



Clostridium difficile Infection: An Integrated Review of Microbiology and Internal Medicine

Jyoti Jaitwani¹, Silky Pandey², Hemendra Pratap Srivastava³, Rajnish Sethi⁴

¹Professor, Microbiology, Santosh Medical College, Ghaziabad, Mail id: gagjot@gmail.com

²Professor, Medicine, Saraswati institute of Medical Sciences, Hapur

³Assistant Professor, Medicine, Rama Medical College, Mandhana, Kanpur

⁴Assistant Professor, Orthopedics, Rama Medical College, Mandhana, Kanpur

Abstract

Clostridium difficile infection (CDI) is an important cause of antibiotic-associated diarrhea and healthcare-associated gastrointestinal disease. Between 2009 and 2014, understanding of CDI changed considerably as epidemic strains, molecular diagnostic methods, disruption of the intestinal microbiome, new antimicrobial agents, and microbiota-based therapies became increasingly important. The disease represents a close interface between clinical microbiology and Internal Medicine. Microbiologically, C. difficile is an anaerobic, spore-forming, toxin-producing organism whose pathogenicity is largely mediated by toxins A and B. Antibiotic-induced disruption of intestinal colonization resistance permits expansion of toxigenic organisms, while environmentally resistant spores facilitate transmission and recurrence. Clinically, disease ranges from mild diarrhea to pseudomembranous colitis, toxic megacolon, sepsis, and death. Accurate diagnosis requires correlation of compatible symptoms with laboratory findings because detection of a toxigenic organism alone cannot reliably distinguish active infection from asymptomatic colonization. During the reviewed period, multistep diagnostic algorithms incorporating glutamate dehydrogenase, toxin immunoassays, and nucleic-acid amplification became increasingly important. Treatment was guided by disease severity and recurrence, with metronidazole and oral vancomycin forming the traditional therapeutic foundation and fidaxomicin emerging as an alternative associated with lower recurrence in selected patients. Fecal microbiota transplantation provided compelling evidence that restoration of the intestinal microbial ecosystem can successfully treat recurrent disease. This review integrates the microbiological mechanisms, diagnostic principles, clinical manifestations, medical management, recurrence, and infection-control considerations of CDI using literature published between 2009 and 2014.

Keywords: Clostridium difficile; antibiotic-associated diarrhea; microbiology; internal medicine; toxin; dysbiosis; PCR; vancomycin; fidaxomicin; fecal microbiota transplantation

1. Introduction

Clostridium difficile, currently classified as *Clostridioides difficile*, is an anaerobic, Gram-positive, spore-forming bacillus capable of producing a clinical spectrum ranging from asymptomatic intestinal colonization to fulminant colitis. During the early twenty-first century, CDI became one of the most important healthcare-associated infections, accompanied by changes in incidence, strain distribution, virulence, and clinical severity. The emergence of epidemic lineages such as PCR ribotype 027/BI/NAP1 stimulated renewed investigation into toxin production, sporulation, antimicrobial resistance, transmission, and clinical outcome [1].

The epidemiology of CDI also broadened beyond the traditional hospitalized patient. Cases increasingly occurred in long-term-care facilities and the community, while healthcare outbreaks remained important. Disease burden reflected interactions among antimicrobial exposure, susceptible hosts, environmental contamination, strain characteristics, and transmission within healthcare systems [2]. A European hospital-based survey demonstrated marked differences in incidence and ribotype distribution between institutions and countries, illustrating how microbiological surveillance can influence medical and infection-control practice [3].

The key biological concept is that disease does not result simply from acquiring *C. difficile*. In most patients, antimicrobial or host-related disturbance of the intestinal microbial ecosystem permits a toxigenic strain to proliferate and produce clinically relevant toxin. Thus, the laboratory identifies the organism or its products, while Internal Medicine determines whether those microbiological findings explain the patient's illness [1-3].

2. Epidemiology and Predisposing Factors

Antibiotic exposure is the most important modifiable risk factor for CDI. Different antimicrobial classes disturb the indigenous intestinal flora to different degrees. A meta-analysis of community-associated disease demonstrated particularly strong associations with clindamycin, fluoroquinolones, and cephalosporins [4]. The clinical risk depends not only on antibiotic class but also on duration, combination therapy, repeated exposure, hospitalization, and underlying illness [2,4].

