Journal of Population Therapeutics & Clinical Pharmacology

RESEARCH ARTICLE DOI: 10.53555/jptcp.v31i1.4213

HEAD AND NECK CANCER RISK AND HUMAN PAPILLOMAVIRUS: BRADFORD HILL CRITERIA BASED EVALUATION

Kinza Khan¹, Rizwana Sultan², Syed Qaswar Ali Shah³, Zahid Farooq⁴, Najeeb Ur Rehman⁵, Hafsa Munir⁶, Jamal Muhammad Khan^{7*}

¹Department of Microbiology, Faculty of Veterinary and Animal Sciences, The Islamia University of Bahawalpur - Pakistan

²Department of Pathology, Faculty of Veterinary and Animal Sciences, Cholistan University of Veterinary and Animal Sciences Bahawalpur - Pakistan

³Department of Zoology, Cholistan University of Veterinary and Animal Sciences Bahawalpur - Pakistan

⁴Department of Zoology, Cholistan University of Veterinary and Animal Sciences, Bahawalpur - Pakistan

⁵Department of Zoology, Cholistan University of Veterinary and Animal Sciences, Bahawalpur - Pakistan

⁶Department of Microbiology, Cholistan University of Veterinary and Animal Sciences, Bahawalpur - Pakistan

^{7*}Department of Parasitology, Cholistan University of Veterinary and Animal Sciences, Bahawalpur - Pakistan

*Corresponding Author: Jamal Muhammad Khan

*Department of Parasitology, Cholistan University of Veterinary and Animal Sciences, Bahawalpur - Pakistan, jamalmkhan@cuvas.edu.pk

Abstract

The involvement of human papillomavirus (HPV) has effectively been decoded in Head and neck cancer (HNC) worldwide with contradicting findings. Although the different groups of researchers explored the potential association of HPV with HNC using statistical meta-analysis, however, the association remained still controversial due to the major shortcomings of meta-analysis. Therefore, we arranged the present study to investigate a potential link of HPV with HNC using an additional method (Bradford Hill criteria) which helps to get a more clear picture. Methodology: Initially using PubMed, we extracted all of the studies that associated HPV to HNC. Then, to assess the potential association of HPV with HNC, an examination of the available data on HPV in HNC, normal/benign samples was done using all the major Bradford Hill criteria postulates. Furthermore, to improve the authenticity of our findings, we have also critically evaluated the methodologies of the identified studies to check the possibility of false-negative and false-positive results. Results: After a careful assessment of the previous studies against Bradford Hill criteria postulates, we observed that all the major postulates were not fulfilled. Conclusion: Therefore, our findings recommended no casual association of HPV with HNC.

Key words: Head and neck cancer (HNC); Bradford Hill criteria; Human papillomavirus (HPV)

INTRODUCTION

Head and neck cancers include a diverse variety of tumors that have been clustered together as an entity, based on etiology, anatomy, and sensitivity to chemotherapy (1-3). These malignancies include tumors of the oral cavity, paranasal sinuses, nasal cavity, pharynx, and larynx that arise from the upper aerodigestive tract (4-6). Around 95% of head and neck cancer (HNC) comes from squamous cells and thus are referred to as head and neck squamous cell carcinoma (HNSCC) (7-9). Alcohol consumption and tobacco smoking are classified as the major predisposing factors for developing HNC, but now Human Papillomavirus (HPV), particularly HPV-16, has also been identified to be linked with HNC (7, 10-13).

Keeping in view the role of HPV in the cancer in HNC, various studies have been conducted so far to document the role of HPV in the pathogenesis of HNC but their results were conflicting (14-22). Various group of researchers used statistical meta-analysis to resolve this disagreement and obtain more accurate association between HPV and HNC. However, due to significant limitations of the statistical meta-analysis including inability to critically evaluate the methodologies, providing no information regarding heterogeneity of the studied populations, and publication biasness, the evaluation of a correlation among HPV and HNC is due with an additional strategy.

In our study, we evaluated the correlation among HPV and HNC using Bradford Hill criteria postulates. These postulates are worldwide effective for linking a presumed cause with an effect [20]. In the evaluation, we analyzed the data of previous studies to document, whether or not previous studies met the Bradford Hill criteria postulates to declare a causal association among HPV and HNC. Additionally, to make our outcomes more authentic, we also critically reviewed the methodologies of identified studies to address the propensity of false results.

MATERIAL AND METHODS

In our study, we implemented a two-phase methodology (Fig. 1).

LITERATURE IDENTIFICATION

Related studies associating HPV with HNC were searched via PubMed using the keywords: "Head and neck cancer" AND "Human papillomavirus". We also defined "Retroviridae" AND "Head and neck neoplasm" as medical subject headings (MeSH) terms. All the original articles were searched available till December 2020. In the end, we found a total of 6831 original articles.

RELEVANT DATA ACQUISITION

From 6831 original articles, in total 52 relevant studies were shortlisted which studied the association between HPV and HNC reading their titles, abstract, and the complete article. In addition, a detailed table was built after acquiring the required data from shortlisted studies..

EVALUATION OF THE RESULTS USING BRADFORD HILL CRITERIA POSTULATES

Based on the acquired data, we critically evaluated the selected studies using eight major Bradford Hill criteria postulates:

(1) Strength, (2) Temporality, (3) Consistency, (4) Plausibility, (5) Biological gradient, (6) Experiment, (7) Specificity, and (8) Analogy (23).

