	4.35± 0.31	7.94± 0.04	9.21± 0.14
Amylase(µm/g)	2.69± 0.14	5.66± 0.16	6.93± 0.03	4.44± 0.32	7.62 ±0.43	8.79± 0.24
Glucose (mg/dl)	70.18± 0.12	72.90±0.11	85.65±0.08	77.79± 0.18	93.79± 0.18	110.16± 0.06
Total protein (g/d)	5.46 ±0.30	6.57 ±0.29	7.33± 0.15	2.68± 0.22	3.64 ±0.45	4.64± 0.20
Triglycerides(mmol/dL)	57.47± 0.21	46.49±0.20	39.51±0.39	58.26± 0.09	49.43± 0.08	42.83± 0.19
Urea (mg/dl)	19.52 ±0.19	14.89±0.08	11.54±0.17	21.43± 0.31	17.45± 0.39	16.16± 0.16
Lipase(µ/g)	2.72± 0.36	3.76± 0.20	5.69± 0.31	4.95± 0.04	6.46 ±0.22	7.65 ±0.29
ALP (U/l)	113.53±0.21	118.65±0.15	122.40±0.17	103.70±0.22	110.63 ±0.31	119.35± 0.11
ALT (U/l)	27.19±0.13	25.69± 0.03	18.59±0.14	30.65± 0.42	29.66± 0.09	22.19 ±0.13
AST (U/l)	33.21± 0.09	29.11± 0.08	25.17± 0.12	34.65± 0.45	27.55± 0.17	22.20± 0.14

values are mean±SE of three observations. Mean bearing different superscript in a row differ significantly (p≤0.05). AST: Aspartate aminotransferase; ALT: Alanine aminotransferase; ALP: Alkaline phosphatase. Significant level (p≥0.001)

However, T₂ (10.78±0.16) exhibited significant increase in protease that was found to be 91.98% as compared to T₀ and T₁ with a significance level of P≤0.05. But T₃ (4.35±0.31) exhibited significantly decrease in protease which was observed 22.43% compared to T₀. Whereas, administration of 1% FOS with heat dramatically enhanced protease in T₄ (7.94±0.04) which was 41.48% higher than T₀. Similarly, protease of T₅ (9.21± 0.14) increased that was found 64.03% as compared to T₃ and T₄ (Table 2). The ALP significantly increased in T₁ (118.65±0.15) that was observed to be 4.31% compared to T₀. Similarly, T₂ (122.40±0.17) showed further increase in ALP 7.81% than T₀ and T₁ at P≤0.05. While ALP of T₃ (103.70±0.22) significantly decreased compared to T₀. Whereas, T₅ (119.35± 0.11) showed significant decrease in ALP 2.55% compared to T₀ and T₁, at a significance level of P≤0.05. As shown in (Table 2) amylase of T₁ (5.66± 0.16) showed significantly increase that was observed 110.53% compared to T₀. Similarly, further 157.74% increase in amylase was observed in T₂ (6.93±0.03) compared to control at P≤0.05. Moreover, amylase of T₃ (4.44± 0.32) showed significant decrease which was 65.30% compared to T₀ while, amylase in T₅ (8.79± 0.24) increased 226.88% compared to T₀. As presented in Table 2 significant impacts of HS and FOS on lipase content of (*Labeo Rohita*) fish were observed. The lipase content of T₁ (3.76± 0.20) increased significantly which was 38.23% compared to T₀. Furthermore, lipase (5.69± 0.31) of T₂ increased that was 109.31% compared to T₀ at P≤0.05. Whereas, lipase content of T₃ (4.95±0.04) exhibited a significant difference compared to T₀ and other treatment groups. While, lipase of T₄ and T₅ (6.46±0.22, 7.65±0.29 respectively) significantly increased which was 137.5% compared to 181.37% of T₀ and T₁. Furthermore, glucose level was significantly increased due to FOS supplementation. The T₁ (72.90±0.11) exhibited significant increase in glucose level that was noted to be 3.88% compared to T₀. However, supplementation of FOS in T₂ (85.65±0.08) significantly increased which was 22.04% compared to the other treatment groups. While in T₃ (77.79±0.18) glucose level was significantly increased which was found to be 10.85% at HS. Although further increase was observed in T₅ (110.16±0.06) which was 56.97% higher than T₄.

