RESEARCH ARTICLE DOI: 10.53555/s51mpj44

# PREVALENCE AND ANTIFUNGAL SUSCEPTIBILITY OF CANDIDA SPECIES IN BLOODSTREAM INFECTIONS AMONG PEDIATRIC PATIENTS IN A TERTIARY CAREHOSPITAL

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#### **Abstract**

**Introduction:** Candidemia, a severe and potentially life-threatening bloodstream infection caused by *Candida* species, remains a significant concern in pediatric populations, especially in developing countries such as India. The increasing prevalence of non-*albicans Candida* (NAC) species, which often exhibit reduced susceptibility to commonly used antifungal agents, has further complicated clinical management and therapeutic outcomes.

**Aim:** The present study aimed to evaluate the prevalence of *Candida* species in bloodstream infections among pediatric patients and to characterize their antifungal resistance patterns.

**Material and methods:** Blood samples from patients with suspected candidemia were collected in automated blood culture bottles. Identification of *Candida* species was performed using standard microbiological techniques, including the VITEK 2 Compact system. Antifungal susceptibility was assessed using the AST-YS09 card, which determines the minimum inhibitory concentration (MIC) for six antifungal agents i.e., voriconazole, caspofungin, flucytosine, micafungin, fluconazole, and amphotericin B.

**Results:** A total of 97 (6.2%) *Candida* isolates were identified, with *Candida albicans* (32%) being the most prevalent species, followed by *Candida tropicalis* (22.6%), *Candida parapsilosis* (19.5%), *Candida pelliculosa* (12.3%), *Candida glabrata* (7.2%), *Candida krusei* (4.1%), and *Candida guilliermondii* (2%). Antifungal resistance was notable, with 36% of isolates resistant to flucytosine, 32% to fluconazole, 26% to amphotericin B, 21.6% to voriconazole, 14.4% to micafungin and 12.3% to caspofungin.

**Conclusion:** Our study highlights a higher incidence of non-albicans Candida species compared to Candida albicans, with a notable resistance to various antifungal agents. These findings underscore the urgent need for antifungal stewardship programs and stringent preventive control measures to mitigate the further spread of antifungal resistance.

**Keywords:** Candida albicans; Non-albicans Candida; Candidemia; Antifungals; Candidemia; Resistance; VITEK

#### **Introduction:**

Candidemia is a leading cause of invasive fungal infections in hospitalized patients [1,2]. *Candida* species have now emerged as the fourth most common causative agent of nosocomial sepsis [3]. Pediatric patients diagnosed with candidemia experience higher mortality rates, prolonged hospital stays, and a significant increase in total hospital costs per patient [4].

According to data from the Centers for Disease Control and Prevention (CDC) and the National Healthcare Safety Network (NHSN), *Candida* species rank as the fourth most common pathogens causing bloodstream infections (BSIs) and the fifth most prevalent hospital-acquired pathogens [5–7]. While *Candida albicans* was historically the predominant cause of candidemia, non-albicans Candida (NAC) species, such as *C. tropicalis* and *C. parapsilosis* in developing countries and *C. glabrata* in developed nations, are increasingly recognized as significant contributors to nosocomial infections [8,9]

Pediatric patients with Candida bloodstream infections (BSIs) remain at risk for other nosocomial infections, including recurrent candidemia or secondary bacterial infections [10,11]. The rising antifungal resistance among Candida species significantly contributes to morbidity and mortality in neonates and children. Fluconazole remains the most commonly used antifungal for treating candidemia; however, Candida krusei exhibits intrinsic resistance to fluconazole, while other nonalbicans Candida (NAC) species, such as Candida guilliermondii, Candida glabrata, and Candida inconspicua, demonstrate reduced susceptibility to azoles. Although echinocandin resistance has traditionally been considered rare, its incidence is increasing. Both intrinsic and acquired resistance to antifungal agents pose significant challenges in the management of candidemia [12]. Therefore, accurate identification of Candida species and determination of their antifungal susceptibility patterns are crucial for selecting the most effective treatment and ensuring appropriate antifungal coverage. Unfortunately, data on antifungal susceptibility patterns in pediatric patients with candidemia remain limited. As a result, empirical treatment for suspected invasive candidiasis in children is often based on extrapolated data from adult populations. Given this gap in pediatric-specific data, the present study aims to evaluate the prevalence of *Candida* species in bloodstream infections among pediatric patients and to characterize their antifungal resistance patterns.