The postulate's evaluation was descriptive, with no quantitative assigned score. The evidence for each postulate is given in (Table 1) and results part with a final verdict of whether or not the postulate was fulfilled.

RESULTS

On PubMed, A total of 52 original studies (14-17, 24-71) (Table 1) were identified worldwide that examined the potential link of HPV with HNC. Table 1 summarizes the selected studies and includes the important acquired data from these studies essential for the assessment of Bradford Hill criteria postulates including information of the studied population, names of the technique utilized

for the HPV identification, targeted gene name, name of the HPV detected strain, CI and P values, name of the prevalent identified HPV strain, total analyzed samples count (normal, benign and HNC) with respective population-wide detection positivity ratios.

The positivity ratio of HPV detection in the HNC samples was varied population-wide from 3.33% (14) to 78% (15). While, the positivity ratio of HPV detection in normal and adjacent/benign samples was varied from 0% (33, 52, 55, 57, 64) to 55 (71) and 0% (65) to 82% (17), respectively.

THE EVIDENCE FOR EACH OF THE BRADFORD-HILL POSTULATES STRENGTH

The existence of a weak association does not rule out the possibility of a causal association; however, weak this situation is more likely to be clarified by undetected prejudices. The point that stronger relationships tend to be more causative is rational. In total, 17 case-control studies (16, 17, 24, 30, 33, 34, 36, 48, 52, 55-57, 63-65, 67, 71) were found in the literature reporting association between HPV and HNC. Only 05 (16, 24, 34, 56, 71) of them have reported the CI (16, 24, 34), P-values (56, 71), and higher HPV detection ratio in HNC samples as compare to controls except one study (17), and found a significant association between HPV and HNC in Mexican, Pakistani, Japanese and Chinese populations. However, none of the study reported both CI and P-value. These data overall support a negligible strength of association between HPV and HNC.

CONSISTENCY

Among 17 case-control studies, 15 studies (16, 24, 30, 33, 34, 36, 48, 52, 55-57, 63-65, 71) have reported the higher HPV detection ratios in HNC samples relative to controls while two studies (17, 67) have documented the opposite results. Therefore, consistent findings have not been observed in different populations using different populations strengthening the existence of a non-casual association.

BIOLOGICAL GRADIENT

In certain circumstances, the effect can be the outcome of the minor existence of a factor while, in other cases, generally a greater exposures lead to the higher induction of an effect. Viral load measurements may predict whether HPV differential viral load leads to the differential outcomes in HNC. Unfortunately, no study has reported the HPV viral load either in HNC samples or controls. Therefore, biological gradient postulate was not fulfilled.

TEMPORALITY

Temporality refers to the necessity for HPV to precede HNC. The HPV detection ratios scenario in the current study has shown different outcomes. In total, 11 case-control studies (16, 17, 24, 30, 34, 36, 48, 56, 63, 67, 71) reported that HPV was detected in both normal and HNC samples. Moreover, in two case-control studies (17, 67) HPV detection ratio was higher in normal controls relative to HNC sample. Such conflicting result thus, failed to fulfill the temporal postulates.

PLAUSIBILITY

Plausibility refers to a proper mechanism between cause and effect. HPV is well recognized as a potent inhibitor of TP53 in cervical cancer by making aE6/E6AP/p53 complex, resulting in the degradation of TP53 protein (72). In literature, five studies (35, 43, 57, 68, 72) were found analyzing the association between HPV presence and expression variations in TP53 level: they failed to validate their results. Thus, the role of HPV in the etiology of HNC is biologically not plausible.

EXPERIMENT

This postulate refers to the evidence from either animal or clinical studies. Evidence based on animal models and clinical studies, however, were absent in all the studies found in literature. Therefore, this postulate was not fulfilled.

SPECIFICITY

Causation is possible if a certain population develop HNC in a certain region where the suspected cause is not clarified otherwise. Higher the specificity of the association between a factor and its effect, the more precise the relationship between a factor and its effect. HNC is multi-factorial disease (73) and together with HPV the role of other non-infectious factors and oncogenic viruses (Epstein–Barr virus and John Cunningham virus) in the development of HNC is also well studied worldwide (74-76). Thus, the complexity of the involved factors in HNC development suggested no specificity.

ANALOGY

The similar diseases to HNC that can considered to be HNC analogous are breast cancer and cervical cancer caused by other viral agents like Epstein–Barr virus (EBV), and Mouse mammary tumor virus (MMTV) (77, 78). However, the role of MMTV and EBV in the development of breast cancer and cervical cancer is yet not fully established. Thus, in the present study, the scenario of analogy also suggests no association between HPV and HNC.

DISCUSSION

HNC is the sixth most common types of cancer that affect that affect people all over the world. So far, many studies were conducted worldwide documenting the relationship between HPV and HNC to identify the possible oncogenic pathways regulating by HPV in the development of HNC, however the findings were inconsistent. In addition, a statistical meta-analysis has also been performed by different groups of scientist worldwide to generate a more meaningful relationship between HPV and HNC, due to statistical meta-analysis shortcoming, scientists yet again failed to find a reliable relationship among HPV and HNC. Therefore, in the present study our aim is to find a relationship between HPV and HNC using Bradford Hill criteria postulates.