The relationship between antibiotic use and CDI is fundamentally ecological. Broad-spectrum therapy suppresses organisms that normally compete with *C. difficile*, creating an intestinal environment in which spores can germinate and vegetative organisms can expand. Consequently, discontinuation of unnecessary antibiotics is both a clinical intervention and a microbiologically rational strategy for restoring colonization resistance [1,4].

Acid-suppressive therapy was also extensively evaluated during this period. A large meta-analysis demonstrated an association between proton-pump inhibitor use and incident CDI, although heterogeneity among observational studies limited certainty regarding causation [5]. In practice, unnecessary acid suppression should be reconsidered, especially in hospitalized patients receiving antibiotics [5].

Advanced age, severe comorbidity, prolonged hospitalization, immunosuppression, gastrointestinal procedures, and previous CDI further increase susceptibility. Community-associated cases demonstrate that lack of recent hospitalization does not exclude the diagnosis, particularly when recent antimicrobial exposure is present [2,4].

3. The Intestinal Microbiome and Colonization Resistance

One of the major advances in CDI research during 2009-2014 was recognition of the intestinal microbiota as an active defense system. A diverse microbial community competes for nutrients, modifies bile acids, occupies ecological niches, and produces metabolites that restrict expansion of potential pathogens. Antibiotic therapy can disrupt these protective functions even after relatively limited exposure [6].

Experimental work demonstrated that a single dose of clindamycin could cause profound and prolonged changes in the intestinal microbiota and sustained susceptibility to *C. difficile* colitis. The biological effect of antibiotic exposure therefore persists longer than the measurable concentration of the drug itself [6].

Human microbiome studies also demonstrated marked dysbiosis in CDI, including reduced bacterial diversity and depletion of important butyrate-producing organisms. Patients with CDI showed microbial patterns distinct from healthy controls, providing a biological explanation for recurrent disease and for therapies designed to restore rather than further suppress the intestinal ecosystem [7].

This microbiome concept provides a direct bridge between microbiology and medicine: antibiotic exposure alters microbial ecology, ecological disruption permits *C. difficile* expansion, bacterial toxins cause mucosal injury, treatment modifies the microbial environment again, and failure of ecological recovery contributes to recurrence [6,7].

4. Microbiology and Virulence

C. difficile is a strictly anaerobic bacillus capable of producing metabolically dormant spores. Vegetative organisms are relatively vulnerable to oxygen and environmental stress, whereas spores can survive for prolonged periods on healthcare surfaces. Sporulation is therefore central to both transmission and recurrence [1,2].

The major virulence determinants are toxin A (TcdA) and toxin B (TcdB). These large exotoxins disrupt intracellular signaling, alter the actin cytoskeleton, damage the epithelial barrier, and trigger cell death and inflammation. Toxin-mediated injury causes fluid secretion, neutrophilic infiltration, mucosal necrosis, and formation of pseudomembranes. Evidence accumulated during this period that toxin B is particularly important in human disease [8].

Virulence is influenced by additional bacterial factors, including adhesins, surface-layer proteins, motility structures, proteases, toxin regulation, and sporulation characteristics. Strain-specific variation may therefore alter clinical behavior even when the same major toxin genes are present [1,8].

Some strains also produce the binary toxin *C. difficile* transferase (CDT). Binary toxin modifies the actin cytoskeleton through a mechanism distinct from toxins A and B and is associated with important epidemic lineages, including ribotype 027. Its independent contribution to disease severity remained uncertain during the reviewed period, although it became an important molecular epidemiological marker [9].

5. Epidemic Strains

PCR ribotype 027, also referred to as NAP1 or BI, became a major focus of CDI research because of its association with hospital outbreaks, fluoroquinolone resistance, binary toxin production, and severe disease. Its expansion demonstrated how antimicrobial selection pressure can influence the success of particular bacterial lineages [1,2].

However, ribotype alone does not determine individual prognosis. Age, host immune response, renal function, leukocytosis, comorbidity, and disease severity strongly influence outcome. Molecular typing is therefore valuable mainly for surveillance and outbreak investigation rather than for determining treatment of an individual patient [2,3].

The emergence of fluoroquinolone-resistant epidemic strains strengthened the medical rationale for antimicrobial stewardship. Reducing unnecessary exposure to high-risk antibiotics can simultaneously preserve the patient's microbiota and decrease selection pressure favoring resistant *C. difficile* populations [2,4].