The current investigation found that several serum biochemical markers including urea, cholesterol, triglycerides, AST and ALT, significantly decreased in fishes fed with diets supplemented with FOS

as demonstrated in Table 2. Cholesterol of T₁ (83.35±0.12) significantly decreased that was 21.40% compared to T₀. Whereas, T₂ (75.74±0.04) showed a comparable decrease that was observed to be 28.58% compared to T₀ at P≤0.05. While in T₃ (109.48±0.30) cholesterol level was significantly increased which was found to be 3.23% under influence of HS. While, cholesterol of T₄ and T₅ (89.51±0.38, 79.11±0.06 respectively) exhibited significant decrease which was observed to be 15.59%, 25.40% respectively as compared to T₀ and T₁. Results revealed that triglycerides of T₁ (46.49±0.20) significantly decreased that was seen in the form of 19.10% compared to T₀. Furthermore, a decrease was observed in T₂ (39.51±0.39) which was found to be 31.24% compared to T₁ at P<0.05 as well as control. While T₃ (58.26±0.09) showed significant increase that was 1.38% compared to T₂ and T₀. The T₅ (42.83±0.19) exhibited slight decrease in triglycerides that was observed to be 25.47% compared to T₄ and T₃.

Urea of T₁ (14.89±0.08) significantly decreased to 23.73% compared to T₀. Furthermore, a decrease in T₂ (11.54±0.17) was noted to be 40.86% compared to T₁, at significance level of P≤0.05. But in case of T₃ (21.43±0.31) exhibited significantly higher level of urea which was observed to be 9.78%. while T₄ revealed significant decrease in urea level that was observed to be 10.58% compared to T₀ and T₁. Similarly, T₅ (16.16±0.16) to some extent of revealed decreased in urea that was 17.2% as compared to T₃. In this study, liver enzymes AST was significantly decreased in T₁ (29.11±0.08) that was 12.35% compared to the control group. The AST level was significantly decreased in T₂ (25.17±0.12) that was 24.20% compared to T₀ at a significance level of P≤0.05. While T₃ (34.65±0.45) exhibited significant increase that was 4.31% compared to T₂ and T₀. Urea of T₅ (22.20±0.14) significantly decreased that was found to be 33.16% compared to T₄ and T₃. Results revealed a considerable impact of FOS on the ALT of the fish (*Labeo rohita*), as shown in Table 2. The ALT was significantly decreased in T₁ (25.69±0.03) that was 5.51% as compared to T₀. Similarly, further decrease was observed in T₂ (18.59±0.14) that was noted to be 31.64% compared to control at P≤0.05. Whereas, ALT of T₃ (30.65±0.42) significantly increased which was 12.72% compared to T₀. Moreover, T₅ (22.19±0.13) significantly decreased ALT which was 18.41% compared to T₄ and T₃.

DISCUSSIONS

The FOS are prebiotic carbohydrates that selectively stimulates the formation of beneficial bacteria in the gut and mitigate the impact of HS on fish (*Labeo rohita*). Results of the present study conclusively demonstrated that dietary supplementation of FOS at 1% and 2% considerably boosted the digestive enzyme activities in (*Labeo rohita*) fishes which are comparable to the findings of Soleimani *et al.* (2012) who reported similar effects in Caspian roach (*Rutilus Rutilus*) fry and snout bream (*Megalobrama Amblycephala*) fingerlings (Wu *et al.*, 2013). Protease, amylase and lipase activities were reported to increase with increasing doses of dietary FOS. The causes of the higher levels are results of endogenous and exogenous microbial activities regulated by FOS. The findings of the present experiment demonstrated that fishes fed with diets supplemented with FOS exhibited significantly higher levels of selected serum biochemical markers which included glucose, albumin, globulin, ALP and total protein as shown in above table which are also comparable with the findings of Ali *et al.* (2020) but the decreased albumin and total protein in the treatment groups suggest an abnormality in kidney and liver function. In contrast to our findings Reza *et al.* (2009) in juvenile beluga reported that in an 8-week feeding trial various biochemical parameters were not significantly affected by dietary prebiotic inulin. The enzyme activities ALP, ALT and AST levels were not affected by supplementation of mannoooligosaccharides at 0, 0.5, 1, 1.5, and 2% in Asian Seabass fingerlings. The current investigation found that several serum biochemical markers including urea, cholesterol, triglycerides, AST and ALT significantly decreased in fish fed with diets supplemented with FOS which are in line to the findings of Feng and Xia (2019) who fed FOS to yellow broiler hens to lower the levels of total cholesterol and serum triglyceride which are two of the key elements of blood lipids. The FOS possesses the properties of dietary fiber that is water-soluble and may absorb lipids from the body while enhancing lipid metabolism to lower blood lipid levels. Serum urea nitrogen levels in the FOS group are lower in the experiment. The findings of the abovesaid study is in line to our results