In total 52 original articles (14-17, 24-71) were included in the present study. The HPV detection ratio reported in these studies was varied between 3.33% (14) to 78% (15) in HNC samples. In most of the case-control studies (16, 24, 30, 33, 34, 36, 48, 52, 55-57, 63-65, 71) the positivity ratio of HPV detection was higher in the HNC samples as compared to the controls while in two studies (17, 67) HPV positivity ratio was higher in the controls as compared to the HNC samples.

Best to our knowledge, no study has applied the Bradford Hill postulates so far to identify the association between HPV and HNC, However, one study utilized these postulates to analyze the causal association between Zika infection and microcephaly, and they suggested no link between the studied parameters (79).

Since, from the initial identification of HPV in HNC, more evidence has become available. We systematically applied Bradford Hill's postulates on the available evidence to find an association between HPV and HNC. The results were not in favor of a casual association. Thus, it was proposed that HPV might combine with the other viruses such as human immunodeficiency virus (HIV) and hepatitis C virus (HCV), and other factors including genetic abnormalities, smoking, alcohol consumption to increase a person's risk of developing HNC by affecting the body's immune system (80).

Moreover, deficiencies as well as and some of the major drawbacks linked with the methodologies of the included studies have been discussed below.

POSSIBLE CAUSES OF FALSE-NEGATIVE RESULTS

Some studies failed to detect HPV presence in any of the cancerous or normal controls they were investigating. How we can be sure that negative results for HPV detection were not due to the poor quality of the extracted DNA? Many studies utilized positive control to avoid such situations (14, 15, 17, 24, 25, 28-30, 33, 34, 37, 38, 40, 44, 46, 47, 50, 62, 63, 65, 66, 69, 71) but few studies (26, 41, 42, 48, 52-54, 56) did not use positive control so there is no way to confirm their negative results. Primers selection targeting L1 and E1 genes of HPV might be inefficient for detecting HPV presence in the advanced carcinoma and thus results in false negative, since L1 and E1 regions

might be lost during viral genome integration with the genome of host, whereas, the E6/E7 regions remained consistently present in any circumstances so, this is the plausible explanation for the negative results of (52, 55, 57, 64, 65) studies.

POSSIBLE CAUSES OF THE FALSE-POSITIVE RESULTS

Most of the studies that we summarized used PCR technique (14-17, 24-30, 33-38, 40-42, 44, 46-54, 56-58, 60, 62-66, 68, 69, 71) for the detection of HPV and none of them utilized second technique to confirm their positive results of PCR, except n = 5 studies (29, 33, 40, 58, 64) which utilized Immunohistochemistry (33, 40, 58), Hybrid capture 2 test (29) and Southern blotting technique (64) and the results of their second techniques have deviated from the first one. In HPV positive HNC patients, expression profiling of various genes such as p14, p16, p53, RB, and others may be used as a surrogate biomarker. Along with HPV detection, these surrogate biomarkers were also analyzed by some studies (27, 28, 33, 35, 43, 44, 46, 49, 54, 57, 58, 68) to further validate their findings, out of which two studies (28, 46) has validated their findings by analyzing p16 as surrogate biomarker while the other studies (27, 33, 35, 43, 44, 49, 54, 57, 58, 68) were failed to validate their findings with surrogate biomarkers. Such deviations in the results of previous studies raise a big question mark on the selection of appropriate technique and their sensitivities.

COMPARISON OF NORMAL, BENIGN AND MALIGNANT SAMPLES

Case-control studies are necessary to establish a causal relationship between the causative agent and the disease. Some of the studies we summarized analyzed only the HNC samples (14, 15, 25-29, 31, 32, 35, 37-47, 49-51, 53, 54, 58-62, 66, 68-70) and did not allow us to compare their results with normal or adjacent/benign controls. However, most of the studies (16, 17, 24, 30, 33, 34, 36, 48, 52, 55-57, 63-65, 67, 71) also analyzed the normal and adjacent/benign tissues along with HNC samples and comparison of their results demonstrated that HPV detection positivity ratios in HNC samples were higher in (16, 24, 30, 33, 34, 36, 48, 52, 55-57, 63-65, 71) studies while lower in two studies (17, 67) as compared to the normal controls. However, no study has found a correlation between HPV and a certain HNC subtype or histologic grade.

CONCLUSION

The results of the present study failed to prove a causal relationship between HPV and HNC. However, due to the limitations of the methodologies used by the previous studies to detect the presence of HPV in HNC, additional experiments are recommended to prove the HPV etiology in HNC.

LIST OF ABBREVIATION

HPV = Human Papillomavirus

HNC = Head and neck cancer

HIV = Human immunodeficiency virus

HCV = Hepatitis C virus

PCR = Polymerase chain reaction

ACKNOWLEDGEMENT

None to declare

CONFLICT OF INTEREST

None to declare

REFERENCES

1. Pfeiffer J, Wiech T, Maier W, Ridder GJ, Laszig R, Birkenhäger R. Head and neck cancer in young adults and nonsmokers: Study of cancer susceptibility by genome-wide high-density SNP microarray mapping. Acta oto-laryngologica. 2011;131(10):1091-8.