6. Clinical Manifestations and Severity

The clinical spectrum ranges from uncomplicated watery diarrhea to severe colitis. Common manifestations include frequent loose stools, abdominal pain, fever, nausea, leukocytosis, rising serum creatinine, and electrolyte disturbances. Severe disease can progress to ileus, toxic megacolon, perforation, sepsis, shock, multiorgan failure, and death [10].

Pseudomembranous colitis represents advanced toxin-mediated mucosal injury. Yellow-white inflammatory plaques composed of fibrin, mucus, inflammatory cells, and necrotic epithelial material overlie damaged colonic mucosa. Endoscopy is not required in most patients but may be useful when stool cannot be obtained because of ileus or when an alternative diagnosis is being considered [10].

Clinical severity must be assessed independently of microbiological positivity. Leukocytosis and renal dysfunction were commonly used during 2009-2014 to identify severe disease, while hypotension, shock, ileus, or megacolon indicated severe complicated or fulminant infection [10].

A patient whose diarrhea decreases because ileus has developed may actually be deteriorating. Serial abdominal examination, hemodynamic assessment, leukocyte count, renal function, and systemic markers of illness are therefore more important than stool frequency alone in severe disease [10].

7. Microbiological Diagnosis

The central diagnostic difficulty is that *C. difficile* can colonize the intestine without producing symptomatic disease. A positive laboratory test therefore does not automatically establish CDI. Testing should be directed toward patients with compatible new-onset unexplained diarrhea rather than asymptomatic individuals or patients with formed stool [10,11].

This distinction between colonization and infection is particularly important as testing becomes more sensitive. A highly sensitive assay may correctly detect a toxigenic organism in a patient whose diarrhea is caused by another condition. Clinical selection of patients for testing is therefore as important as analytical test performance [11].

7.1 Culture and toxigenic culture

Anaerobic stool culture is highly sensitive for detecting *C. difficile* and remains useful for strain characterization, epidemiology, and antimicrobial susceptibility investigations. Conventional culture, however, cannot distinguish toxigenic from nontoxigenic isolates. Toxigenic culture adds demonstration of toxin-producing ability but is too slow for many acute clinical decisions [11].

7.2 Toxin immunoassays and glutamate dehydrogenase

Enzyme immunoassays for toxins A and B provide rapid results and are technically straightforward, but their sensitivity is inadequate for use as the sole exclusion test in patients with a convincing clinical syndrome. A negative toxin EIA therefore does not necessarily rule out CDI [11].

Glutamate dehydrogenase (GDH) is produced by both toxigenic and nontoxigenic strains. Because the assay is sensitive but not specific for toxigenic disease, GDH is useful as an initial screening test that requires confirmation with another method. These limitations encouraged adoption of multistep diagnostic algorithms rather than reliance on a single assay [11].

8. Nucleic-Acid Amplification and Colonization

Nucleic-acid amplification tests, particularly PCR assays targeting toxin genes such as *tcdB*, became increasingly common because of their high analytical sensitivity and rapid turnaround. They detect the genetic potential for toxin production directly from stool [12].

The same sensitivity creates an interpretative problem. PCR positivity demonstrates the presence of a toxigenic organism but does not establish that free toxin is responsible for the patient's symptoms. A prospective multicenter study demonstrated important differences in outcomes according to the diagnostic method used and showed that patients with detectable stool toxin differed clinically from individuals who were culture- or molecular-positive without detectable free toxin [12].

These findings reinforce the principle that laboratory results require clinical interpretation. A patient with compatible diarrhea and detectable toxin represents a different clinical situation from a PCR-positive, toxin-negative patient receiving laxatives or suffering from another gastrointestinal disorder [12].

Asymptomatic carriage further complicates diagnosis. A 2014 study demonstrated that a meaningful proportion of hospitalized patients without diarrhea carried toxigenic *C. difficile* [13]. Treating colonization is generally inappropriate because unnecessary antimicrobial therapy may cause further microbiome disruption. Nonetheless, carriers may still contribute to organism transmission within healthcare environments [13].

9. Initial Medical Management

The first general intervention is to discontinue the precipitating antimicrobial whenever clinically feasible. Continuing unnecessary non-CDI antibiotics prolongs disruption of colonization resistance and may increase treatment failure or recurrence. Fluid replacement, electrolyte correction, nutritional support, and review of unnecessary proton-pump inhibitors are additional components of care [10,14].