### Methodology

This retrospective study was conducted over a period of two years in the Department of Microbiology at the Government Institute of Medical Sciences (GIMS), Greater Noida, India.

Blood cultures received from all suspected cases of septicemia and various locations, such as neonatal intensive care units (NICU), Pediatric intensive care units (PICU), Sick intensive care unit (SNCU), outpatient departments (OPDs), and wards during this period were processed using the Becton Dickinson BACTEC TM FX40 automated culture systems. Blood samples were collected under strict aseptic conditions in BacT/ALERT (bioMérieux) culture bottles for aerobic bacterial and fungal growth and incubated at 37°C for up to seven days. Upon a positive signal from the BacT/ALERT system, the culture bottles were unloaded, and a preliminary Gram stain was performed. The samples were then subcultured on Sabouraud dextrose agar (SDA) supplemented with chloramphenicol (0.5 g/L) (Oxoid, UK) and incubated at 37°C for 24–48 hours.

Colonies were identified using standard microbiological methods, including colony morphology, Gram staining, and germ tube testing. Further confirmation was performed using the VITEK 2 Compact system (bioMérieux, USA) with YST ID REF21343 (yeast identification cards), following the manufacturer's instructions.

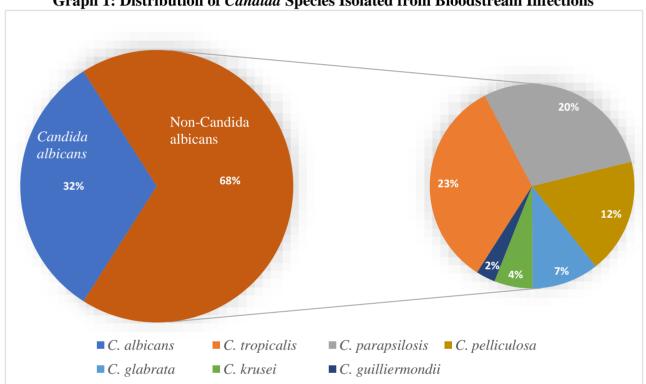
## **Antifungal Susceptibility Testing**

Antifungal susceptibility testing of *Candida* isolates was conducted using the VITEK 2 system, following Clinical and Laboratory Standards Institute (CLSI) guidelines [13]. The AST-YS08 card was used to determine the minimum inhibitory concentration (MIC) of six antifungal agents: voriconazole, caspofungin, flucytosine, micafungin, fluconazole, and amphotericin B.

Data obtained from this study were analyzed using various descriptive statistics like percentage and proportion. The Chi-square test was used to determine the association of various parameters with *C. albicans* and NAC. All the data analysis was performed using the SPSS software (version 28.0).

#### **Results**

Among all 1542 the blood samples processed, a total of 97 (6.2%) samples tested positive for *Candida* species. Of these, *Candida albicans* accounted for 31 (32%) isolates, while the majority, 66 (68%), were non-albicans Candida (NAC) species (Graph 1). Among NAC species, *Candida tropicalis* was the most prevalent, with 22 (22.6%) isolates, followed by *Candida parapsilosis* 19 (19.5%), *Candida pelliculosa* 12 (12.3%), *Candida glabrata* 7 (7.2%), *Candida krusei* 4 (4.1%) and *Candida guilliermondii* 2 (2.1%) (Graph 2).

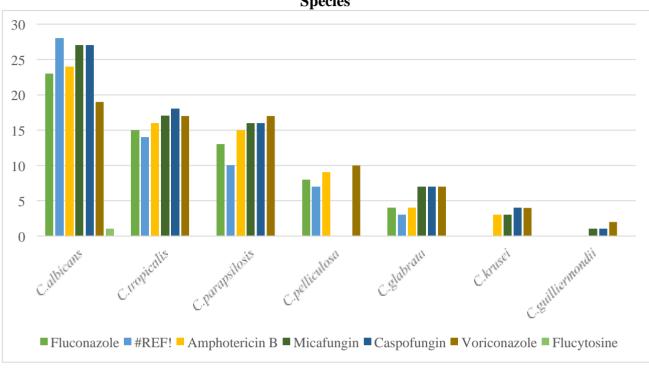


Graph 1: Distribution of Candida Species Isolated from Bloodstream Infections

Among these, 97 samples yielded *Candida* species. The highest incidence of candidemia was observed in infants (41.2%), followed by neonates (37.1) (Table 1).