because in this experiment FOS is used as prebiotic supplement in yellow broiler which promotes development and activity of good bacteria in the gut. The digestion and absorption of nutrients are enhanced by these bacteria especially those of the *Bifidobacterium* and *Lactobacillus* species. Fish fed FOS-supplemented diets exhibited increased nutritional utilization, resulting in lower blood levels of urea, cholesterol and triglycerides randomly. According to Taati *et al.* (2011) changes in biochemical parameters may differ from one species to another depending on numbers of variables that may vary according to the dose of prebiotics added to the diet, type of feed ingredients and the experimental settings. The high levels of AST and ALT in the serum are results of HS.

The result of the current study showed that positive impact of FOS on growth parameters which included final weight, weight gain and survival rate which significantly increased while feed intake significantly decreased. These results are according to the results reported by Hu *et al.* (2019) but differ from the finding of Hashmi *et al.* (2023) in which final weight and weight gain significantly decreased after supplementation of prebiotic. The difference from our results is due to the fact that they used pectin as prebiotic whereas in our study the prebiotic used was FOS. According to the results of the current study, it was observed that the exposure of HS significantly decreased growth parameters of (*Labeo rohita*) which included final weight, weight gain, feed intake and survival rate which is similar to the findings of Mehaisen *et al.* (2017) who reported that growth performance of quails were negatively affected by HS. Moreover, recent studies have shown that colonies of chicken exposed to HS (over 30 °C) showed low production performance along with high morbidity and mortality resulting in significant economic losses on poultry farms (Niu *et al.*, 2009; Quinteiro-Filho *et al.*, 2012).

The findings of this study shows that glucose level of (*Labeo Rohita*) under HS was significantly increased which is similar to the results of Imik *et al.* (2010) because the increasing level of plasma glucose are derived from rise in free radicals along with the release of ACTH and cortisol hormones which restrict insulin release and promotes gluconeogenesis in response to HS Ajakaiye *et al.* (2010). Likewise Rashidi *et al.* (2010) also discovered that HS raised plasma glucose levels. Albumin, globulin and total protein was significantly decreased under influence of HS which is similar to the findings of Sarica *et al.* (2017) as HS disrupts protein synthesis and accelerates its breakdown which lowers the quantity of protein in the body resultantly decreasing albumin, globulin and total protein levels related to alterations caused by HS in protein production and metabolism.

CONCLUSIONS

Nexus to the above it is concluded that cyclic heat stress (HS) adversely effects the health of rohu fishes (*Labeo Rohita*) by decreasing growth performance and negatively altering the selected biochemical parameters. Our results reveal that the detrimental effects of HS can be minimized with fructooligosaccharides (FOS) supplementation. The results of this study are indicative that rohu fishes (*Labeo Rohita*) exposed to HS exhibited significant recovery in growth performance and selected serum biochemical parameters after FOS supplementation. Results of this study reveal that FOS alleviate negative effects of HS. Therefore, it is concluded that FOS mitigate the harmful effects of HS in rohu fishes (*Labeo Rohita*).

REFERENCES

1. ACTION, S.I., 2020. World fisheries and aquaculture. Food and Agriculture Organization, 2020: 1-244.
2. Ajakaiye, J.J., A. Perez-Bello and A. Mollineda-Trujillo, 2010. Impact of vitamins c and e dietary supplementation on leukocyte profile of layer hens exposed to high ambient temperature and humidity. *Acta Veterinaria Brno*, 79(3): 377-383.
3. Ali, S.S.R., K. Ambasankar, P.E. Praveena, S. Nandakumar, S. Balachandran and K. Ramachandran, 2020. Effect of dietary prebiotic fructooligosaccharide on histology, digestive enzyme activity, biochemical and immunological parameters of asian seabass (*Lateolabrax niloticus*) fingerlings. *Indian J. Fish*, 67(4): 80-88.

