



PATHOPHYSIOLOGY OF EPILEPTOGENESIS: A COMPREHENSIVE REVIEW

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Abstract

There are several neurological disorders that can cause recurrent seizures. Epilepsy is one of them, and is characterized by a persistent predisposition to seizures caused by abnormal neuronal activity in the brain. The relative imbalance between excitatory and inhibitory neurotransmitters may result in epileptic seizures. Pathogenesis of epilepsy can be influenced by changes in the expression of receptors and ion channels regulated by neurotransmitters. A neurotransmitter metabolism error affects the synthesis, breakdown, transport or the cofactors of neurotransmitters. The impairment of neuronal receptors, intracellular signaling, vesicle release, or other synaptic abnormalities can also cause neurotransmitter dysfunction. The main clinical hallmark of some diseases is epilepsy. The purpose of this comprehensive review is to describe the epileptogenic mechanisms as well as the implications arising from mutations in neurotransmitter-mediated receptors and ion channels in epilepsy.

Keywords: Epileptogenesis, Epilepsy, Neurotransmitters, Pathophysiology, Signaling

INTRODUCTION

Epilepsy is a neurological disorder caused by the repeated occurrence of seizures resulting from recurrent, spontaneous, and abnormal electrical discharge from a group of neurons in the brain (Vezzani et al., 2008). The term “epilepsy” is derived from the Greek word “epilambanein”, which means “to seize upon” or “to attack” (Patel et al., 2019). In Ayurveda, epilepsy is defined as Apasmara: APA, meaning negation or loss of; smara, meaning recollection or consciousness (Brennan et al., 2018). The estimated population with epilepsy is between 4-10 per 1000 people. However, in low and middle-income countries this proportion is much higher, between 7 and 14 per 1000 people. Each year, approximately 2.4 million people are globally diagnosed with epilepsy (Lang et al., 2022; Beghi et al., 2023). With a conservative estimate of 1% prevalence of epilepsy, there are more than 12 million persons with epilepsy in India, which contributes to nearly one-sixth of the global burden (Chaunsali et al., 2020). Epilepsy belongs to a group of chronic disorders characterized by the periodic or transistor bursting of excessive electric discharge produced by sudden and recurrent

events of seizures. Seizures are proximal events of abnormal discharge of NTs (Victor et al., 2020). Both seizures and epilepsy are interchangeable. Epilepsy has been diagnosed after two unprovoked seizures occurring in twenty-four hours. Seizures had classified according to incidence, characteristics, and clinical signs (Moore et al., 2021). seizures had divided into two broad classes of 'partial' and 'generalized' seizures. A recent classification of seizures proposed by the International League Against Epilepsy (ILAE) in 2017 based on clinical manifestations is characterized into three groups: seizures, epilepsies, and epileptic syndrome but the old system is still running due to some limitations and facing some high criticism. Different type of seizures demonstrates different symptoms, and it also varies from person to person (Sharma et al., 2019).

Based on the meta-analysis of GBD, the prevalence rate was slightly higher (50 million) in 1990 than in 2016. Focal Seizures (Partial Seizures) are very prevalent as compared to generalized seizures (Lang et al., 2022). The age-standardized prevalence rate increased by 13.6% from 1990 to 2017. The incidence rate of epilepsy was 61.4 per 1,00,000 per year. Globally the mortality rate of epilepsy was 125000. India has an extensive burden of epilepsy because about 10-12 million people live with epilepsy, and 3 million population suffer from drug-resistant epilepsy. A large number of cases of epilepsy in India are due to developmental delay. The mortality rate is 5 to 10 times greater than the general population (Begley et al., 2022).

Epilepsy affects the different parts of the brain by hyperexcitation and hyper synchronization of brain cells that why epilepsy is called a brain disorder (Gasparini et al., 2019; Fukuyama et al., 2020). Epileptic seizures are appearing in the cortical and subcortical parts of the brain (Engelborghs et al., 2000). In a normal brain, excitatory synaptic activity is tightly regulating the inhibitory activity of the neurons. Some specific mechanisms are responsible for the production of seizures. Epilepsy arises due to the sudden imbalance and shifting of excitatory and inhibitory signaling of the neurons in the brain, as show in fig 1 (Falco-Walter et al., 2020). Due to this modulation in excitatory and inhibitory NTs causes an increase in the Na^+ or Ca^{2+} influx and a decrease in the Cl^- influx that produces depolarization of the neurons, increases the action potential of the neurons, or stimulates excessive neuronal firing in the brain (Falco-Walter et al., 2020; Fukuyama et al., 2020). Hyperexcitation of Cortical neurons is observed under certain conditions (trauma, TBI, viral infection, high fever, genetic mutation) (Russo et al., 1981). Understand some other pathways and factors which take part in the dysregulation of NTs and ion channels, as show in fig 2.

1. NEUROTRANSMITTERS IN EPILEPTOGENESIS

Several NTs are present in the brain that plays a very crucial role in physical, genetic, behavioral, and psychological development. Dereglulation of NTs causes several diseases from them one is epilepsy (Dhaher et al., 2021). Loss of NTs causes hypersynchronisation of neurons and these neurons can produce GS and PS. Abnormal discharge of NTs produce sign and symptoms with alternation in motor function, sensation, autonomic functions, behavioral functions, and consciousness (Mastrangelo et al., 2021; Sarikaya et al., 2015). Some NTs are involved in the production of excitation. On the other hand, some NTs act as inhibitory NTs, as show in fig 3 (Arafa et al., 2013).

1) GABA

GABA receptor act as an antagonist in epilepsy (Palma et al., 2017). Seizures are generated when the inhibitory GABAergic action decreases. Convulsive and epileptic agents block GABA mediatory inhibition. Repetitive activation of cortical neurons can reduce the inhibitory postsynaptic potential of neurons that decreases the release of GABAergic action (Jacob et al., 2016; Wasterlain et al., 1993). During hypoxia and ischemia, GABA receptors coupled with their agonist produce alteration in ionic gradient, prolong the opening of the Cl^- channel, or increase the conductance of Cl^- ion in the neurons due to accumulation of Cl^- in the brain cell. Increased concentration of Cl^- affects the redistribution of Cl^- and K^+ transport (Mathern et al., 1999). A low level of GABA and GAD (Glutamic acid decarboxylase) is observed in epilepsy. Epilepsy and epilepsy-associated disease decrease the 3H receptor and BDZ receptors binding site which causes an increase in the frequency of seizures. A low concentration of GABA in CSF is increasing the episodes of seizures

(Sgadò et al., 2011). Several endogenous and exogenous substances are involved in the inhibition of GABAergic transmission via make interaction with GABA receptors or inhibition of GABA synthesis (Sarnat et al., 2021).