Table 1: Distribution of Candidemia Cases by *Candida* Species and Age Group

Species	Neonate (birth to <1 month)	Infant (1 month to <1 years)	Children (1 to <12 years	Adolescent (12–14 years)	Total No. (%)
C. albicans	13 (36%)	11 (27%)	5 (33%)	2 (33%)	31 (28.6%)
C. tropicalis	8 (22%)	9 (22%)	3 (20%)	2 (33%)	22 (23.5%)
C. parapsilosis	7 (19%)	9 (22%)	2 (13%)	1 (17%)	19 (17.6%)
C. pelliculosa	4 (11%)	7 (17%)	1 (7%)	0	12 (12.5%)
C. glabrata	2 (5%)	2 (5%)	2 (13%)	1 (17%)	7 (8.8%)
C. krusei	2 (5%)	1 (2.5%)	1 (7%)	0	4 (6.6%)
C. guilliermondii	0	1 (2.5%)	1 (7%)	0	2 (2.2%)
Total	36 (37.1%)	40 (41.2%)	15 (15.4%)	6 (6.5%)	97



**Graph 2: Antifungal Susceptibility Patterns of** *Candida albicans* **and Non-***Candida albicans* **Species** 

Antifungal susceptibility testing (AFST) results revealed *C.pelliculosa* read fluconazole, voriconazole, amphotericin B and flucytosine. *C.krusei* is intrinsic resistant to fluconazole and read caspofungin, flucytosine, micafungin, fluconazole and amphotericin B.

Among *Candida* isolates voriconazole (78.3%) and caspofungin (75.2%) exhibited the highest efficacy against *Candida* species, followed by micafungin (73.1%) and amphotericin B (73%). Fluconazole and flucytosine demonstrated the lowest efficacy, with a susceptibility rate of 65% and 64%.

Among individual species, *Candida albicans* showed the highest resistance to voriconazole (38.7%) and fluconazole (29%). *Candida tropicalis* exhibited maximum resistance to flucytosine (37%) and fluconazole (32%), while *Candida parapsilosis* displayed the highest resistance to flucytosine (47%) and fluconazole (31%). *Candida glabrata* showed notable resistance to flucytosine (57%). *Candida krusei* was intrinsically resistant to fluconazole, whereas *Candida guilliermondii* demonstrated the highest resistance to amphotericin B (100%) and flucytosine (100%).

#### Discussion

The overall prevalence of *Candida* species in our study is 6.2% distribution of *Candida* species in this study revealed that *Candida albicans* accounted for 32%% of cases, while non-*albicans Candida* (NAC) species constituted 68%. This finding is consistent with a study conducted in Haryana, India, which reported a higher prevalence of NAC species (77%) compared to *C. albicans* (33%) [14]. Similar trends have been observed in recent studies from various Indian healthcare settings, where NAC isolation rates have ranged from 66% to 90% [15–18].

In recent years, *Candida* has emerged as a major causative agent of bloodstream infections. In the United States, *Candida* species rank as the fourth most common cause of bloodstream infections [19] and are among the top ten pathogens responsible for hospital-acquired bloodstream infections [12]. Additionally, a study by Chakrabarti A. reported that the incidence of candidemia in developing countries is 4–15 times higher than in developed nations [20].

In this study, *Candida albicans* accounted for 32% of cases, while non-*albicans Candida* (NAC) species constituted 68%. This finding aligns with a study conducted in Haryana, India, which reported a higher prevalence of NAC species (77%) compared to *C. albicans* (33%) [14]. Similar trends have

been observed across various Indian healthcare settings, with NAC isolation rates ranging from 66% to 90% [15–18]. Over the years, *Candida* has emerged as a major cause of bloodstream infections. In the United States, *Candida* species are the fourth most common cause of bloodstream infections [19] and rank among the top ten pathogens responsible for hospital-acquired bloodstream infections [12]. Furthermore, a study by Chakrabarti A. reported that the incidence of candidemia in developing countries is 4–15 times higher than in developed nations. [20]

In the present study, among 97 Candida isolates, the most common species identified was C. albicans (32%), followed by C. tropicalis (23%), C. parapsilosis (19.5%), C. pelliculosa (12.3%), C. glabrata (7.2%), C. krusei (4.1%), and C. guilliermondii (2%). These findings are consistent with studies conducted by Capoor Malini et al. [21] and Naseem Shaikh et al. [31]. Additionally, C. tropicalis was reported as the most common species by Jyoti Pal et al. [32]. Similarly, a study from North India identified C. tropicalis (39%) as the most frequently isolated species, followed by C. parapsilosis (18%) [33]. In contrast, a study from South India found C. parapsilosis to be the predominant pathogen among pediatric patients [34].