4. Bali, V., P.S. Panesar, M.B. Bera and R. Panesar, 2015. Fructo-oligosaccharides: Production, purification and potential applications. *Critical Review Food Science Nutrition*, 55(11): 1475-1490. Available from <https://www.ncbi.nlm.nih.gov/pubmed/24915337>. DOI 10.1080/10408398.2012.694084.
5. Casal, C.M.V., 2006. Global documentation of fish introductions: The growing crisis and recommendations for action. *Biological Invasions*, 8: 3-11.
6. El-Tarabany, M.S., A.A. El-Tarabany and M.A. Atta, 2017. Physiological and lactation responses of egyptian dairy baladi goats to natural thermal stress under subtropical environmental conditions. *International Journal of Biometeorology*, 61: 61-68.
7. FAO, F., 2020. Agriculture organization of the united nations the state of world fisheries and aquaculture 2020: Sustainability in action. Rome: Food and Agriculture Organization of the United Nations: 1-244.
8. Feng, Z. and Z. Xia, 2019. Effects of dietary fructo-oligosaccharides on laying performance and serum biochemical parameters of yellow broiler breeder hens. In: *E3S Web of Conferences*. EDP Sciences: pp: 01081.
9. Genç, E., M.A. Genç, D. Kaya, F.S. Secer, A. Qaranjiki and D. Güroy, 2020. Effect of prebiotics on the growth performance, haematological, biochemical, and histological parameters of african catfish (*clarias gariepinus*) in recirculating aquaculture system. *Turkish Journal of Veterinary & Animal Sciences*, 44(6): 1222-1231.
10. Guerreiro, I., C.R. Serra, P. Enes, A. Couto, A. Salvador, B. Costas and A. Oliva-Teles, 2016. Effect of short chain fructooligosaccharides (scfos) on immunological status and gut microbiota of gilthead sea bream (*sparus aurata*) reared at two temperatures. *Fish & Shellfish Immunology*, 49: 122-131.
11. Handeland, S.O., A.K. Imsland and S.O. Stefansson, 2008. The effect of temperature and fish size on growth, feed intake, food conversion efficiency and stomach evacuation rate of atlantic salmon post-smolts. *Aquaculture*, 283(1-4): 36-42.
12. Hansen, J., R. Ruedy, M. Sato and K. Lo, 2010. Global surface temperature change. *Reviews of Geophysics*, 48(4).
13. Hashmi, H.S., N. Khan, K.J. Iqbal, M. Fatima, K. ANJUM, S. Abbas, M. Awais, S. Nazir, M. Asghar and M. De Zoysa, 2023. Studies on the growth, immunomodulation and gut morphometry of *labeo rohita* fed pectin. *Czech Journal of Animal Science*, 68(4).
14. Hu, X., H.L. Yang, Y.Y. Yan, C.X. Zhang, J.d. Ye, K.L. Lu, L.H. Hu, J.J. Zhang, L. Ruan and Y.Z. Sun, 2019. Effects of fructooligosaccharide on growth, immunity and intestinal microbiota of shrimp (*litopenaeus vannamei*) fed diets with fish meal partially replaced by soybean meal. *Aquaculture Nutrition*, 25(1): 194-204.
15. Imik, H., M.A. Atasever, M. Koc, M. Atasever and K. Ozturan, 2010. Effect of dietary supplementation of some antioxidants on growth performance, carcass composition and breast meat characteristics in quails reared under heat stress. *Czech Journal of Animal Science*, 55(5): 209-220.
16. Kherade, M., S. Solanke, M. Tawar and S. Wankhede, 2021. Fructooligosaccharides: A comprehensive review. *Journal of Ayurvedic and Herbal Medicine*, 7(3): 193-200.
17. Loring, P.A., D.V. Fazzino, M. Agapito, R. Chuenpagdee, G. Gannon and M. Isaacs, 2019. Fish and food security in small-scale fisheries. *Transdisciplinarity for small-scale fisheries governance: analysis and practice*: 55-73.
18. McKenzie, D., B. Geffroy and A.P. Farrell, 2021. Effects of global warming on fishes and fisheries. *Journal of Fish Biology*, 98(6): 1489-1492.
19. Mehaisen, G.M., R.M. Ibrahim, A.A. Desoky, H.M. Safaa, O.A. El-Sayed and A.O. Abass, 2017. The importance of propolis in alleviating the negative physiological effects of heat stress in quail chicks. *Plos One*, 12(10): e0186907.