2) Glutamate

Glutamate receptors act as an agonist and can elicit seizures in the normal brain (Sandhu et al., 2021). Glutamate consists of two types of receptors 1) Ionotropic receptor (iGluR) and 2) Metabotropic receptors [G- protein-coupled receptor (mGluR) (Chiprés-Tinajero et al., 2021). Overexpression of iGluR and mGluR increases the Ca^{2+} influx into the neuronal membrane and increases the excitation of a neuronal membrane (Kovalenko et al., 2022; Sandhu et al., 2021). Metabotropic receptors (GPCR) are also involved in the production of neuronal excitation via the regulation of secondary messenger cAMP and modulate the activity of synapsis (Sarło et al., 2021). NMDAR is a subtype ionotropic receptor of glutamate. It plays a crucial role in the development and generation of neuronal excitability in CNS. NMDARs can induce seizures and selective exocytotic cell death of hippocampal neuronal cells. NMDA may release Ca^{2+} from ER (endoplasmic reticulum) via activates of the mGluR that increase the level of Na^+ and Ca^{2+} . It is able to prolong the opening of the Ca^{2+} channel that cause increase in the release of glutamate (Kovalenko et al., 2021; Sarło et al., 2021).

a) Ionotropic receptor

Neuronal activity increases the release of glutamate in the synapses. NMDA receptors become activated when glutamate is coupled with the NMDA receptors. Activated NMDA receptors increase the Ca^{2+} influx in the synapsis that causes the activation of CaMKII and calcineurin (Akyuz et al., 2021; Needs et al., 2019). Activated CaMKII stimulates the endocytosis of a subtype of AMPA receptor (GluR1) that produced long-term potential excitation of synapsis. Activated calcineurin bind with β 2/3 or γ 2 subunit of GABAA receptors that promote the endocytosis of GABAA receptors and decrease the GABA-mediated inhibition in the synapsis that involve the reduction of expression and function of Chloride potassium symporter 5 (KCC2) (Rossi et al., 2022; Sears et al., 2021). Overexpression of KCC2 modulates the equilibrium of Cl^- potential. Overexpression of NMDA receptors in the synapsis increases the influx of Ca^{2+} that causes the deregulation of CaMKII activity and causes epileptogenesis, as show in fig 4 (Kovalenko et al., 2022; Needs et al., 2019).

b) Metabotropic receptor

Activation of specific group I mGluR promote the epileptogenesis in CA3 pyramidal cell. Group 1 GluR agonist activates the mGluR1/5 (a subtype of group I mGluR) (Cingolani et al., 2019) in the CA3 pyramidal cell of the hippocampus. Activated mGluR1/5 promotes the dissociation of heterotrimeric G protein in $\text{G}\alpha$ and $\text{G}\beta\gamma$, or activation of Src and ERK1/2 MAPK (Cingolani et al., 2019; Wong et al., 2002) Dissociated $\text{G}\alpha$ activates the $\text{PLC}\beta$ (Phospholipase C- β) that involve in the hydrolysis of PIP_2 into IP_3 and DAG. IP_3 increases the release of Ca^{2+} from ER and DAG increases the activity of PKC (Protein Kinase C). Dissociated $\text{G}\beta\gamma$ promotes the reduction of K^+ conductance and increases the voltage dependent intrinsic neuronal activity, as show in fig 2 (Kłodzińska et al., 1999; Qian et al., 2016).

2. ASTROGLIOSIS IN EPILEPTOGENESIS

TBI is the most common cause of epilepsy because the injured area has recovered by reactive astrogliosis. About 14-20% of patients with TBI have a chance to develop epilepsy (Golub et al., 2020). It is a star-shaped glial cell that consists of 30% part of the CNS and provides metabolic and physical support to the neurons. It is found nearly to the synapsis and works as an NTs, transporter, receptor, and ion channel (Song et al., 2022). As we know, astrogliosis regulates the development, plasticity, and hyperexcitability of the neurons, and it also controls the release of NTs. It converted into reactive astrogliosis after the morphological and genetic changes. Some studies demonstrate that alternation in morphology and genetic pattern can cause epilepsy (Song et al., 2020; Yang et al., 2021). Astrocyte proliferation increases after injury because a shield of the scar has formed around

the injured area. The glial scar seals the injured area and protects the healthy area of the brain from damage by preventing the influx of harmful substances (Fukuyama et al., 2022). TBI produces cerebral ischemia after the scar formation that increases the production and accumulation of lactic acid in the brain causing modulation and elevation of excitatory ions and NTs (Verhoog et al., 2020; Sano et al., 2021).

Modulation of NTs increases the glutamate level in the brain and increases the production of ROS species (Sandhu et al., 2021). Overstimulation of glutamate levels causes an increased extracellular NMDA-mediated Ca^{2+} level. Elevated Ca^{2+} concentration in the brain causes hyperexcitability and develops a seizure, as shown in fig 2. The proliferation of astrocytes also plays a crucial role in the generation of inflammation. Astrocytes elevate the liberation of inflammatory mediators such as $\text{TNF-}\alpha$ and $\text{IL-1}\beta$ that activate the NMDA receptors that enhance NMDA and induce Ca^{2+} influx. Excessive Ca^{2+} influx involves the production of ROS that cause hyperexcitability of neurons that produce episodes of epilepsy (Fukuyama et al., 2022; Verhoog et al., 2020).

3. CYTOKINES IN EPILEPTOGENESIS

Cytokines are prototypic inflammatory mediators. It consists of $\text{TNF-}\alpha$, $\text{IL-1}\beta$, IL-6 , $\text{NFK}\beta$, chemokines, an acute-phase protein, and a complementary system. It produces neurotoxicity via autocrine and paracrine transporter mechanisms. Epileptic patients have a high level of cytokines in serum and CSF (Soltani et al., 2022; Vishwakarma et al., 2022).