The pediatric population is particularly vulnerable to *Candida* bloodstream infections (BSI). In the present study, the prevalence of candidemia was highest among infants (41.2%), followed by neonates (3%). A declining trend in infection rates was observed with increasing age, suggesting an inverse relationship between age and candidemia prevalence. This pattern aligns with findings from other studies, which reported a high prevalence of candidemia among neonates (35%) [35–36]. The increased susceptibility in neonates and young children is primarily attributed to their underdeveloped immune system, which is not yet fully equipped to mount an effective defense against infections. Consequently, *Candida* can proliferate and invade the bloodstream, leading to BSI. Significant variations in *Candida* species distribution were observed across different pediatric age groups. In our study, *C. albicans* (36%) and *C. tropicalis* (22%) were the predominant species among neonates. Similarly, a study from Chile reported that neonates had the highest proportion of *C. albicans* cases (60%) [37]. Among infants, *C. albicans* (27%) was the most frequently isolated species, followed by *C. tropicalis* (22%).

Regarding antifungal resistance, the highest resistance was observed against flucytosine (36%), followed by fluconazole (32%) and amphotericin B (26%). Previous studies have also reported fluconazole resistance rates of 10% in Ethiopia (Seyoum E et al., 2020), 15% in China (Bilal H et al., 2022), 5% in Italy (Bedini A et al., 2006), and 34.8% in India (Bhattacharjee P et al., 2016) [38, 24–26]. Many *Candida* species exhibit intrinsic resistance to antifungal agents, with *C. krusei* being inherently resistant to fluconazole. As fluconazole remains one of the most commonly used antifungal agents, its resistance is rising in various parts of the world, posing a significant challenge in treatment [39].

Most *Candida* isolates in our study were susceptible to caspofungin based on the revised CLSI breakpoints. However, resistance was observed against fluconazole, amphotericin B, micafungin, voriconazole, and caspofungin. This resistance may be attributed to reduced susceptibility to fluconazole and cross-resistance among azole antifungals. Consistent with previous studies, we found higher mortality rates among patients infected with non-*albicans Candida* (NAC) species compared to those infected with *C. albicans* (68% vs. 32 respectively), highlighting the clinical significance of NAC infections and the need for effective antifungal stewardship.

# **CONCLUSION**

In the present study, candidemia accounted for 6.2% of all bloodstream infections (BSIs) among pediatric patients, with a predominance of non-albicans Candida (NAC) species. The most commonly isolated species were Candida tropicalis and Candida parapsilosis. Among the antifungal agents tested, voriconazole exhibited the highest activity, whereas fluconazole showed the lowest efficacy. Early diagnosis of Candida BSI and determination of its resistance profile are crucial for optimizing treatment strategies and improving clinical outcomes, particularly in critically ill neonates and young children.

#### **Conflicts of Interest**

The authors declare no conflicts of interests.