During the reviewed period, treatment guidance generally differentiated mild-to-moderate disease from severe and complicated infection. Metronidazole remained commonly used for initial mild-to-moderate CDI, whereas oral vancomycin was favored for severe disease [10,14].

Oral vancomycin achieves high intraluminal concentrations because systemic absorption is minimal. In severe complicated disease with ileus, intravenous metronidazole and rectal vancomycin could be added because oral drug distribution throughout the colon may be unreliable [10,14].

Management of CDI must also account for other active infections. A patient who continues to require broad-spectrum antimicrobial treatment may remain microbiologically vulnerable despite receiving adequate CDI therapy because recovery of the normal intestinal community remains impaired [14].

10. Fidaxomicin

Fidaxomicin was one of the major therapeutic developments of the period. It is a minimally absorbed, relatively narrow-spectrum antimicrobial with potent activity against *C. difficile*. In a randomized controlled trial, fidaxomicin was noninferior to vancomycin for initial clinical cure and was associated with a lower overall recurrence rate, particularly in patients not infected with the epidemic NAP1/BI/027 strain [15].

A second randomized trial conducted in Europe, Canada, and the United States similarly demonstrated noninferiority of fidaxomicin to vancomycin and supported its use as an alternative therapy [16]. These studies shifted attention away from initial resolution of diarrhea alone toward the clinically important outcome of sustained cure without recurrence [15,16].

The microbiological explanation is highly relevant. Fidaxomicin has less activity against many components of the normal intestinal microbiota than broader antimicrobial agents and may therefore permit faster restoration of colonization resistance. Its clinical benefit illustrates how antimicrobial spectrum can influence recurrence through effects on microbial ecology as well as direct killing of the pathogen [15,16].

Susceptibility studies of isolates collected during the fidaxomicin trials confirmed potent in vitro activity against diverse strains of *C. difficile* [17]. Nevertheless, antimicrobial susceptibility alone cannot predict recurrence because spores, host immunity, ongoing antibiotic exposure, and microbiome recovery all contribute to outcome [17].

11. Recurrence

Recurrence is one of the major therapeutic challenges of CDI. After apparently successful treatment, patients may develop renewed symptoms because of relapse with the original strain or reinfection with a different strain. Without molecular typing these mechanisms cannot usually be distinguished in routine clinical practice [1,14].

The probability of further recurrence rises after repeated episodes. Older age, major comorbidity, continuing antimicrobial exposure, persistent dysbiosis, and inadequate host immune responses contribute to susceptibility. Importantly, recurrence does not necessarily imply acquired resistance to the treatment previously used [14].

Traditional approaches included retreatment and tapered or pulsed oral vancomycin schedules. Intermittent administration was intended to eliminate vegetative organisms emerging during repeated cycles of spore germination while allowing gradual recovery of competing intestinal microorganisms [14].

12. Fecal Microbiota Transplantation

Fecal microbiota transplantation (FMT) represented a major conceptual change because it targeted the disturbed intestinal ecosystem rather than simply applying additional antimicrobial pressure [18].

In the landmark randomized trial by van Nood and colleagues, infusion of donor feces following a short course of vancomycin and bowel lavage was substantially more effective for recurrent CDI than conventional vancomycin-based treatment. Most patients receiving donor microbiota achieved resolution, and microbial diversity increased following transplantation [18].

The importance of the study extends beyond treatment efficacy. It provided direct clinical evidence that dysbiosis is central to recurrent CDI. By restoring a complex microbial community, FMT re-establishes

ecological competition and colonization resistance rather than targeting one specific toxin or bacterial enzyme [18].

During 2009-2014, questions remained regarding donor screening, optimal preparation, route of administration, long-term safety, and standardization. Nevertheless, FMT became an increasingly important option for patients with multiple recurrences who had failed appropriate antimicrobial treatment [14,18].

13. Fulminant Disease and Surgery

Patients with hypotension, shock, ileus, toxic megacolon, increasing lactate, rapidly rising leukocyte count, or progressive organ dysfunction require intensive monitoring and early surgical assessment. Severe disease may actually present with reduced stool output when ileus prevents passage of diarrhea [10,19].

Imaging may demonstrate bowel-wall thickening, marked colonic dilation, pericolonic inflammation, ascites, or perforation, but radiological findings cannot establish the microbiological etiology of colitis. Imaging is most valuable when complications such as toxic megacolon or perforation are suspected [19].