Interleukine-1 consist three ligands such as: $\text{IL-1}\alpha$, $\text{IL-1}\beta$ and IL-1Ra . The level of IL-1 is low in the CNS. In certain pathological conditions, the level of IL-1 has elevated. $\text{IL-1}\beta$ is responsible for ROS generation and takes part in excitability (Yazdanpanah et al., 2022). Cytokines $\text{IL-1}\beta$ are located on hippocampal pyramidal neurons along with NMDA receptors. $\text{IL-1}\beta$ activates the sphingomyelinase and Src kinase in the hippocampus and, they provoke phosphorylation of the NR2B subunit of the NMDA receptor that produces NMDA-mediated Ca^{2+} influx that causes hyperexcitability (Vishwakarma et al., 2022; Numis et al., 2019). $\text{IL-1}\beta$ inhibits the glutamate reuptake that increases the glial release via $\text{TNF-}\alpha$ production. $\text{TNF-}\alpha$ is (a tumor necrosis factor) present in the hippocampus, and it works as a dichromate. It activates the p55 and p75 receptors (Chen et al., 2023). $\text{TNF-}\alpha$ modulates the chemical balance of the AMPA receptor on synapsis that increases the excitability through the interaction of the p55 receptor that activates the apoptosis signal-regulating kinase-1 is a key event in the death of neurons and also prevents GLuR2 subunit at the neuronal membrane that produces Ca^{2+} influx into the neurons that causes hyperexcitability (Soltani et al., 2022). $\text{TNF-}\alpha$ increases extracellular glutamate levels, and it also increases the glutamate release via activation of NO synthetase in astrocytes that produces hyperexcitability. It also activates the IL-1R1 which may trigger the $\text{NF-}\kappa\text{B}$. $\text{NF-}\kappa\text{B}$ directly takes part in the production of Pro IL-6 (that stimulates the IL-6) and NO (Atabakin et al., 2023; Aulická et al., 2022; Meng et al., 2020).

IL-6 receptor complex consists of a unit of IL-6R and two molecules of gp130 (glycoprotein130) transmembrane protein found in all classes of cytokines receptors. When IL-6 is expressed in the brain acts as an agonist of glutamate and drops the concentration of GABA that leads to a decrease in the GABA-mediated inhibition and causing depolarization of neurons (Castañeda et al., 2020; Korotkov et al., 2020). HMGB1 (high-mobility group box-1) is release signals in dangerous conditions like injury or stress. HMGB1 activates several kinases (MAPK, PKA, PKC) that alter voltage-dependent K^+ , Na^+ , and Ca^{2+} channels rapidly and produce neuronal excitability in dangerous conditions (Castañeda et al., 2020).

a) NEUROPEPTIDES IN EPILEPTOGENESIS

It consists of two types of receptors P1 and P2. Out of them, P2 takes part in epileptogenesis. P2 receptors are further subdivided into P2X and P2Y. The subclass of P2X is P2X7 that express in neurons and glial cells of the brain. P2X7 is an ATP-gated non-selective cation permeable ionotropic receptor (Tekgul et al., 2020). When ATP converts into ADP activates the P2X7 receptor. This is a key event in the stimulation of microglia, modulates neuronal excitability in the hippocampus, and produces a neuroinflammatory response (Tekgul et al., 2020; Cui et al., 2019). These events elevate

the damage in the brain and generate excitability of neurons or alter the ion channels. On the other side, P2Y receptor subclasses bind to the different G-protein coupled receptors and exert their action. P2Y_{1,2,4,6,11} receptors bind with Gq/G11 receptors that activate the phospholipase C and IP3 activity and increase the Ca influx into the cell. P2Y_{12,13,14} receptors coupled with Gi/o inhibit adenylyl cyclase and modulate the ion channel. Adenylyl cyclase activates the cyclic AMP that produces neuronal excitability (Santos et al., 2023; Wang et al., 2021; Yeo et al., 2022).

b) REACTIVE OXYGEN SPECIES (ROS) IN EPILEPTOGENESIS

Naturally, oxygen is present in divalent forms (unreactive form) and univalent forms (reactive form). The univalent oxygen species has generated from the atoms or groups of atoms that have a tendency to accept electrons and consist of unpaired electrons. This univalent species is known as reactive oxygen species (ROS) (Godinho et al., 2021). The single oxygen species, superoxide, hydroxyl ion, free radicals, or hydrogen peroxide, all are ROS. The brain is the hub of the generation of ROS (Terrone et al., 2020; Devi et al., 2008). ROS generated from the non-enzymatic process (UV irradiation) as well as the enzymatic process (ROS generated in the cell due to modulation in enzyme activity). The frequency of ROS production has to depend on the presence or availability of oxygen and the number of ROS-producing enzymes in the tissue. ROS generated by NOX (NADPH oxidase), NADPH, mitochondria, xanthine oxidase, and lipoxygenase as shown in fig 4 (Terrone et al., 2019; Walker et al., 2023; Xu et al., 2022).

The brain generates ROS in a resting state, and the production of ROS has increased along with the activity of the brain. Generally, ROS generated when the imbalance between free radicals and antioxidants has occurred (Godinho et al., 2021; Walker et al., 2023). The mechanism of ROS production has started when the Ca²⁺ and Na⁺ enter via NMDA receptors into the cell. Several disease conditions may enhance the level of Ca²⁺ and Na⁺ in the cell which causes it to convert NADPH to NADP⁺ by activation of NOX. NADP⁺ takes part in the production of ROS (PATEL et al., 2022). Both ROS and NADP⁺ convert NO (nitric acid) into the reactive form OONO⁻ (peroxynitrite) and OONO⁻ causing the damage of DNA, inactivation of the enzyme, and lipid peroxidation (Frantseva et al., 2000). Excessive efflux of Ca²⁺ in the cell with ROS causes the depolarization of the mitochondrial membrane via activation of DNA repair enzymes [Poly ADP ribose (PAR), PAR polymerase (PARP)] that decrease the level of NADPH which results from the decrease in the production of ATP (Devi et al., 2008, Puttachary et al, 2015). The deficiency of ATP produces energy failure and imbalances the cellular ionic gradients. ROS production and accumulation of Ca²⁺ ion into the mitochondria result from the formation of mitochondrial permeability transition pore (mPTP) that disturb the function of mitochondria and permit the movement of cytochrome C into the cytosol that causes cell death (apoptosis) (Frantseva et al., 2000; Roma-Mateo et al., 2015). In mitochondria complex, I and complex III are the primary sites for the production of ROS. They produce ROS by electron leak mechanism from ETC (electron transport chain). ROS decreases the production of ATP by inhibiting complex I, which causes a decrease in the membrane potential of mitochondria and a decrease in the production of ATP (Puttachary et al, 2015). Complex III is mainly involved in the production of superoxide ion and directly contribute to the production of ROS (Geronzi et al., 2018; Méndez-Armenta et al., 2014).