- 2. Khan M, Hameed Y. Discovery of novel six genes-based cervical cancer-associated biomarkers that are capable to break the heterogeneity barrier and applicable at the global level. Journal of Cancer Research and Therapeutics. 9000.
- 3. Usman M, Hameed Y, Ahmad M. Does human papillomavirus cause human colorectal cancer? Applying Bradford Hill criteria postulates. ecancermedicalscience. 2020;14.
- 4. Pai SI, Westra WH. Molecular pathology of head and neck cancer: implications for diagnosis, prognosis, and treatment. Annual Review of Pathology: Mechanisms of Disease. 2009;4:49-70.
- 5. Yasir M, Nawaz A, Ghazanfar S, Okla M, Chaudhary A, Al WH, et al. Anti-bacterial activity of essential oils against multidrug-resistant foodborne pathogens isolated from raw milk. Brazilian Journal of Biology. 2022;84:e259449.
- 6. Khalil T, Okla M, Al-Qahtani W, Ali F, Zahra M, Shakeela Q, et al. Tracing probiotic producing bacterial species from gut of buffalo (Bubalus bubalis), South-East-Asia. Brazilian Journal of Biology. 2022;84:e259094.
- 7. Aboagye E, Agyemang-Yeboah F, Duduyemi BM, Obirikorang C. Human papillomavirus detection in head and neck squamous cell carcinomas at a tertiary hospital in sub-Saharan Africa. The Scientific World Journal. 2019;2019.
- 8. Zhang L, Sahar A, Li C, Chaudhary A, Yousaf I, Saeedah M, et al. A detailed multi-omics analysis of GNB2 gene in human cancers. Brazilian Journal of Biology. 2022;84.
- 9. Hameed Y, Ejaz S. TP53 lacks tetramerization and N-terminal domains due to novel inactivating mutations detected in leukemia patients. Journal of Cancer Research and Therapeutics. 2021;17(4):931-7.
- 10. Ullah L, Hameed Y, Ejaz S, Raashid A, Iqbal J, Ullah I, et al. Detection of novel infiltrating ductal carcinoma-associated BReast CAncer gene 2 mutations which alter the deoxyribonucleic acid-binding ability of BReast CAncer gene 2 protein. Journal of Cancer Research and Therapeutics. 2020;16(6):1402-7.
- 11. Hameed Y, Usman M, Liang S, Ejaz S. Novel diagnostic and prognostic biomarkers of colorectal cancer: capable to overcome the heterogeneity-specific barrier and valid for global applications. Plos one. 2021;16(9):e0256020.
- 12. Zhu X, Tang L, Mao J, Hameed Y, Zhang J, Li N, et al. Decoding the mechanism behind the pathogenesis of the focal segmental glomerulosclerosis. Computational and Mathematical Methods in Medicine.2022.
- 13. Usman M, Hameed Y, Ahmad M, Iqbal MJ, Maryam A, Mazhar A, et al. SHMT2 is associated with tumor purity, CD8+ T immune cells infiltration, and a novel therapeutic target in four different human cancers. Current Molecular Medicine. 2023;23(2):161-76.
- 14. de Abreu PM, Có ACG, Azevedo PL, do Valle IB, de Oliveira KG, Gouvea SA, et al. Frequency of HPV in oral cavity squamous cell carcinoma. BMC cancer. 2018;18(1):1-8.
- 15. Attner P, Du J, Näsman A, Hammarstedt L, Ramqvist T, Lindholm J, et al. The role of human papillomavirus in the increased incidence of base of tongue cancer. International journal of cancer. 2010;126(12):2879-84.
- 16. Baig S, Zaman U, Lucky MH. Human papilloma virus 16/18: Fabricator of trouble in oral squamous cell carcinoma. International Journal of Infectious Diseases. 2018;69:115-9.
- 17. García-Milián R, Hernández H, Panadé L, Rodríguez C, González N, Valenzuela C, et al. Detection and typing of human papillomavirus DNA in benign and malignant tumours of laryngeal epithelium. Acta Otolaryngol. 1998 Sep;118(5):754-8.
- 18. Hameed Y, Usman M, Ahmad M. Does mouse mammary tumor-like virus cause human breast cancer? Applying Bradford Hill criteria postulates. Bulletin of the National Research Centre. 2020;44(1):1-13.
- 19. Mao J, Huang X, Okla MK, Abdel-Maksoud MA, Mubarak A, Hameed Z, et al. Risk Factors for TERT promoter mutations with papillary thyroid carcinoma patients: a meta-analysis and systematic review. Computational and Mathematical Methods in Medicine.2022.
- 20. Sial N, Rehman JU, Saeed S, Ahmad M, Hameed Y, Atif M, et al. Integrative analysis reveals methylenetetrahydrofolate dehydrogenase 1-like as an independent shared diagnostic and