Subtotal colectomy with preservation of the rectum was the conventional operative approach to fulminant disease during this period, although alternative strategies were being explored. Delay until irreversible multiorgan failure significantly reduces the likelihood of surgical rescue, emphasizing the need for early collaboration between internists, critical-care teams, infectious-disease specialists, and surgeons [19].

14. Infection Prevention and Antimicrobial Stewardship

Environmental persistence of spores makes CDI particularly difficult to control. Contact precautions, gloves, gowns, environmental cleaning with sporicidal agents, and careful hand hygiene are critical components of infection prevention. Alcohol-based hand sanitizers do not reliably eliminate spores, so handwashing with soap and water remains important in appropriate clinical settings [10,20].

Microbiology laboratories contribute directly to infection prevention by rapidly identifying positive patients and supporting strain typing during suspected outbreaks. At the same time, inappropriate testing may inflate reported infection rates by identifying colonized individuals rather than true cases [11,12].

Antimicrobial stewardship is among the most important population-level preventive measures. Restricting unnecessary high-risk antibiotics reduces disruption of host microbiota and decreases selection pressure favoring resistant epidemic strains [2,4]. Thus, stewardship operates at two microbiological levels: it preserves the patient's colonization resistance and influences the composition of the circulating pathogen population [2,4].

15. Integrated Microbiology-Medicine Approach

The central principle is that microbiological detection and clinical diagnosis are not synonymous. The laboratory establishes whether *C. difficile*, toxin, or toxin genes are present; Internal Medicine determines whether those findings explain the patient's illness [11,12].

Conversely, clinical suspicion without appropriate microbiological evaluation may lead to unnecessary treatment of patients whose diarrhea has another cause. Repeated empiric antimicrobial exposure may

worsen dysbiosis, demonstrating why microbiology and medicine must function as an integrated diagnostic system [20].

Table 1. Integrated microbiology-medicine interpretation of common CDI scenarios.

Clinical issue	Microbiological interpretation	Medical implication
Antibiotic-associated diarrhea	Disrupted colonization resistance permits toxigenic <i>C. difficile</i> expansion	Review and discontinue unnecessary antibiotics
Positive GDH, negative toxin	Organism antigen detected but active toxin not demonstrated	Additional testing and clinical correlation required
PCR positive, toxin negative	Toxigenic strain present; active toxin-mediated disease uncertain	Avoid diagnosing CDI from molecular positivity alone
Toxin detected with compatible diarrhea	Strong evidence of active disease	Initiate severity-appropriate treatment
Recurrent CDI	Spores, persistent dysbiosis, relapse, or reinfection may contribute	Use recurrence-specific treatment strategies
Multiple recurrences	Failure of microbial ecosystem recovery becomes increasingly important	Consider FMT
Fulminant colitis	Severe toxin-mediated inflammatory injury and organ dysfunction	Intensive care and early surgical involvement
Epidemic strain detected	Important epidemiological information	Strengthen surveillance and infection-control measures

16. Practical Diagnostic and Therapeutic Algorithm

Step 1: Establish clinical compatibility. Test patients with otherwise unexplained new-onset diarrhea. Avoid routine testing of asymptomatic individuals or formed stool [10,11].

Step 2: Review predisposing medications. Identify recent antibiotics, continuing broad-spectrum treatment, and unnecessary acid suppression [4,5].

Step 3: Use an appropriate diagnostic algorithm. Understand whether the laboratory reports GDH, toxin, NAAT, or a combination of these tests [11,12].

Step 4: Interpret results clinically. A positive molecular test in a low-probability patient may represent colonization rather than disease [12,13].

Step 5: Assess severity. Evaluate leukocyte count, renal function, hemodynamic status, abdominal findings, ileus, and organ dysfunction [10,14].

Step 6: Initiate appropriate therapy. Metronidazole, oral vancomycin, and increasingly fidaxomicin were used during this period according to disease severity, recurrence history, and patient factors [14-16].

Step 7: Monitor for complications. Ileus, hypotension, increasing lactate, abdominal distension, and multiorgan dysfunction should prompt critical-care and surgical assessment [19].

Step 8: Address recurrence differently from primary disease. Review ongoing antimicrobial exposure and consider vancomycin taper/pulse regimens, fidaxomicin, or other recurrence-directed approaches [14-16].