CONCLUSION

Epileptogenic changes in the brain are caused by inflammation and increased neurogenic activity post-seizure. The contribution of microglia to this process needs to be better understood in order to control it. This balance is largely mediated by glutamate and gamma-aminobutyric acid (GABA); abnormal changes in these molecules may result in irreversible neuronal damage. Neurotransmitter systems and ion channels play a crucial role in neuronal excitability. This will contribute to the development of still more specific and efficient therapeutic interactions in the area of clinical neurology by understanding epilepsy pathophysiology and epileptogenesis.

REFRENECE

1. Akyuz, E., Polat, A. K., Eroglu, E., Kullu, I., Angelopoulou, E., & Paudel, Y. N. (2021). Revisiting the role of neurotransmitters in epilepsy: An updated review. *Life sciences*, 265, 118826.
2. Arafa, N. M., Abdel-Rahman, M., El-khadragy, M. F., & Kassab, R. B. (2013). Evaluation of the possible epileptogenic activity of ciprofloxacin: the role of *Nigella sativa* on amino acids neurotransmitters. *Neurochemical research*, 38, 174-185.
3. Atabaki, R., Khaleghzadeh-Ahangar, H., Esmaeili, N., & Mohseni-Moghaddam, P. (2023). Role of pyroptosis, a pro-inflammatory programmed cell death, in epilepsy. *Cellular and Molecular Neurobiology*, 43(3), 1049-1059.
4. Aulická, S., Česká, K., Šána, J., Siegl, F., Brichtová, E., Ošlejšková, H., ...& Nestršil, I. (2022). Cytokine-chemokine profiles in the hippocampus of patients with mesial temporal lobe epilepsy and hippocampal sclerosis. *Epilepsy Research*, 180, 106858.
5. Beghi, E., Giussani, G., Costa, C., DiFrancesco, J. C., Dhakar, M., Leppik, I., ...& ILAE Task Force on Epilepsy in the Elderly (2017–2021). (2023). The epidemiology of epilepsy in older adults: A narrative review by the ILAE Task Force on Epilepsy in the Elderly. *Epilepsia*.
6. Begley, C., Wagner, R. G., Abraham, A., Beghi, E., Newton, C., Kwon, C. S., ...& Winkler, A. S. (2022). The global cost of epilepsy: a systematic review and extrapolation. *Epilepsia*, 63(4), 892-903.
7. Brennan, G. P., & Henshall, D. C. (2018). microRNAs in the pathophysiology of epilepsy. *Neuroscience letters*, 667, 47-52.
8. Castañeda-Cabral, J. L., Ureña-Guerrero, M. E., Beas-Zárate, C., Colunga-Durán, A., Nuñez-Lumbreras, M. D. L. A., Orozco-Suárez, S., ...& Rocha, L. (2020). Increased expression of proinflammatory cytokines and iNOS in the neocortical microvasculature of patients with temporal lobe epilepsy. *Immunologic Research*, 68, 169-176.
9. Chaunsali, L., Tewari, B. P., & Sontheimer, H. (2021). Perineuronal net dynamics in the pathophysiology of epilepsy. *Epilepsy Currents*, 21(4), 273-281.
10. Chen, Yu, Marwa M. Nagib, Nelufar Yasmen, Madison N. Sluter, Taylor L. Littlejohn, Ying Yu, and Jianxiong Jiang. "Neuroinflammatory mediators in acquired epilepsy: an update." *Inflammation Research* 72, no. 4 (2023): 683-701.
11. Chiprés-Tinajero, G. A., Núñez-Ochoa, M. A., & Medina-Ceja, L. (2021). Increased immunoreactivity of glutamate receptors, neuronal nuclear protein and glial fibrillary acidic protein in the hippocampus of epileptic rats with fast ripple activity. *Experimental Brain Research*, 239(6), 2015-2024.
12. Cingolani, L. A., Vitale, C., & Dityatev, A. (2019). Intra-and extracellular pillars of a unifying framework for homeostatic plasticity: a crosstalk between metabotropic receptors and extracellular matrix. *Frontiers in Cellular Neuroscience*, 13, 513.
13. Cui, Z., Zhang, X., Song, H., Yang, F., Feng, S., Feng, L., ...& Xu, B. (2019). Differential long non-coding RNA (lncRNA) profiles associated with hippocampal sclerosis in human mesial temporal lobe epilepsy. *International Journal of Clinical and Experimental Pathology*, 12(1), 259.
14. Devi, P. U., Manocha, A., & Vohora, D. (2008). Seizures, antiepileptics, antioxidants and oxidative stress: an insight for researchers. *Expert Opinion on Pharmacotherapy*, 9(18), 3169-3177.
15. Dhaher, R., Gruenbaum, S. E., Sandhu, M. R. S., Ottestad-Hansen, S., Tu, N., Wang, Y., ...& Eid, T. (2021). Network-related changes in neurotransmitters and seizure propagation during rodent epileptogenesis. *Neurology*, 96(18), e2261-e2271.
16. Engelborghs, S., D'hooge, R., & De Deyn, P. P. (2000). Pathophysiology of epilepsy. *Actaneurologica belgica*, 100(4), 201-213.
17. Falco-Walter, J. (2020, December). Epilepsy—definition, classification, pathophysiology, and epidemiology. In *Seminars in neurology* (Vol. 40, No. 06, pp. 617-623). Thieme Medical Publishers, Inc..