- prognostic biomarker in five different human cancers. Bioscience Reports. 2022; 42(1):BSR20211783.
- 21. Hameed A, Condò C, Tauseef I, Idrees M, Ghazanfar S, Farid A, et al. Isolation and characterization of a cholesterol-lowering bacteria from Bubalus bubalis raw milk. Fermentation. 2022;8(4):163.
- 22. Ahmad M, Hameed Y, Khan M, Usman M, Rehman A, Abid U, et al. Up-regulation of GINS1 highlighted a good diagnostic and prognostic potential of survival in three different subtypes of human cancer. Brazilian Journal of Biology. 2021;84.
- 23. Fedak KM, Bernal A, Capshaw ZA, Gross S. Applying the Bradford Hill criteria in the 21st century: how data integration has changed causal inference in molecular epidemiology. Emerging themes in epidemiology. 2015;12(1):1-9.
- 24. Anaya-Saavedra G, Ramírez-Amador V, Irigoyen-Camacho ME, García-Cuellar CM, Guido-Jiménez M, Méndez-Martínez R, et al. High association of human papillomavirus infection with oral cancer: a case-control study. Archives of medical research. 2008;39(2):189-97.
- 25. Balaram P, Nalinakumari KR, Abraham E, Balan A, Hareendran NK, Bernard HU, et al. Human papillomaviruses in 91 oral cancers from Indian betel quid chewers--high prevalence and multiplicity of infections. Int J Cancer. 1995;61(4):450-4.
- 26. Boy S, Rensburg EJV, Engelbrecht S, Dreyer L, Van Heerden M, Van Heerden W. HPV detection in primary intra-oral squamous cell carcinomas—commensal, aetiological agent or contamination? Journal of oral pathology & medicine. 2006;35(2):86-90.
- 27. Cao J, Zhang ZY, Patima, Zhang YX, Chen WT. Human papillomavirus infection and p53 alteration in oral squamous cell carcinoma. Chin J Dent Res. 2000;3(3):44-9.
- 28. Charfi L, Jouffroy T, de Cremoux P, Le Peltier N, Thioux M, Fréneaux P, et al. Two types of squamous cell carcinoma of the palatine tonsil characterized by distinct etiology, molecular features and outcome. Cancer letters. 2008;260(1-2):72-8.
- 29. Chaudhary AK, Pandya S, Mehrotra R, Bharti AC, Singh M, Singh M. Comparative study between the Hybrid Capture II test and PCR based assay for the detection of human papillomavirus DNA in oral submucous fibrosis and oral squamous cell carcinoma. Virology Journal. 2010;7(1):253.
- 30. Chen F, Yan L, Liu F, Huang J, Liu F, Wu J, et al. Oral human papillomavirus infection, sexual behaviors and risk of oral squamous cell carcinoma in southeast of China: a case-control study. Journal of clinical virology. 2016;85:7-12.
- 31. Chernock RD, Zhang Q, El-Mofty SK, Thorstad WL, Lewis JS. Human papillomavirus—related squamous cell carcinoma of the oropharynx: a comparative study in whites and African Americans. Archives of Otolaryngology—Head & Neck Surgery. 2011;137(2):163-9.
- 32. Dillner J, Knekt P, Schiller J, Hakulinen T. Prospective seroepidemiological evidence that human papillomavirus type 16 infection is a risk factor for oesophageal squamous cell carcinoma. Bmj. 1995;311(7016):1346.
- 33. Elango KJ, Suresh A, Erode EM, Subhadradevi L, Ravindran HK, Iyer SK, et al. Role of human papilloma virus in oral tongue squamous cell carcinoma. Asian Pac J Cancer Prev. 2011;12(4):889-96.
- 34. Gan L-L, Zhang H, Guo J-H, Fan M-W. Prevalence of human papillomavirus infection in oral squamous cell carcinoma: a case-control study in Wuhan, China. Asian Pac J Cancer Prev. 2014;15(14):5861-5.
- 35. Hannisdal K, Schjølberg A, De Angelis PM, Boysen M, Clausen OPF. Human papillomavirus (HPV)-positive tonsillar carcinomas are frequent and have a favourable prognosis in males in Norway. Acta oto-laryngologica. 2010;130(2):293-9.
- 36. Hansson BG, Rosenquist K, Antonsson A, Wennerberg J, Schildt E-B, Bladström A, et al. Strong association between infection with human papillomavirus and oral and oropharyngeal squamous cell carcinoma: a population-based case-control study in southern Sweden. Acta oto-laryngologica. 2005;125(12):1337-44.