Step 9: Restore the microbial ecosystem when appropriate. FMT should be considered in appropriately selected patients with repeated recurrent disease [18].

Step 10: Prevent future disease. Apply contact precautions, environmental cleaning, and antimicrobial stewardship [10,20].

17. Conclusion

Between 2009 and 2014, CDI evolved conceptually from a relatively straightforward form of antibiotic-associated diarrhea into a complex disease involving microbial ecology, toxin-mediated injury, environmental transmission, and host susceptibility. The ability of *C. difficile* to produce environmentally resistant spores permits persistence and transmission, while toxins A and B damage the intestinal epithelium and trigger inflammatory colitis [1,8].

Clinical microbiology is essential to diagnosis, but no laboratory method available during this period perfectly distinguished active toxin-mediated disease from asymptomatic colonization. Culture, GDH, toxin immunoassays, and nucleic-acid amplification tests measure different biological aspects of CDI and must be interpreted according to the patient's clinical presentation [11-13].

Internal Medicine transforms these findings into management decisions. Clinicians must identify modifiable risk factors, assess disease severity, correct fluid and electrolyte abnormalities, select appropriate antimicrobial therapy, recognize recurrence, and escalate rapidly when fulminant colitis develops [10,14,19].

The therapeutic developments of the period were particularly important. Fidaxomicin demonstrated efficacy comparable to vancomycin while reducing recurrence in selected patient populations, showing how preservation of microbial ecology can influence clinical outcomes [15,16]. FMT went further by directly restoring microbial diversity and provided compelling evidence that recurrent CDI is fundamentally a disease of disrupted colonization resistance as well as pathogen overgrowth [18].

The most effective clinical model is therefore an integrated one: antibiotic exposure leads to microbiome disruption, toxigenic *C. difficile* expansion and toxin-mediated colitis; clinically appropriate laboratory diagnosis then guides severity-based treatment, restoration of microbial ecology, and prevention through infection control and antimicrobial stewardship. CDI remains an important example of modern infectious-disease medicine in which understanding the pathogen, the microbial

ecosystem, and the patient is equally necessary for accurate diagnosis and successful treatment [1,10,18,20].

References

1. Rupnik M, Wilcox MH, Gerding DN. Clostridium difficile infection: new developments in epidemiology and pathogenesis. *Nat Rev Microbiol.* 2009;7(7):526-536. doi:10.1038/nrmicro2164. PMID: 19528959.
2. Freeman J, Bauer MP, Baines SD, Corver J, Fawley WN, Goorhuis B, et al. The changing epidemiology of Clostridium difficile infections. *Clin Microbiol Rev.* 2010;23(3):529-549. doi:10.1128/CMR.00082-09. PMID: 20610822.
3. Bauer MP, Notermans DW, van Benthem BHB, Brazier JS, Wilcox MH, Rupnik M, et al. Clostridium difficile infection in Europe: a hospital-based survey. *Lancet.* 2011;377(9759):63-73. doi:10.1016/S0140-6736(10)61266-4. PMID: 21084111.
4. Deshpande A, Pasupuleti V, Thota P, Pant C, Rolston DDK, Sferri TJ, et al. Community-associated Clostridium difficile infection and antibiotics: a meta-analysis. *J Antimicrob Chemother.* 2013;68(9):1951-1961. doi:10.1093/jac/dkt129. PMID: 23620467.
5. Kwok CS, Arthur AK, Anibueze CI, Singh S, Cavallazzi R, Loke YK. Risk of Clostridium difficile infection with acid suppressing drugs and antibiotics: meta-analysis. *Am J Gastroenterol.* 2012;107(7):1011-1019. doi:10.1038/ajg.2012.108. PMID: 22525304.
6. Buffie CG, Jarchum I, Equinda M, Lipuma L, Gorboun A, Viale A, et al. Profound alterations of intestinal microbiota following a single dose of clindamycin results in sustained susceptibility to Clostridium difficile-induced colitis. *Infect Immun.* 2012;80(1):62-73. doi:10.1128/IAI.05496-11. PMID: 22006564.
7. Antharam VC, Li EC, Ishmael A, Sharma A, Mai V, Rand KH, Wang GP. Intestinal dysbiosis and depletion of butyrogenic bacteria in Clostridium difficile infection and nosocomial diarrhea. *J Clin Microbiol.* 2013;51(9):2884-2892. doi:10.1128/JCM.00845-13. PMID: 23804381.
8. Denève C, Janoir C, Poilane I, Fantinato C, Collignon A. New trends in Clostridium difficile virulence and pathogenesis. *Int J Antimicrob Agents.* 2009;33 Suppl 1:S24-S28. doi:10.1016/S0924-8579(09)70012-3. PMID: 19303565.
9. Gerding DN, Johnson S, Rupnik M, Aktories K. Clostridium difficile binary toxin CDT: mechanism, epidemiology, and potential clinical importance. *Gut Microbes.* 2014;5(1):15-27. doi:10.4161/gmic.26854. PMID: 24253566.
10. Cohen SH, Gerding DN, Johnson S, Kelly CP, Loo VG, McDonald LC, et al. Clinical practice guidelines for Clostridium difficile infection in adults: 2010 update by the Society for Healthcare Epidemiology of America and the Infectious Diseases Society of America. *Infect Control Hosp Epidemiol.* 2010;31(5):431-455. doi:10.1086/651706. PMID: 20307191.
11. Crobach MJT, Dekkers OM, Wilcox MH, Kuijper EJ. European Society of Clinical Microbiology and Infectious Diseases: data review and recommendations for diagnosing Clostridium difficile infection. *Clin Microbiol Infect.* 2009;15(12):1053-1066. doi:10.1111/j.1469-0691.2009.03098.x. PMID: 19929972.
12. Planche TD, Davies KA, Coen PG, Finney JM, Monahan IM, Morris KA, et al. Differences in outcome according to Clostridium difficile testing method: a prospective multicentre