#### References

- 1. Beck-Sagué C, Jarvis WR. Secular trends in the epidemiology of nosocomial fungal infections in the United States, 1980-1990. National nosocomial infections surveillance system. J Infect Dis 1993:167:1247-51.
- 2. Edmond MB, Wallace SE, McClish DK, Pfaller MA, Jones RN, Wenzel RP, et al. Nosocomial bloodstream infections in United States hospitals: A three year analysis. Clin Infect Dis 1999;29:239-44.
- 3. Pfaller M. A., Diekema D. J.2007. Epidemiology of invasive candidiasis: a persistent public health problem. Clin. Microbiol. Rev. 20:133163.
- 4. Zaoutis TE, Argon J, Chu J, Berlin JA, Walsh TJ, Feudtner C, et al. The epidemiology and attributable outcomes of candidemia in adults and children hospitalized in the United States: A propensity analysis. Clin Infect Dis 2005;41:1232-9.
- 5. Pfaller MA, Diekema D. Epidemiology of invasive candidiasis: a persistent public health problem. Clin microbiol rev. 2007;20(1):133-163.
- 6. Sievert DM, Ricks P, Edwards JR, Schneider A, Patel J, Srinivasan A, Kallen A, Limbago B, Fridkin S, National Healthcare Safety Network (NHSN) Team and Participating NHSN Facilities. Antimicrobial- resistant pathogens associated with healthcare- associated infections: summary of data reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention 2009–2010. Infect Control Hosp Epidemiol. 2013;34(1):1–14.
- 7. Wisplinghoff H, Bischoff T, Tallent SM, Seifert H, Wenzel RP, Edmond MB. Nosocomial bloodstream infections in US hospitals: analysis of 24,179 cases from a prospective nationwide surveillance study. Clin infect dis. 2004;39(3):309-317.
- 8. Kaur H, Singh S, Rudramurthy SM, Ghosh AK, Jayashree M, Narayana Y, et al. Candidaemia in a tertiary care centre of developing country: Monitoring possible change in spectrum of agents and antifungal susceptibility. Indian J Med Microbiol. 2020;38(1):109 16.
- 9. Shivaprakasha S, Radhakrishnan K, Karim PM. Candida spp. other than Candida albicans: a major cause of fungaemia in a tertiary care centre. Indian J Med Microbiol. 2007;25(4):405–7.
- 10. Lai, M.Y.; Hsu, J.F.; Chu, S.M.; Wu, I.H.; Huang, H.R.; Chiang, M.C.; Fu, R.-H.; Tsai, M.-H. Risk factors and outcomes of recurrent candidemia in children: Relapsed or re-infection? J. Clin. Med. 2019, 8, E99.
- 11. Lai, M.Y.; Hsu, J.F.; Chu, S.M.; Wu, I.H.; Huang, H.R.; Lin, C.C.; Lee, I.-T.; Chiang, M.-C.; Fu, R.-H.; Tsai, M.-H. Breakthrough candidemia in children: Clinical and microbiological characteristics, therapeutic strategies and impact on outcomes. Future Microbiol. 2017, 12, 695–705.
- 12. Sanguinetti M, Posteraro B. Lass-Flörl C. Antifungal drug resistance among Candida species: mechanisms and clinical impact. Mycoses. 2015;58(2):2–13
- 13. Clinical and Laboratory Standard Institute. CLSI (2017). M60. Performance Standards for Antifungal Susceptibility Testing of Yeasts, 1st Edn. Wayne, PA: Clinical and Laboratory Standards Institute.
- 14. Sandhu R, Dahiya S, Sayal P, et al. Increased role of nonalbicans Candida, potential risk factors and attributable mortality in hospitalized patients. J Health Res Rev 2017;4(3):78-83.
- 15. Chakrabarti A, Mohan B, Shrivastava SK, Marak RS, Ghosh A, Ray P, et al. Change in distribution and amp; antifungal susceptibility of Candida species isolated from candidaemia cases in a tertiary care centre during 1996-2000. Indian J Med Res 2002;116:5-12. 21.
- 16. Chakrabarti A, Singh K, Das S. Changing face of candidemia. Ind J Med Microbiol 1999;17:160-6. 22.
- 17. Singhi SC, Reddy TC, Chakrabarti A. Candidemia in a pediatric intensive care unit. Pediatr Crit Care Med 2004;5:369-74. 23.