- 37. Ibieta BR, Lizano M, Frías-Mendivil M, Barrera JL, Carrillo A, Ruíz-Godoy LM, et al. Human papilloma virus in oral squamous cell carcinoma in a Mexican population. Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology. 2005;99(3):311-5.
- 38. Jalouli J, Ibrahim SO, Sapkota D, Jalouli MM, Vasstrand EN, Hirsch JM, et al. Presence of human papilloma virus, herpes simplex virus and Epstein–Barr virus DNA in oral biopsies from Sudanese patients with regard to toombak use. Journal of oral pathology & medicine. 2010;39(8):599-604.
- 39. Kane S, Patil V, Joshi A, Noronha V, Muddu V, Dhumal S, et al. Neoadjuvant Chemotherapy in Technically Unresectable Oral Cancers: Does Human Papillomavirus Make a Difference? Clinical oncology (Royal College of Radiologists (Great Britain)). 2015;27(12):751.
- 40. Kouketsu A, Sato I, Abe S, Oikawa M, Shimizu Y, Takahashi T, et al. Detection of human papillomavirus infection in oral squamous cell carcinoma: a cohort study of Japanese patients. Journal of oral pathology & medicine. 2016;45(8):565-72.
- 41. Kozomara R, Jović N, Magić Z, Branković-Magić M, Minić V. p53 mutations and human papillomavirus infection in oral squamous cell carcinomas: correlation with overall survival. Journal of Cranio-Maxillofacial Surgery. 2005;33(5):342-8.
- 42. Kulkarni SS, Kulkarni SS, Vastrad PP, Kulkarni BB, Markande AR, Kadakol G, et al. Prevalence and distribution of high risk human papillomavirus (HPV) types 16 and 18 in carcinoma of cervix, saliva of patients with oral squamous cell carcinoma and in the general population in Karnataka, India. Asian Pac J Cancer Prev. 2011;12(3):645-8.
- 43. Kumar RV, Kadkol S, Daniel R, Shenoy A, Shah K. Human papillomavirus, p53 and cyclin D1 expression in oropharyngeal carcinoma. International journal of oral and maxillofacial surgery. 2003;32(5):539-43.
- 44. Kuo K-T, Hsiao C-H, Lin C-H, Kuo L-T, Huang S-H, Lin M-C. The biomarkers of human papillomavirus infection in tonsillar squamous cell carcinoma—molecular basis and predicting favorable outcome. Modern Pathology. 2008;21(4):376-86.
- 45. Lai K, Killingsworth M, Matthews S, Caixeiro N, Evangelista C, Wu X, et al. Differences in survival outcome between oropharyngeal and oral cavity squamous cell carcinoma in relation to HPV status. Journal of oral pathology & medicine. 2017;46(8):574-82.
- 46. Lau HY, Brar S, Klimowicz AC, Petrillo SK, Hao D, Brockton NT, et al. Prognostic significance of p16 in locally advanced squamous cell carcinoma of the head and neck treated with concurrent cisplatin and radiotherapy. Head & neck. 2011;33(2):251-6.
- 47. Lee L-A, Huang C-G, Tsao K-C, Liao C-T, Kang C-J, Chang K-P, et al. Human papillomavirus infections are common and predict mortality in a retrospective cohort study of taiwanese patients with oral cavity cancer. Medicine. 2015;94(47).
- 48. Lee S, Cho N, Choi E, Baek S, Kim W, Shin D, et al. Relevance of human papilloma virus (HPV) infection to carcinogenesis of oral tongue cancer. International journal of oral and maxillofacial surgery. 2010;39(7):678-83.
- 49. Lindquist D, Romanitan M, Hammarstedt L, Näsman A, Dahlstrand H, Lindholm J, et al. Human papillomavirus is a favourable prognostic factor in tonsillar cancer and its oncogenic role is supported by the expression of E6 and E7. Molecular Oncology. 2007;1(3):350-5.
- 50. Lingen MW, Xiao W, Schmitt A, Jiang B, Pickard R, Kreinbrink P, et al. Low etiologic fraction for high-risk human papillomavirus in oral cavity squamous cell carcinomas. Oral oncology. 2013;49(1):1-8.
- 51. Mathew A, Mody R, Patait MR, Razooki AA, Varghese NT, Saraf K. Prevalence and relationship of human papilloma virus type 16 and type 18 with oral squamous cell carcinoma and oral leukoplakia in fresh scrappings: A PCR study. Indian journal of medical sciences. 2011;65(5).
- 52. Mellin H, Friesland S, Lewensohn R, Dalianis T, Munck-Wikland E. Human papillomavirus (HPV) DNA in tonsillar cancer: clinical correlates, risk of relapse, and survival. International journal of cancer. 2000;89(3):300-4.