- diagnostic validation study. *Lancet Infect Dis.* 2013;13(11):936-945. doi:10.1016/S1473-3099(13)70200-7. PMID: 24007915.
13. Alasmari F, Seiler SM, Hink T, Burnham CAD, Dubberke ER. Prevalence and risk factors for asymptomatic *Clostridium difficile* carriage. *Clin Infect Dis.* 2014;59(2):216-222. doi:10.1093/cid/ciu258. PMID: 24755858.
 14. Debast SB, Bauer MP, Kuijper EJ. European Society of Clinical Microbiology and Infectious Diseases: update of the treatment guidance document for *Clostridium difficile* infection. *Clin Microbiol Infect.* 2014;20 Suppl 2:1-26. doi:10.1111/1469-0691.12418. PMID: 24118601.
 15. Louie TJ, Miller MA, Mullane KM, Weiss K, Lentnek A, Golan Y, et al. Fidaxomicin versus vancomycin for *Clostridium difficile* infection. *N Engl J Med.* 2011;364(5):422-431. doi:10.1056/NEJMoa0910812. PMID: 21288078.
 16. Cornely OA, Crook DW, Esposito R, Poirier A, Somero MS, Weiss K, et al. Fidaxomicin versus vancomycin for infection with *Clostridium difficile* in Europe, Canada, and the USA: a double-blind, non-inferiority, randomised controlled trial. *Lancet Infect Dis.* 2012;12(4):281-289. doi:10.1016/S1473-3099(11)70374-7. PMID: 22321770.
 17. Goldstein EJC, Citron DM, Sears P, Babakhani F, Sambol SP, Gerding DN. Comparative susceptibilities to fidaxomicin of isolates collected at baseline, recurrence, and failure from patients in two phase III trials. *Antimicrob Agents Chemother.* 2011;55(11):5194-5199. doi:10.1128/AAC.00625-11. PMID: 21844318.
 18. van Nood E, Vrieze A, Nieuwdorp M, Fuentes S, Zoetendal EG, de Vos WM, et al. Duodenal infusion of donor feces for recurrent *Clostridium difficile*. *N Engl J Med.* 2013;368(5):407-415. doi:10.1056/NEJMoa1205037. PMID: 23323867.
 19. To KB, Napolitano LM. *Clostridium difficile* infection: update on diagnosis, epidemiology, and treatment strategies. *Surg Infect (Larchmt).* 2014;15(5):490-502. doi:10.1089/sur.2013.186. PMID: 25314344.
 20. Wilcox MH. Overcoming barriers to effective recognition and diagnosis of *Clostridium difficile* infection. *Clin Microbiol Infect.* 2012;18 Suppl 6:13-20. doi:10.1111/1469-0691.12057. PMID: 23121550.