18. Frantseva, M. V., Velazquez, J. P., Tsoraklidis, G., Mendonca, A. J., Adamchik, Y., Mills, L. R., ... & Burnham, M. W. (2000). Oxidative stress is involved in seizure-induced neurodegeneration in the kindling model of epilepsy. *Neuroscience*, 97(3), 431-435.
19. Fukuyama, K., & Okada, M. (2022). Brivaracetam and levetiracetam suppress astroglial l-glutamate release through hemichannel via inhibition of synaptic vesicle protein. *International Journal of Molecular Sciences*, 23(9), 4473.
20. Fukuyama, K., & Okada, M. (2022). High frequency oscillations play important roles in development of epileptogenesis/ictogenesis via activation of astroglialsignallings. *Biomedicine & Pharmacotherapy*, 149, 112846.
21. Fukuyama, K., Fukuzawa, M., Shiroyama, T., & Okada, M. (2020). Pathogenesis and pathophysiology of autosomal dominant sleep-related hypermotor epilepsy with S284L-mutant $\alpha 4$ subunit of nicotinic ACh receptor. *British Journal of Pharmacology*, 177(9), 2143-2162.
22. Gasparini, S., Ferlazzo, E., Sueri, C., Cianci, V., Ascoli, M., Cavalli, S. M., ...& Epilepsy Study Group of the Italian Neurological Society. (2019). Hypertension, seizures, and epilepsy: a review on pathophysiology and management. *Neurological Sciences*, 40, 1775-1783.
23. Geronzi, U., Lotti, F., &Grosso, S. (2018). Oxidative stress in epilepsy. *Expert Review of Neurotherapeutics*, 18(5), 427-434.
24. Godinho, A. M. G. (2021). *Exploring the activation of NLRP3 inflammasome by reactive oxygen species (ROS) in a model of epileptogenesis* (Doctoral dissertation).
25. Golub, V., & Reddy, S. (2020). Anatomical Basis of Epileptogenesis in a Mouse Model of Traumatic Brain Injury. *The FASEB Journal*, 34(S1), 1-1.
26. Jacob, J. (2016). Cortical interneuron dysfunction in epilepsy associated with autism spectrum disorders. *Epilepsia*, 57(2), 182-193.
27. Kłodzińska, A., Chojnacka-Wójcik, E., &Pilc, A. (1999). Selective group II glutamate metabotropic receptor agonist LY354740 attenuates pentetrazole-and picrotoxin-induced seizures. *Polish journal of pharmacology*, 51(6), 543-545.
28. Korotkov, A., Broekaart, D. W., Banchaewa, L., Pustjens, B., van Scheppingen, J., Anink, J. J., ... &Aronica, E. (2020). microRNA-132 is overexpressed in glia in temporal lobe epilepsy and reduces the expression of pro-epileptogenic factors in human cultured astrocytes. *Glia*, 68(1), 60-75.
29. Kovalenko, A. A., Zakharova, M. V., Schwarz, A. P., Dyomina, A. V., Zubareva, O. E., &Zaitsev, A. V. (2022). Changes in Metabotropic Glutamate Receptor Gene Expression in Rat Brain in a Lithium–Pilocarpine Model of Temporal Lobe Epilepsy. *International Journal of Molecular Sciences*, 23(5), 2752.
30. Kovalenko, A. A., Zakharova, M. V., Schwarz, A. P., Dyomina, A. V., Zubareva, O. E., &Sarnat, H. B., & Flores-Sarnat, L. (2021). Excitatory/inhibitory synaptic ratios in polymicrogyria and down syndrome help explain epileptogenesis in malformations. *Pediatric Neurology*, 116, 41-54.
31. Lang, J. D., &Hamer, H. M. (2022). Epidemiology of epilepsy in old age–English Version. *ZeitschriftfürEpileptologie*, 1-4.
32. Mastrangelo, M. (2021). Epilepsy in inherited neurotransmitter disorders: Spotlights on pathophysiology and clinical management. *Metabolic Brain Disease*, 36(1), 29-43.
33. Mathern, G. W., Mendoza, D., Lozada, A., Pretorius, J. K., Dehnes, Y., Danbolt, N. C., ... &Adelson, P. D. (1999). Hippocampal GABA and glutamate transporter immunoreactivity in patients with temporal lobe epilepsy. *Neurology*, 52(3), 453-453.
34. Méndez-Armenta, M., Nava-Ruíz, C., Juárez-Rebollar, D., Rodríguez-Martínez, E., &Yescas Gómez, P. (2014). Oxidative stress associated with neuronal apoptosis in experimental models of epilepsy. *Oxidative medicine and cellular longevity*, 2014.
35. Meng, F., & Yao, L. (2020). The role of inflammation in epileptogenesis . *ActaEpileptologica*, 2(1), 1-19.