- 53. Nagpal JK, Patnaik S, Das BR. Prevalence of high-risk human papilloma virus types and its association with P53 codon 72 polymorphism in tobacco addicted oral squamous cell carcinoma (OSCC) patients of Eastern India. Int J Cancer. 2002;97(5):649-53.
- 54. Nemes JA, Deli L, Nemes Z, Márton IJ. Expression of p16INK4A, p53, and Rb proteins are independent from the presence of human papillomavirus genes in oral squamous cell carcinoma. Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology. 2006;102(3):344-52.
- 55. Niedobitek G, Pitteroff S, Herbst H, Shepherd P, Finn T, Anagnostopoulos I, et al. Detection of human papillomavirus type 16 DNA in carcinomas of the palatine tonsil. Journal of clinical pathology. 1990;43(11):918-21.
- 56. Nishioka KF, Kazunori Nishizaki, Mehmet Gunduz, Susumu Tominaga, Motoharu Fukazawa, Nobuya Monden, Shuichi Watanabe, Yu Masuda, Hajime Ogura, Shinji. Human papillomavirus as a risk factor for head and neck cancers-a case-control study. Acta Oto-Laryngologica. 1999;119(540):77-80.
- 57. PALLESEN G, AUER G, LINDHOLM J, LINDE A, ÅBERG B, RUBIO C, et al. Involvement of Aberrant p53 Expression and Human Papillo-mavirus in Carcinoma of the Head, Neck and Esophagus. Anticancer research. 1994;14:1281-6.
- 58. Palve V, Bagwan J, Krishnan NM, Pareek M, Chandola U, Suresh A, et al. Detection of highrisk human papillomavirus in oral cavity squamous cell carcinoma using multiple analytes and their role in patient survival. Journal of global oncology. 2018;4:1-33.
- 59. Pillai MR, Phanidhara A, Kesari AL, Nair P, Nair MK. Cellular manifestations of human papillomavirus infection in the oral mucosa. J Surg Oncol. 1999;71(1):10-5.
- 60. Premoli-De-Percoco G, Ramirez JL. High risk human papillomavirus in oral squamous carcinoma: evidence of risk factors in a Venezuelan rural population. Preliminary report. J Oral Pathol Med. 2001;30(6):355-61.
- 61. Premoli-De-Percoco G, Ramírez JL, Galindo I. Correlation between HPV types associated with oral squamous cell carcinoma and cervicovaginal cytology: An in situ hybridization study. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 1998;86(1):77-81.
- 62. Ritchie JM, Smith EM, Summersgill KF, Hoffman HT, Wang D, Klussmann JP, et al. Human papillomavirus infection as a prognostic factor in carcinomas of the oral cavity and oropharynx. International journal of cancer. 2003;104(3):336-44.
- 63. Smith EM, Summersgill KF, McCulloch T, Allen J, Turek LP, Hoffman HT, et al. Human papillomavirus and risk of laryngeal cancer. Annals of Otology, Rhinology & Laryngology. 2000;109(11):1069-76.
- 64. Snijders PJ, Cromme FV, Van Brule AJD, Schrijnemakers HF, Snow GB, Meijer CJ, et al. Prevalence and expression of human papillomavirus in tonsillar carcinomas, indicating a possible viral etiology. International journal of cancer. 1992;51(6):845-50.
- 65. Strome SE, Savva A, Brissett AE, Gostout BS, Lewis J, Clayton AC, et al. Squamous cell carcinoma of the tonsils: a molecular analysis of HPV associations. Clinical Cancer Research. 2002;8(4):1093-100.
- 66. Sugiyama M, Bhawal UK, Kawamura M, Ishioka Y, Shigeishi H, Higashikawa K, et al. Human papillomavirus-16 in oral squamous cell carcinoma: clinical correlates and 5-year survival. British Journal of Oral and Maxillofacial Surgery. 2007;45(2):116-22.
- 67. Syrjänen S, Syrjänen K, Happonen RP. Human papillomavirus (HPV) DNA sequences in oral precancerous lesions and squamous cell carcinoma demonstrated by in situ hybridization. Journal of oral pathology & medicine. 1988;17(6):273-8.
- 68. Tang X, Jia L, Ouyang J, Takagi M. Comparative study of HPV prevalence in Japanese and North-east Chinese oral carcinoma. Journal of oral pathology & medicine. 2003;32(7):393-8.
- 69. Tsuhako K, Nakazato I, Miyagi J, Iwamasa T, Arasaki A, Hiratsuka H, et al. Comparative study of oral squamous cell carcinoma in Okinawa, Southern Japan and Sapporo in Hokkaido, Northern Japan; with special reference to human papillomavirus and Epstein-Barr virus infection. Journal of oral pathology & medicine. 2000;29(2):70-9.

- 70. Xue J, Chen H, Fan M, Zhu F, Diao L, Chen X, et al. Use of quantum dots to detect human papillomavirus in oral squamous cell carcinoma. Journal of oral pathology & medicine. 2009;38(8):668-71.
- 71. Zhang Z-Y, Sdek P, Cao J, Chen W-T. Human papillomavirus type 16 and 18 DNA in oral squamous cell carcinoma and normal mucosa. International journal of oral and maxillofacial surgery. 2004;33(1):71-4.
- 72. Narisawa-Saito M, Kiyono T. Basic mechanisms of high-risk human papillomavirus-induced carcinogenesis: Roles of E6 and E7 proteins. Cancer science. 2007;98(10):1505-11.
- 73. Galbiatti ALS, Padovani-Junior JA, Maníglia JV, Rodrigues CDS, Pavarino ÉC, Goloni-Bertollo EM. Head and neck cancer: causes, prevention and treatment. Brazilian journal of otorhinolaryngology. 2013;79(2):239-47.
- 74. Al-Thawadi H, Gupta I, Jabeen A, Skenderi F, Aboulkassim T, Yasmeen A, et al. Co-presence of human papillomaviruses and Epstein–Barr virus is linked with advanced tumor stage: a tissue microarray study in head and neck cancer patients. Cancer Cell International. 2020;20(1):1-13.
- 75. Deng Z, Uehara T, Maeda H, Hasegawa M, Matayoshi S, Kiyuna A, et al. Epstein-Barr virus and human papillomavirus infections and genotype distribution in head and neck cancers. PLoS One. 2014;9(11):e113702.
- 76. Kutsuna T, Zheng H, Abdel-Aziz HO, Murai Y, Tsuneyama K, Furuta I, et al. High JC virus load in tongue carcinomas may be a risk factor for tongue tumorigenesis. Virchows Arch. 2008;452(4):405-10.
- 77. Zammarchi F, Pistello M, Piersigilli A, Murr R, Cristofano CD, Naccarato AG, et al. MMTV-like sequences in human breast cancer: a fluorescent PCR/laser microdissection approach. The Journal of Pathology: A Journal of the Pathological Society of Great Britain and Ireland. 2006;209(4):436-44.
- 78. Al Dossary R, Alkharsah KR, Kussaibi H. Prevalence of Mouse Mammary Tumor Virus (MMTV)-like sequences in human breast cancer tissues and adjacent normal breast tissues in Saudi Arabia. BMC cancer. 2018;18(1):1-10.
- 79. Awadh A, Chughtai AA, Dyda A, Sheikh M, Heslop DJ, MacIntyre CR. Does Zika virus cause microcephaly-applying the Bradford Hill viewpoints. PLoS currents. 2017;9.
- 80. Song D, Li H, Li H, Dai J. Effect of human papillomavirus infection on the immune system and its role in the course of cervical cancer. Oncology letters. 2015;10(2):600-6.