20. Niu, Z., F. Liu, Q. Yan and W. Li, 2009. Effects of different levels of vitamin e on growth performance and immune responses of broilers under heat stress. *Poultry Science*, 88(10): 2101-2107.
21. Obi, F., B. Ugwuishiwu and J. Nwakaire, 2016. Agricultural waste concept, generation, utilization and management. *Nigerian Journal of Technology*, 35(4): 957–964-957–964.
22. Quinteiro-Filho, W.M., A. Gomes, M.L. Pinheiro, A. Ribeiro, V. Ferraz-de-Paula, C.S. Astolfi-Ferreira, A.J.P. Ferreira and J. Palermo-Neto, 2012. Heat stress impairs performance and induces intestinal inflammation in broiler chickens infected with salmonella enteritidis. *Avian Pathology*, 41(5): 421-427.
23. Rashidi, A., Y.G. Ivvari, A. Khatibjoo and R. Vakili, 2010. Effects of dietary fat, vitamin e and zinc on immune response and blood parameters of broiler reared under heat stress. *Res. J. Poult. Sci*, 3(2): 32-38.
24. Reverter, M., N. Bontemps, D. Lecchini, B. Banaigs and P. Sasal, 2014. Use of plant extracts in fish aquaculture as an alternative to chemotherapy: Current status and future perspectives. *Aquaculture*, 433: 50-61.
25. Reza, A., H. Abdolmajid, M. Abbas and A.K. Abdolmohammad, 2009. Effect of dietary prebiotic inulin on growth performance, intestinal microflora, body composition and hematological parameters of juvenile beluga, *huso huso* (linnaeus, 1758). *Journal of the World Aquaculture Society*, 40(6): 771-779.
26. Sarica, S., H. Aydin and G. Ciftci, 2017. Effects of dietary supplementation of some antioxidants on liver antioxidant status and plasma biochemistry parameters of heat-stressed quail. *Turkish Journal of Agriculture-Food Science and Technology*, 5(7): 773-779.
27. Soleimani, N., S.H. Hoseinifar, D.L. Merrifield, M. Barati and Z.H. Abadi, 2012. Dietary supplementation of fructooligosaccharide (fos) improves the innate immune response, stress resistance, digestive enzyme activities and growth performance of caspian roach (*rutilus rutilus*) fry. *Fish & Shellfish Immunology*, 32(2): 316-321.
28. Stankus, A., 2021. State of world aquaculture 2020 and regional reviews: Fao webinar series. *FAO Aquaculture Newsletter*(63): 17-18.
29. Taati, R., M. Soltani, M. Bahmani and A. Zamini, 2011. Growth performance, carcass composition, and immunophysiological indices in juvenile great sturgeon (*huso huso*) fed on commercial prebiotic, immunoster.
30. Tajima, K., I. Nonaka, K. Higuchi, N. Takusari, M. Kurihara, A. Takenaka, M. Mitsumori, H. Kajikawa and R.I. Aminov, 2007. Influence of high temperature and humidity on rumen bacterial diversity in holstein heifers. *Anaerobe*, 13(2): 57-64.
31. Tódero, L.M., C.G.V. Rechia and L.H.S. Guimarães, 2019. Production of short-chain fructooligosaccharides (scfos) using extracellular β -d-fructofuranosidase produced by *aspergillus thermomutatus*. *Journal of Food Biochemistry*, 43(8): e12937.
32. Wu, Y., W.B. Liu, H.Y. Li, W.N. Xu, J.X. He, X.F. Li and G.Z. Jiang, 2013. Effects of dietary supplementation of fructooligosaccharide on growth performance, body composition, intestinal enzymes activities and histology of blunt snout bream (*m egalobrama amblycephala*) fingerlings. *Aquaculture nutrition*, 19(6): 886-894.
33. Zhou, C., S. Yang, W. Ka, P. Gao, Y. Li, R. Long and J. Wang, 2022. Association of gut microbiota with metabolism in rainbow trout under acute heat stress. *Frontiers in Microbiology*, 13: 846336.
34. Zhu, L., R. Liao, N. Wu, G. Zhu and C. Yang, 2019. Heat stress mediates changes in fecal microbiome and functional pathways of laying hens. *Applied microbiology and biotechnology*, 103: 461-472.