36. Moore, J. L., Carvalho, D. Z., St Louis, E. K., & Bazil, C. (2021). Sleep and epilepsy: a focused review of pathophysiology, clinical syndromes, co-morbidities, and therapy. *Neurotherapeutics*, 18, 170-180.
37. Needs, H. I., Henley, B. S., Cavallo, D., Gurung, S., Modebadze, T., Woodhall, G., & Henley, J. M. (2019). Changes in excitatory and inhibitory receptor expression and network activity during induction and establishment of epilepsy in the rat Reduced Intensity Status Epilepticus (RISE) model. *Neuropharmacology*, 158, 107728.
38. Numis, A. L., Foster-Barber, A., Deng, X., Rogers, E. E., Barkovich, A. J., Ferriero, D. M., & Glass, H. C. (2019). Early changes in pro-inflammatory cytokine levels in neonates with encephalopathy are associated with remote epilepsy. *Pediatric research*, 86(5), 616-621.
39. Palma, E., Ruffolo, G., Cifelli, P., Roseti, C., Vliet, E. A. V., & Aronica, E. (2017). Modulation of GABAA Receptors in the Treatment of Epilepsy. *Current pharmaceutical design*, 23(37), 5563-5568.
40. Patel, D. C., Tewari, B. P., Chaunsali, L., & Sontheimer, H. (2019). Neuron–glia interactions in the pathophysiology of epilepsy. *Nature Reviews Neuroscience*, 20(5), 282-297.
41. PATEL, M., & WALKER, M. (2022). METABOLISM, REACTIVE OXYGEN SPECIES, AND EPILEPSY. *Neurobiology of the Epilepsies: From Epilepsy: A Comprehensive Textbook*.
42. Puttachary, S., Sharma, S., Stark, S., & Thippeswamy, T. (2015). Seizure-induced oxidative stress in temporal lobe epilepsy. *BioMed research international*, 2015.
43. Qian, F., & Tang, F. R. (2016). Metabotropic glutamate receptors and interacting proteins in epileptogenesis. *Current Neuropharmacology*, 14(5), 551-562.
44. Roma-Mateo, C., Aguado, C., García-Giménez, J. L., Knecht, E., Sanz, P., & Pallardo, F. V. (2015). Oxidative stress, a new hallmark in the pathophysiology of Lafora progressive myoclonus epilepsy. *Free Radical Biology and Medicine*, 88, 30-41.
45. Rossi, J., Cavallieri, F., Biagini, G., Rizzi, R., Russo, M., Cozzi, S., ...& Valzania, F. (2022). Epileptogenesis and Tumorigenesis in Glioblastoma: Which Relationship?. *Medicina*, 58(10), 1349.
46. Russo, M. E. (1981). The pathophysiology of epilepsy. *The Cornell Veterinarian*, 71(2), 221-247.
47. Sandhu, M. R. S., Gruenbaum, B. F., Gruenbaum, S. E., Dhaher, R., Deshpande, K., Funaro, M. C., ...& Eid, T. (2021). Astroglial glutamine synthetase and the pathogenesis of mesial temporal lobe epilepsy. *Frontiers in neurology*, 12, 665334.
48. Sandhu, M. R. S., Gruenbaum, B. F., Gruenbaum, S. E., Dhaher, R., Deshpande, K., Funaro, M. C., ...& Eid, T. (2021). Astroglial glutamine synthetase and the pathogenesis of mesial temporal lobe epilepsy. *Frontiers in neurology*, 12, 665334.
49. Sano, F., Shigetomi, E., Shinozaki, Y., Tsuzukiya, H., Saito, K., Mikoshiba, K., ...& Koizumi, S. (2021). Reactive astrocyte-driven epileptogenesis is induced by microglia initially activated following status epilepticus. *JCI insight*, 6(9).
50. Santos, V. R., Tilelli, C. Q., Fernandes, A., de Castro, O. W., Del-Vecchio, F., & Garcia-Cairasco, N. (2023). Different types of Status Epilepticus may lead to similar hippocampal epileptogenesis processes. *IBRO Neuroscience Reports*.
51. Sarikaya, I. (2015). PET studies in epilepsy. *American journal of nuclear medicine and molecular imaging*, 5(5), 416.
52. Sarlo, G. L., & Holton, K. F. (2021). Brain concentrations of glutamate and GABA in human epilepsy: A review. *Seizure*, 91, 213-227.
53. Sarnat, H. B., & Flores-Sarnat, L. (2021). Excitatory/inhibitory synaptic ratios in polymicrogyria and down syndrome help explain epileptogenesis in malformations. *Pediatric Neurology*, 116, 41-54.
54. Sears, S. M., & Hewett, S. J. (2021). Influence of glutamate and GABA transport on brain excitatory/inhibitory balance. *Experimental Biology and Medicine*, 246(9), 1069-1083.

55. Sgadò, P., Dunleavy, M., Genovesi, S., Provenzano, G., & Bozzi, Y. (2011). The role of GABAergic system in neurodevelopmental disorders: a focus on autism and epilepsy. *International journal of physiology, pathophysiology and pharmacology*, 3(3), 223.
56. Sharma, R., Leung, W. L., Zamani, A., O'Brien, T. J., Casillas Espinosa, P. M., & Semple, B. D. (2019). Neuroinflammation in post-traumatic epilepsy: pathophysiology and tractable therapeutic targets. *Brain sciences*, 9(11), 318.
57. SoltaniKhaboushan, A., Yazdanpanah, N., & Rezaei, N. (2022). Neuroinflammation and proinflammatory cytokines in epileptogenesis. *Molecular Neurobiology*, 59(3), 1724-1743.
58. Song, L. J., Zhang, H., Jin, J. G., Wang, C., Qu, X. P., Jiang, X., ... & Shen, L. L. (2020). Rho-associated Protein Kinase 2 Confers Epileptogenesis through the Activation of Astroglial Stat3 Pathway.
59. Song, L. J., Zhang, H., Qu, X. P., Jin, J. G., Wang, C., Jiang, X., ... & Liu, B. (2022). Increased expression of Rho-associated protein kinase 2 confers astroglial Stat3 pathway activation during epileptogenesis. *Neuroscience Research*, 177, 25-37.
60. Tekgul, H., Serin, H. M., Simsek, E., Kanmaz, S., Gazeteci, H., Azarsiz, E., ... & Gokben, S. (2020). CSF levels of a set of neurotrophic factors (brain-derived neurotrophic factor, nerve growth factor) and neuropeptides (neuropeptide Y, galanin) in epileptic children. *Journal of Clinical Neuroscience*, 76, 41-45.
61. Tekgul, H., Simsek, E., Erdoğan, M. A., Yiğittürk, G., Erbaş, O., & Taşkiran, D. (2020). The potential effects of anticonvulsant drugs on neuropeptides and neurotrophins in pentylenetetrazol kindled seizures in the rat. *International Journal of Neuroscience*, 130(2), 193-203.
62. Terrone, G., Balosso, S., Pauletti, A., Ravizza, T., & Vezzani, A. (2020). Inflammation and reactive oxygen species as disease modifiers in epilepsy. *Neuropharmacology*, 167, 107742.
63. Terrone, G., Frigerio, F., Balosso, S., Ravizza, T., & Vezzani, A. (2019). Inflammation and reactive oxygen species in status epilepticus: Biomarkers and implications for therapy. *Epilepsy & Behavior*, 101, 106275.
64. Verhoog, Q. P., Holtman, L., Aronica, E., & van Vliet, E. A. (2020). Astrocytes as guardians of neuronal excitability: mechanisms underlying epileptogenesis. *Frontiers in Neurology*, 11, 591690.
65. Vezzani, A., Balosso, S., & Ravizza, T. (2008). The role of cytokines in the pathophysiology of epilepsy. *Brain, behavior, and immunity*, 22(6), 797-803.
66. Victor, T. R., & Tsirka, S. E. (2020). Microglial contributions to aberrant neurogenesis and pathophysiology of epilepsy. *Neuroimmunology and neuroinflammation*, 7, 234.
67. Vishwakarma, S., Singh, S., & Singh, T. G. (2022). Pharmacological modulation of cytokines correlating neuroinflammatory cascades in epileptogenesis. *Molecular Biology Reports*, 1-16.
68. Walker, M. C. (2023). Reactive oxygen species in status epilepticus. *Epilepsia Open*, 8, S66-S72.
69. Wang, N., Wang, D., Zhou, H., Xu, C., Hu, X., Qian, Z., & Xu, X. (2021). Serum neuropeptide Y level is associated with post-ischemic stroke epilepsy. *Journal of Stroke and Cerebrovascular Diseases*, 30(2), 105475.
70. Wasterlain, C. G., Fujikawa, D. G., Penix, L., & Sankar, R. (1993). Pathophysiological mechanisms of brain damage from status epilepticus. *Epilepsia*, 34, S37-S53.
71. Wong, R. K., Chuang, S. C., & Bianchi, R. (2002). Metabotropic glutamate receptors and epileptogenesis. *Epilepsy Currents*, 2(3), 81-85.
72. Xu, X. X., Shi, R. X., Fu, Y., Wang, J. L., Tong, X., Zhang, S. Q., ... & Guo, F. (2022). Neuronal nitric oxide synthase/reactive oxygen species pathway is involved in apoptosis and pyroptosis in epilepsy. *Neural Regeneration Research*.
73. Yang, T. T., Qian, F., Liu, L., Peng, X. C., Huang, J. R., Ren, B. X., & Tang, F. R. (2021). Astroglial connexins in epileptogenesis. *Seizure*, 84, 122-128.
74. Yazdanpanah, N., & Rezaei, N. (2022). Neuroinflammation and Proinflammatory Cytokines in Epileptogenesis. *Molecular Neurobiology*.