- 18. Verma AK, Prasad KN, Singh M, Dixit AK, Ayyagari A. Candidaemia in patients of a tertiary health care hospital from North India. Indian J Med Res 2003;117:122-8.
- 19. Morrell M, Fraser VJ, Kollef MH. Delaying the empiric treatment of candida bloodstream infection until positive blood culture results are obtained: a potential risk factor for hospital mortality. Antimicrob Agents Chemother. 2005;49(9):3640–5.
- 20. Chakrabarti A. Fungal Infections in Asia: Eastern Frontier of Mycology. India: Elsevier; 2014.
- 21. Capoor MR, Nair D, Deb M, Verma PK, Srivastava L, Aggarwal P. Emergence of non-albicans Candida species and antifungal resistance in a tertiary care hospital. Japanese j infect dis. 2005;58(6):344-348.
- 22. Khadka S, Sherchand JB, Pokhrel BM, Parajuli K, Mishra SK, Sharma S, Shah N, Kattel HP, Dhital S, Khatiwada S, Parajuli N. Isolation, speciation and antifungal susceptibility testing of Candida isolates from various clinical specimens at a tertiary care hospital, Nepal. BMC res notes. 2017;10(1):1-5.
- 23. Seyoum E, Bitew A, Mihret A. Distribution of Candida albicans and non-albicans Candida species isolated in different clinical samples and their in vitro antifungal suscetibity profile in Ethiopia. BMC Infect Dis. 2020;20(1):231. doi:10.1186/s12879-020-4883-5.
- 24. Bhattacharjee P. Epidemiology and antifungal susceptibility of Candida species in a tertiary care hospital. Curr Med Mycol. 2016;2(2):20–7.
- 25. Bilal H, Hou B, Shafiq M, Chen X, Shahid MA, Zeng Y, et al. Antifungal susceptibility pattern of Candida isolated from cutaneous candidiasis patients in eastern Guangdong region: A retrospective study of the past 10 years. Front Microbiol. 2022;13:981181. doi:10.3389/fmicb.2022.981181.
- 26. Bedini A, Venturelli C, Mussini C, Guaraldi G, Codeluppi M, Borghi V, et al. Epidemiology of candidaemia and antifungal susceptibility patterns in an Italian tertiary-care hospital. Clin Microbiol Infect. 2006;12(1):75–80.
- 27. Bhattacharya S, Sae-Tia S, Fries BC. Candidiasis and Mechanisms of Antifungal Resistance. Antibiotics (Basel). 2009;9(6):312. doi:10.3390/antibiotics9060312.
- 28. SnyderGM, WrightSB. The Epidemiology and Prevention of Candida auris. Curr Infect Dis Rep. 2019;21(6):19. doi:10.1007/s11908-0190675-8.
- 29. Arastehfar A, Yazdanpanah S, Bakhtiari M, Fang W, Pan W, Mahmoudi S, et al. Epidemiology of candidemia in Shiraz, southern Iran: A prospective multicenter study (2016–2018) Get access Arrow. Med Mycol. 2016;59(5):422–30
- 30. Sridharan S, Gopalakrishnan R, Nambi PS, Kumar S, Sethuraman N, Ramasubramanian V, et al. Clinical Profile of Non-neutropenic Patients with Invasive Candidiasis: A Retrospective Study in a Tertiary Care Center. Indian J Crit Care Med. 2021;25(3):267–72.
- 31. Naseema Shaik, Uma Penmetcha, Ramesh Babu Myneni, Padmaja Yarlagadda and Susmitha Singamsetty. 2016. A study of Identification and Antifungal Susceptibility Pattern of Candida species Isolated from Various Clinical Specimens in a Tertiary care Teaching Hospital, Chinakakani, Guntur, Andhra Pradesh, South India. *Int. J. Curr. Microbiol. App. Sci.* 5(7):71-91. doi: http://dx.doi.org/10.20546/ijcmas.2016.507.006
- 32. Jyoti Pal and Dakshina Bisht/ International Journal of Biomedical and Advance Research 2016: 7(5): 226-229
- 33. Kaur R, Jaggi S, Dhakad MS, et al. An etiological and antifungal profile of candidemia in children. Int J Community Med Public Health 2019;6(9):3899–3904. DOI: 10.18203/2394-6040.ijcmph20193990.
- 34. Singh R, Parija SC. Candida parapsilosis: an emerging fungal pathogen. Indian J Med Res 2012;136(4):671–673.
- 35. Khairat SM, Sayed AM, Nabih M, Soliman NS, Hassan YM. Prevalence of Candida blood stream infections among children in tertiary care hospital: detection of species and antifungal susceptibility. Infect Drug Resist. 2019;12:2409-2416.

- 36. Rajeshwari R, Vyasam S, Chandran J, Porwal S, Ebenezer K, Thokchom M, James EJ, Karuppusami R. Risk Factors for Candida Infection among Children Admitted to a Pediatric Intensive Care Unit in a Tertiary Care Centre in Southern India. Indian J Crit Care Med. 2022;26(6):717-722.
- 37. Santolaya ME, Thompson L, Benadof D, et al. A prospective, multi center study of Candida bloodstream infections in Chile. PLoS ONE 2019;14(3):e0212924. DOI: 10.1371/journal.pone.0212924
- 38. Seyoum E, Bitew A, Mihret A. Distribution of Candida albicans and non-albicans Candida species isolated in different clinical samples and their in vitro antifungal suscetibity profile in Ethiopia. BMC Infect Dis. 2020;20(1):231. doi:10.1186/s12879-020-4883-5.
- 39. Ahmed S, Shahid M, Fatima N, Khan F, Tayyaba U. Candidemia Changing Trends from Candida albicans to Non-Albicans Candida from a Tertiary Care Center in Western UP, India. CHRISMED J Health Res. 2020;7(3):167–72.