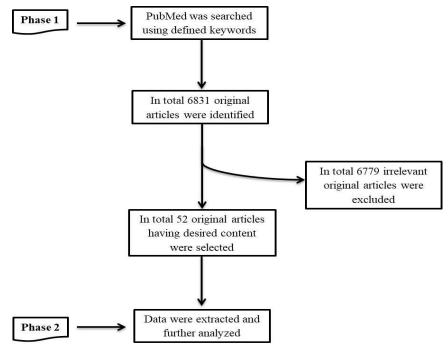


Figure 1: Overview of the methodology implemented during the present study.

Table 1: Summary of the Detection of HPV and positivity rate in normal and HNC samples relative to the different selected articles.

Studied Population	Technique used for viral genome detection	Prevalent strain	Number of the normal sample screened	Percentage positivity of HPV in normal samples (%)	Number of the adjacent or benign samples screened	Percentage positivity of HPV in adjacent or benign samples (%)	Number of the total head and neck cancer samples screened	Percentage positivity of HPV in head and neck cancer samples (%)	References	P- value	CI (%)
United kingdom	In situ hybridization	16	30	0	0	0	28	21	(55)		
Hungary	PCR	16	0	0	0	0	79	41.7	(54)		
Netherlands	PCR, Southern blotting	16	7	0	0	0	10	50	(64)		
Sweden	PCR, ELISA	16	2	0	0	0	34	11	(57)		
	PCR	16	10	0	0	0	60	43	(52)		
	PCR	16	320	4.4	0	0	85	20	(36)		
	PCR	16	0	0	0	0	87	78	(15)		
	RT-PCR	16	0	0	0	0	203	49	(49)		
	PCR		0	0	0	0	72	20.8	(38)		
Brazil	PCR.	16	0	0	0	0	90	3.33	(14)		
Finland	ELISA	16	0	0	0	0	66	9	(32)		
	In situ hybridization	16	0	ő	21	28.6	51	11.8	(67)		
Cuba	PCR, Southern blotting	16	25	16	29	82	33	48.5	(17)	0.05	
Japan	PCR	16	70	4.2	0	0	74	16.2	(56)	<0.05	
	PCR.	16, 18	0	0	0	0	24	54.1	(40)		
	PCR.	16	0	0	ō	0	66	36	(66)		
	PCR, in situ hybridization	16, 18	ő	ő	ů o	0	101	57.4	(69)		
	PCR, immunohistochemistry	16, 18	0	0	Ö	0	60	55.0	(68)		
USA	PCR.	16	12	16.7	ő	0	44	25	(63)		
USA	PCR.	16	0	0	48	0	108	46	(65)		
	In situ hybridization		0	0	0	0	174	56	(31)		
	PCR.	16	0	0	Ö	0	94	10.6	(62)		
	PCR	16	0	0	0	0	409	5.9	(50)		95
Pakistan	PCR	16, 18	200	4	0	0	100	46	(16)		95
Norway	PCR.	16, 18	0	0	0	0	137	51.8	(35)		93
	PCR PCR		0		0	0	50				-
Venezuela				0				60	(60)		
A 4 11	In situ hybridization		0	0	0	0	50 95	70 61.1	(61)		
Australia	Immunohistochemistry			0		0			(45)		
Korea	PCR.	16	25	4	0		36	36	(48)		
China	PCR.	18	189	3.1	0	0	178	14.04	(30)		
	PCR	16, 18	68	2.94	0	0	200	27.5	(34)		95
	Quantum dots, in situ hybridization	16, 18	0	0	0	0	21	33.3	(70)		
	PCR	16	0	0	0	0	1002	19.4	(47)		
	PCR.	16, 18	0	0	0	0	40	72.5	(27)	0.037	
	PCR.	16, 18	40	55	0	0	73	73.9	(71)	0.040	
India	PCR.	16, 18	0	0	0	0	91	41	(25)		
	PCR, Hybrid Capture II test	16	0	0	0	0	222	32.4	(29)		
	Immunohistochemistry		0	0	0	0	124	12.9	(39)		
	PCR	16	46	0	0	0	60	48	(33)		
	qPCR.	16, 18	0	0	0	0	106	33	(58)		
	Immunohistochemistry, In situ hybridization		0	0	0	0	42	31	(43)		
	PCR.	16, 18	0	0	0	0	110	33.6	(53)		
	PCR	16, 18	0	0	0	0	34	70.6	(42)		
	PCR	16, 18	0	0	0	0	45	77.3	(51)		
	In situ hybridization		0	0	0	0	61	27.8	(59)	0.008	
										9	T
South Africa	PCR	18	0	0	0	0	59	11.8	(26)		
Taiwan	PCR	16	0	0	0	0	92	75	(44)		
	PCR		0	0	0	0				_	_
France		16					52	62	(28)		
Canada	PCR, Immunohistochemistry	16	0	0	0	0	55	58	(46)		
Serbia	PCR	16	0	0	0	0	50	64	(41)		
Mexico	PCR	16, 18	248	17.3	0	0	62	43.5	(24)		95
	PCR	16	0	0	0	0	51	42	(37)		

PCR = Polymerase chain reaction