75. Yeo, X. Y., Cunliffe, G., Ho, R. C., Lee, S. S., & Jung, S. (2022). Potentials of neuropeptides as therapeutic agents for neurological diseases. *Biomedicines*, 10(2), 343.

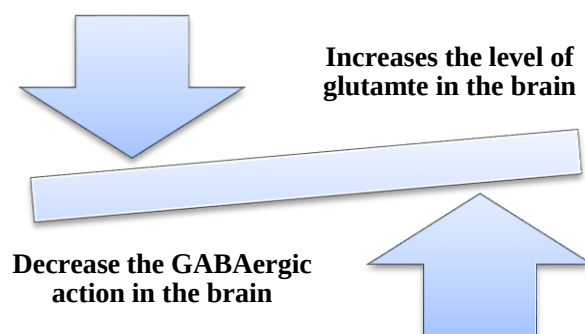


Fig 1 Imbalance of GABA inhibitory and glutamate excitatory NTs in the neurons

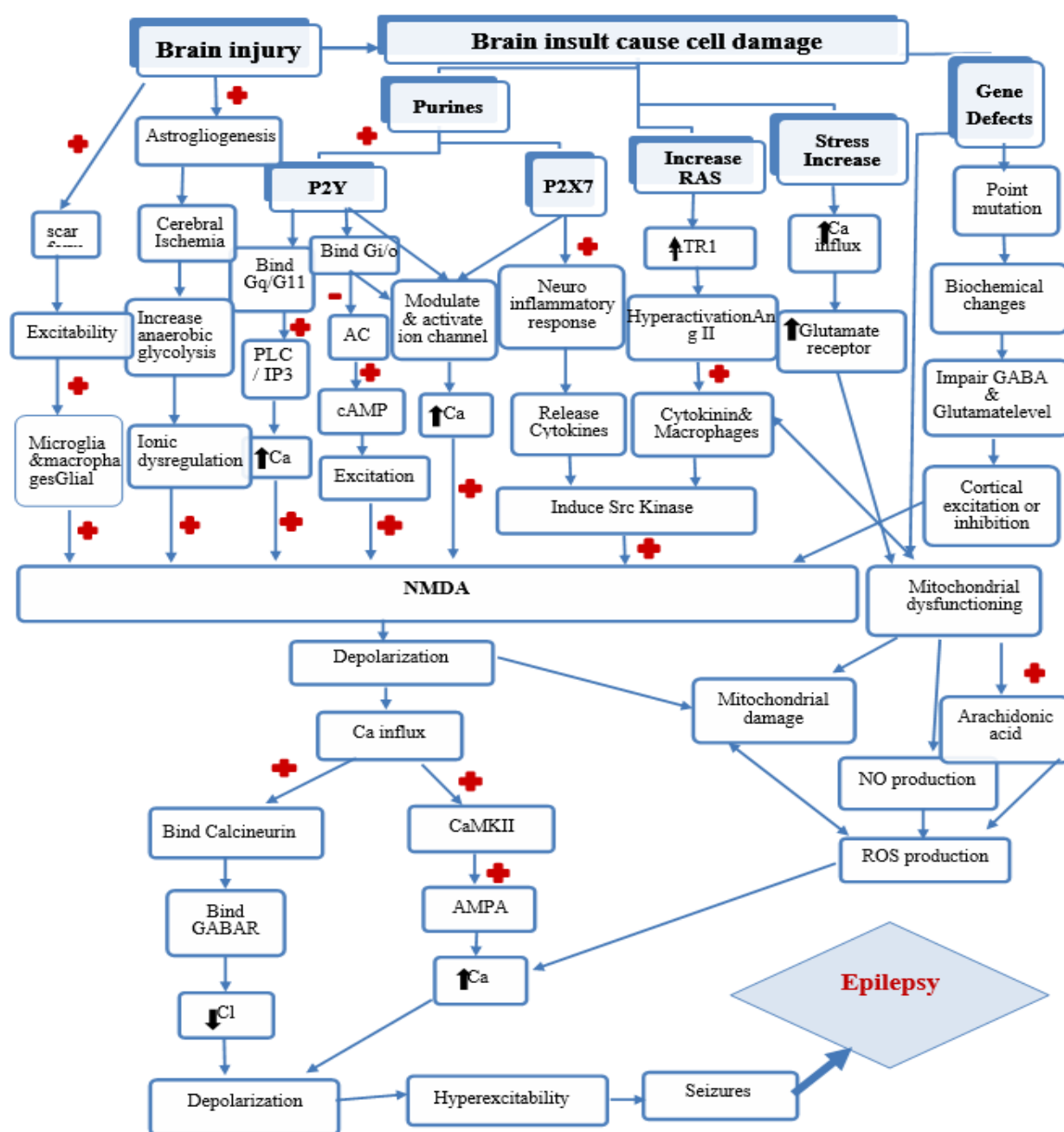


Fig 2 Pathophysiology in epileptogenesis

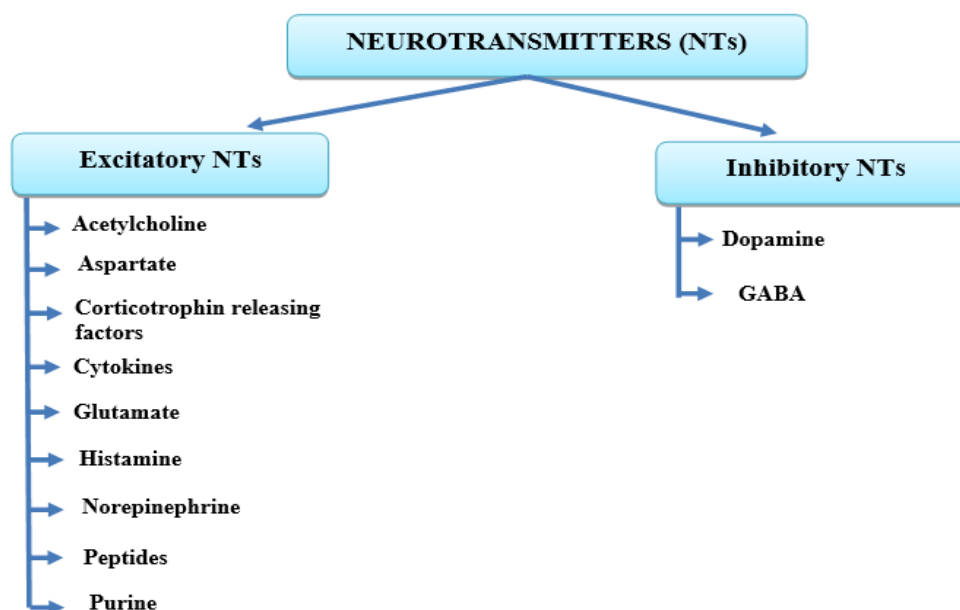


Fig 3 Type's of excitatory and inhibitory NTs involved in epileptogenesis

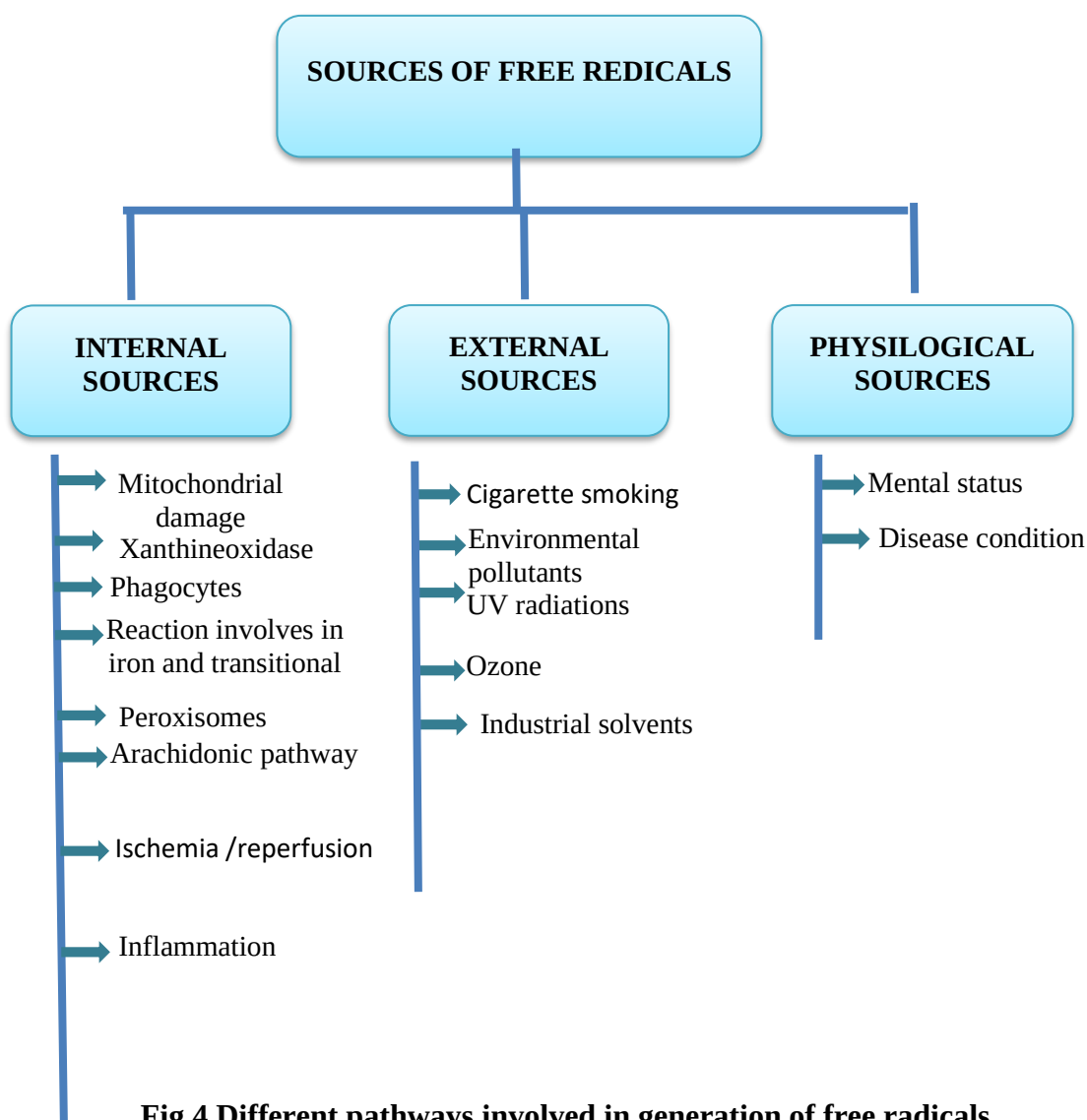


Fig 4 Different pathways involved in generation of free radicals